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**MEIOFAUNAL ASSEMBLAGES
OF THE THAMES ESTUARY**

April 1989 - March 1990

Modules I - III

**A Study of the Meiofaunal Communities Present in Sediment Samples
Collected by the National Rivers Authority, Thames Region**

by

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ENVIRONMENT AGENCY



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The Meiofaunal Assemblages of the Thames Estuary

April 1989 - March 1990

Module I

EXECUTIVE SUMMARY

by

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1. The aim of the study reported here is to examine the use of the assemblages of meiofaunal organisms (sediment-dwelling species measuring less than 1 mm in length) to assess the pollution status of the Thames Estuary and the possible effects of reduced freshwater flow and mechanical disturbance. It also demonstrates the value of meiofaunal techniques as a sensitive means of longer-term monitoring of changes in conditions that occur within the Tideway. A number of parameters and methods of examining the data were used to make inferences on the environmental quality within the Estuary and these are described and discussed in detail. These include information on the tolerance of different species to pollution or other forms of environmental stress, effects on the numbers of species present in different communities (species richness), effects on the densities of individuals and on the trophic structure of the communities (the relative abundances of different feeding types). These parameters were also used to assess the effects of reduced freshwater flow and mechanical disturbance.

2. Meiofauna offer many distinct advantages in the study of environmental conditions. Being nearer to the base of food webs in aquatic ecosystems than larger invertebrate animals, meiofauna respond more rapidly to any changes in environmental conditions that affect the quality and nature of their food supply. Unlike other animals, the less mobile meiofauna are subjected continuously to the effects and constraints of any "foreign" materials that enter their environment and this is reflected in the composition of their communities.

Other advantages of using meiofauna as biological indicator organisms for assessing and monitoring environmental conditions can be summarised as follows:

- i. meiofaunal populations are inherently stable, that is, under similar environmental conditions similar meiofaunal populations will develop (similar species complements, densities, species richness and trophic structure). This enables changes in the structure of meiofaunal assemblages to be related more easily to changes in environmental conditions;

- ii. the short generation times of certain species and high diversity of species present in a given habitat, especially in the case of the Nematoda, enable communities to respond more rapidly than larger invertebrates (macrofauna) to changing conditions;

- iii. certain meiofaunal species are amongst the last to survive in grossly polluted conditions; consequently meiofaunal indices can be used to assess changes in conditions in the entire range of stressed and polluted habitats;

- iv. the high densities of meiofaunal species in a given environment make statistically valid sampling simpler than for other groups; sampling macrofauna on a comparable scale would itself have a



marked impact on the ecology of area under examination;

- v. given the equivalent effort required to produce the same resolution using macrofaunal indices, the costs of meiofaunal surveys and analyses in terms of time, effort and expenditure are relatively low. This has proved to be an important consideration where industries and authorities have had to strike a balance between competing demands on limited resources.

3. Sediment samples were collected by NRA (Thames Region) staff as part of the Thames Estuary Benthic Programme from intertidal and subtidal sites between Teddington and Shoeburyness between April 1989 and March 1990. This programme included examination of macrofauna and sediment chemistry. The sediment samples were fixed at the time of collection or shortly after with formaldehyde solution. In the laboratory, meiofaunal organisms were elutriated and concentrated using polymer density gradient solutions. Harpacticoid copepods were removed for dissection (necessary for accurate identification). Halacarid mites were also removed and were cleared in polyvinyl lactophenol. Remaining meiofaunal groups, principally Nematoda, were processed to glycerol and mounted on slides for identification and enumeration. All specimens were examined using Nomarski differential interference contrast microscopy and identification confirmed using the meiofaunal reference collections held at *Physalia*.

4. k-Dominance plots examine the number of species present in a given habitat and their relative abundance. These were used to provide information about fundamental changes that occurred in dominance:diversity characteristics of assemblages at each sampling site. In meiofaunal communities, dominance often reflects prevailing environmental conditions. Consequently, the degree of dominance and changes in this value can provide a valuable indication of ambient conditions.

The trophic structure of each community was determined and the relative abundances of selective epigrowth feeders, diatomivorous species, microbivorous species, non-selective deposit feeders (detritivores) and predators/omnivores documented. Changes in conditions are often reflected directly in the nature and quality of the food supply, especially in populations of sediment bacteria, fungi and algae. These are difficult to assess directly. However, populations of certain meiofaunal species that feed on these groups are a convenient way in which such changes can be detected and monitored.

Species-richness (numbers of species present) and densities of individuals combined with a knowledge of the biology of the species (including reproductive strategies and sensitivity to stress/disturbance), were used throughout the study to assess conditions at the stations sampled.

The results of the surveys are summarised briefly below.



5. Amongst the principal meiofaunal groups, a total of 207 nematode species, 49 harpacticoid copepod species and 6 acarine species were observed during the course of the survey. Species from 10 further meiofaunal phyla were also recorded along with the larval and juvenile stages of several macrofaunal species (see Module IV).
6. Principal *natural* factors governing the structure of species assemblages were salinity, sediment granulometry, position relative to intertidal zone and season (see Module IV).

- i. Salinity

The estuary supported marine, estuarine and freshwater meiofauna and encompassed species capable of tolerating broad changes in salinity (euryhaline species) through to those that are restricted to narrow salinity ranges (stenohaline species). Euryhaline species were ubiquitous and predominated at the mid-estuary sampling stations. The distributions of these species formed overlapping, successional series along the Tideway. The stenohaline species were located at either end of the estuary and formed characteristic assemblages of freshwater or marine species.

- ii. Sediment Granulometry

Sediment types ranged from coarse grained sands that contained stones and shell fragments to compacted clay muds and flocculant, high silt-clay fraction sediments. In general, the coarse sediments supported a high diversity of meiofaunal species (up to 98 taxa; Chapman Buoy) at low densities. In these habitats dominance was usually low. The mud communities were less species-rich than those of neighbouring sands but were capable of supporting exceptionally high meiofaunal densities (greater than 153,000 individuals litre⁻¹ sediment; Southend intertidal site). Dominance was often high in these sediments. Amongst the nematodes, selective epigrowth feeding species and diatomivorous species predominated in the coarser substrates which also favoured interstitial harpacticoid copepods. In contrast, non-selective deposit feeding nematodes and epibenthic harpacticoid copepods predominated in the Thames muds.

- iii. Proximity to Intertidal Zone.

With few exceptions, the intertidal sites sampled in the Thames supported higher densities and numbers of meiofaunal species than equivalent subtidal sites. The reasons for this phenomenon are discussed.



iv. Season

Successions of species were found to occur at given sampling sites at different times of the year. This was largely related to the trophic mode of the species and is thought to be dependent on ambient water temperatures, availability of food and competition pressure. Within different groups, opportunist species (r-strategists) with high natural rates of increase and K-strategists with longer life-cycles could be recognised.

7. Against this background of *natural* variation, the effects of external, anthropogenic stresses and perturbations were documented. These stresses included diffuse, non-specific pollution, discharge of sewage effluents, reduced freshwater flows resulting from high levels of water abstraction during dry periods and mechanical disturbance.

i. Non-specific Pollution

With reference to the status of meiofaunal assemblages at neighbouring sampling stations, attention is drawn to depressed meiofaunal densities and diversities of species at certain estuarine sites. Possible causes are considered and potential changes in the meiofaunal assemblages resulting from worsening or improving conditions postulated with a view to aiding monitoring of the status of the Tideway (see Module II).

ii. Sewage Effluents

Two different effects of sewage effluents were noted that related to the position of the treatment works within the estuary. At Beckton and Crossness sewage works meiofaunal densities and species richness were depressed around the outfalls in comparison to neighbouring sampling stations. Tolerant, euryhaline non-selective detritivores persisted along with low salinity microbivorous species in species-poor communities at sites at which macrofauna was either absent or impoverished. At Southend, salinity effects of the sewage effluent appeared to be secondary as marine species were present throughout the year. Dominance, however, was high for a sandy sediment (up to 40%) exceeding that of the muddier intertidal sediments. Examination of the species complements revealed the dominant species to be a non-selective deposit feeding nematode commonly found in association with elevated levels of organic material. Densities of this species may provide a useful indicator of the effects of this outfall in the higher salinity outer reaches of the estuary (see Module II).

iii. Reduced Freshwater Flow

Direct effects of decreased freshwater flows were the reduction in densities of stenohaline freshwater species at the head of the estuary (Teddington and Kew), the extended colonisation of upstream stations by euryhaline mid-estuary species and the ingress of marine species into the outer estuary. Increased



sedimentation leading to changes in sediment types from coarse grained to finer substrates along the Tideway may have been an indirect effect of reduced flow that produced associated changes in meiofaunal communities outlined in 4 ii above (see Module III).

iv. Mechanical Disturbance

Some evidence of mechanically disturbed or unstable sediments was derived from meiofaunal indices. These were based on the ability of the more mobile epibenthic harpacticoid copepods to recolonise disturbed sediments more rapidly than nematodes. The value of this index is considered in terms of monitoring conditions in the Thames Estuary (see Module III).

8. The adoption of a routine meiofaunal sampling programme would greatly enhance the sensitivity of existing faunal surveys in the establishment and monitoring of the pollution status of the Thames Estuary. Comparatively small improvements in prevailing conditions in certain reaches could be documented and attention drawn at an early stage to the onset of adverse conditions. The effects of reduced freshwater flow and mechanical disturbance could also be assessed. Multivariate techniques for the analysis of complex meiofaunal communities in the existing data sets are suggested as a means of defining key species assemblages at each of the sampling stations at different times in the year. This would facilitate the detection of changes in benthic communities in response to altered environmental conditions (see Modules II and III). The multivariate analyses could be integrated with those of the macrofaunal surveys and correlated with the physical and chemical parameters of the sediments. This would greatly improve our understanding of the factors that regulate populations of meio- and macrofaunal species and could strengthen existing pollution management strategies for the Thames Estuary.



The Meiofaunal Assemblages of the Thames Estuary

April 1989 - March 1990

Module II

**An Assessment of the Pollution Status of the Thames Estuary
as Revealed by its Meiofaunal Assemblages**

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1. Summary

Examination of the meiofauna indicated that sites sampled at the Teddington and seaward ends of the Thames Estuary supported comparatively healthy assemblages and densities of infaunal species. The communities at these sites were characterised by the presence of several pollution-intolerant invertebrate species. At the freshwater end of the estuary, these included rhizopod amoebae, microbivorous nematodes, tardigrades, halacarid and hydracarine mites and freshwater harpacticoid copepods. At the outer estuarine sites, clean, unstressed conditions were indicated by the presence of marine ciliates, gastrotrichs, kinorhynch, oxystominid and desmoscolecid nematodes and interstitial harpacticoid copepods.

Meiofaunal dominance and diversity proved to be useful indicators of stressed conditions within the Estuary. Depressed species-richness and increased dominance were apparent at certain sites in the mid-estuary that were in close proximity to the two sewage treatment works (Beckton and Crossness). Here the subtidal sites appeared to be most strongly affected, with reduced populations of selective epigrowth and diatom feeding species (nematodes and copepods). Amongst the nematodes, non-selective, euryhaline detritivores predominated and secernentean species, more usually associated with freshwater habitats, were also noted.

Reduced densities of meiofauna also appeared to correlate with stressed conditions. At the subtidal sites at Southend and Sea Reach No. 2 Buoy, for example, densities of harpacticoid copepods and nematodes were lower than at surrounding stations. These comparatively species rich communities comprised a modified assemblage of species and were dominated throughout the year by a single nematode species. The latter was present at low densities only at other sampling sites. It is believed that the modified assemblages and reduced densities might relate to the Southend sewage outfall.

Nematode:copepod ratios were not thought to be useful indices of pollution status owing to the wide variations in densities of both meiofaunal groups in response to natural factors such as granulometry and salinity.



2. The Value of Meiofauna in the Determination of Pollution Status

Meiofauna are sediment-dwelling species that measure less than 1 mm. They are of fundamental importance in aquatic ecosystems, forming food for macrofaunal invertebrates as well as fish and possibly certain avifauna (e.g. waders). They also offer many distinct advantages in the study of prevailing environmental conditions. Essentially, being nearer to the base of food webs in aquatic ecosystems they respond rapidly to any changes in environmental conditions that affect the quality and nature of their food supply. Unlike other animals, the less mobile meiofauna are subjected continuously to the effects and constraints of any "foreign" materials that enter their environment and this is reflected in the composition of their communities.

The advantages of using meiofauna as biological indicator organisms in monitoring environmental conditions can be summarised as follows:

1. the existence of inherently stable populations enables changes in the structure of meiofaunal assemblages to be related more easily to changes in environmental conditions;
2. short generation times and high species diversity, especially in the Nematoda, enable communities to respond more rapidly than macrofauna to changing conditions;
3. certain meiofaunal species are amongst the last to survive in grossly polluted conditions; consequently meiofaunal indices can be used to assess the entire range of polluted conditions;
4. the high densities of certain meiofaunal species in a given environment make statistically valid sampling simpler than for other groups; sampling macrofauna on a comparable scale would itself have a marked impact on benthic communities;
5. given the equivalent effort required to produce the same resolution of using macrofaunal indices, the costs of meiofaunal surveys and analyses in terms of time, effort and expenditure are relatively low. This has proved to be an important consideration where industries and authorities have had to strike a balance between competing demands on limited resources.

Meiofaunal studies have gained acceptance throughout the world for the delineation of impact zones of pipe-borne effluents (Newell *et al.*, 1990a and b). *Physalia* undertakes annual examination of meiofaunal assemblages associated with industrial outfalls in Britain (Humber and Tees Estuaries), France, Holland, Spain, Italy, Canada (St. Lawrence River), Malaysia, Southern Africa and Tasmania. These surveys form a key part of the environmental audit for the industries concerned and enables them to assess the efficacy of their recovery and treatment processes. Other meiofaunal projects have included the accurate delineation of the impact zones of marine sewage outfalls, where macrofaunal studies had failed to reveal effects (Irish Sea), the detection of impacts of fish farm effluents, the assessment of the effects of drainage from mining operations and the evaluation of the effects of constructional activity on aquatic



ecosystems.

Until now, there have been almost no studies or published information on the meiofaunal assemblages of the Thames Estuary. Feil (1989) includes some observations on meiofaunal communities at the mouth of the estuary in relation to the biology of O-group Dover sole, *Solea solea* and Trett, Feil and Forster (unpublished) have examined meiofaunal communities of a limited number of sites as part of an engineering consultancy for the location of a thermo-electric power station.

This section of the report considers the pollution status of the Thames Estuary in the light of the findings outlined in Module IV.



3. Interpretation of Meiofaunal Results

i. General Comments

As in other estuarine systems, the meiofaunal communities were dominated by nematodes. However, the harpacticoid copepods that are usually the main sub-dominant meiofaunal group were poorly represented, especially in the middle reaches of the Estuary. The reason for this is unknown but it might reflect general pollution stress. Many of the larger epibenthic harpacticoid copepods are episammic species that browse epigrowth from sediment particles. Consequently, any factor that affects the microbiology of sediments may influence the populations of these species.

Abundance of individuals, species richness and dominance are inter-related characteristics of meiofaunal communities that can provide important information concerning prevailing environmental conditions. These factors, along with the trophic structure of the meiobenthic assemblages, are discussed in connection with the pollution status of the Estuary in the following sections.

ii. Species Richness and Abundance

In general, species richness at a given salinity is highest in coarse grained sediments and lowest in high silt-clay fraction muds. This may relate to the heterogeneity of habitats present in coarse sediments that can be exploited by interstitial fauna. The converse situation is true of total densities of meiofauna; muds can support considerably higher densities of meiofauna than sands.

Examination of the results of the 4 meiofaunal surveys, presented in Module IV, suggests that both the Teddington and seaward ends of the Estuary support relatively healthy assemblages and densities of the smaller aquatic invertebrates. In comparison to marine and estuarine systems, there is little published information on the meiofauna of freshwater sediments. However, surveys that we have undertaken of lake and river meiofauna throughout the world have established that unpolluted freshwater sediments usually sustain moderate densities of a comparatively small number of meiofaunal species. The freshwater sediments sampled between Teddington to Cadogan Pier supported a high diversity of predominantly freshwater meiofauna. Many of these species are pollution intolerant. These groups included rhizopod amoebae, such as *Centropyxis* and *Euglypha* species, microbivorous nematodes (type 1A species) including species of *Rhabditis* and *Alaimus*, hydracarine mites and freshwater harpacticoids, such as *Bryocamptus* and *Moraria* species. Consequently, pollution stress on these sediment assemblages would appear to be minimal. However, evidence for changes in their communities with respect to altered freshwater flow was noted and is considered in detail in Module III.

At the seaward end of the estuary, the coarser grained, high salinity sediment assemblages are exceptionally species rich comprising, for example, up to 98 principal meiofaunal species (i.e. species of Nematoda, harpacticoid Copepoda and Acari). This is all the more notable given the high densities of meiofauna recorded; in the January - March 1990 survey, 153,403 nematodes litre⁻¹ sediment were observed at the intertidal site at Southend (19i) and 125,012 nematodes litre⁻¹ sediment at Shoeburyness East (21).



Again, 'pollution-sensitive' species and groups were present at the outermost stations. In this particular habitat these species included representatives of marine ciliates, kinorhynchs, gastrotrichs, oxystominid and desmoscolecoid nematodes and interstitial harpacticoid species. With certain possible exceptions (see below), there is little evidence to suggest that these communities are pollution stressed or impacted.

In most of the estuaries that have been studied, meiofaunal diversity and densities rise with increasing mean salinity ranges (see Platt and Warwick, 1980). In practice this means that species numbers and abundances increase progressively towards the mouth of an estuary. The numbers of nematode and harpacticoid species observed at each sampling site in the Thames throughout the study are illustrated in Figures 1 to 4. Whilst there is a general trend for increased species richness towards the mouth of the estuary, sites are present at which diversity is lower than in neighbouring, upstream assemblages. These centre around the mid-tideway, although other examples can be seen at certain stations in the outer estuary. In several cases sites with reduced numbers of species exhibited reduced densities of meiofauna (Figures 5 to 8). However, in one or two instances this could also be explained by scouring effects of currents and changes in sediment types (see Module III).

In the mid-estuary, sampling stations between Woolwich (8) and Purfleet (11) supported depauperate meiofaunal communities throughout the survey. This correlated with the location and potential sphere of influence of the Beckton (9) and Crossness (10) sewage treatment works. Examination of the species complements present at each of the stations highlights the fact that the subtidal sites are most severely affected. In each case, the dominance and trophic structure of the communities are modified (see below). Euryhaline species, such as *Sabatieria punctata* and species of *Daptonema*, were the most resilient of those present and were, on occasion, the only species to be observed at these stations. Both groups are euryhaline and readily tolerate alternating periods of reduced and high salinity and, consequently, may be pre-adapted to survive in the mobile, low salinity effluent plumes of the treatment works. Secernentean nematode species, commonly seen in the freshwater samples at the Teddington end of the estuary, were also common components of the meiofauna in this reach of the Thames although present at low densities. These were represented by microbivorous species such as *Mononchoides striatus* and members of the family Diplogasteridae. Where species richness rose during the year, this usually related to the appearance of a few individuals of marine nematode species. Copepod densities and numbers of species were low at each of the stations in this region with the exception of Purfleet (11) where estuarine, epibenthic detritivorous species were noted.

Densities and diversity of meiofauna were routinely lower at Grain Flats (20) than at its neighbouring stations (see Figures 1 to 8). Again dominance was also affected (see below). Taken together, these factors indicate a stressed community. The reasons for this stress are not entirely obvious. Direct influence of effluent from the Southend outfall is not seen as a likely explanation and, although *Richtersia inaequalis* was present at this site in three of the surveys, it was never dominant. The low similarities between the species assemblages and the reduced numbers of harpacticoid species (3 species identified over the year; see Module IV) suggests that conditions at this site were not stable. A progressive change in sediment type from a sand (April - June) to a mud (October - December) before returning to a muddy sand in the final survey was noted. It is possible that this southern shore station is affected by the backed up outflow of the Medway Estuary (see Module IV; Figure 1). Lower salinity water and allochthonous



organic material from the Medway might account for the dominance of euryhaline, non-selective detritivore nematodes at Grain Flats (*Daptonema tenuispiculum* and *Sabatieria punctata*). However, the toxic effects of domestic and industrial material originating from the Medway cannot be ruled out. Whatever the cause, the changes in community structure are clearly reflected in the k-dominance plots (see below).

A further area of note in terms of modified species richness and abundance was the South Bank Centre (5). In each of the surveys, sediments at this station supported fewer species and lower densities of meiofauna than at either of the neighbouring stations (Cadogan Pier (4) and London Bridge (6); see Module IV; Appendix I, Section 2). The variability of the species complements at this station suggests that the sediment or prevailing environmental conditions may have been unstable but without detailed knowledge of the locality of the habitat sampled it is impossible to speculate further. The sediment type varied slightly from a coarse-fine sand mix to a sand with low silt content. This alone would not account for the reduced densities and numbers of species observed and indicates the operation of other factors that might include localised pollution.

Throughout the surveys it was noted that halacarid mite densities (and possibly diversity) were lower than observed in similar estuarine systems. In the Humber, *Thalassarachna baltica* and *Copidognathus* species exhibit high densities even around industrial and sewage outfalls where they may thrive in impoverished communities as a result of reduced competition and/or reduced predation (Newell *et al.*, 1989a). It is not impossible that improved conditions within the Thames would allow such species to breed here if introduced, their absence reflecting zoological history rather than current pollution status.

iii. Dominance

In meiofaunal assemblages, knowledge of dominance *per se* is of little value in determining the pollution status of a sampling station. Dominance varies naturally from site to site and tends, for example, to be highest in fine sediments and lowest in sands. Information on the biology of the dominant species and the numbers of species present in the assemblage can provide a better insight into prevailing conditions. Further, changes in dominance with time can indicate modification of communities in response to an environmental stress. This is well illustrated at several sites in the Thames Estuary where increased dominance in sediments of a given type and at similar salinities are accompanied by reductions in species richness. k-Dominance curves are a convenient way in which to express and assess these changes over time. For example, in comparison with other sites, dominance amongst the nematode populations was elevated during the first 3 surveys at Grain Flats (20; see above and Figures 9 to 12).

At the mouth of the Estuary, 2 subtidal sampling sites, Southend (19s) and Sea Reach No. 2 Buoy (22), whilst possessing seemingly high diversities of nematode species, supported lower numbers of species and densities of both nematodes and harpacticoid copepods than surrounding stations. Again, examination of their species complements shows them to have possessed modified assemblages which might relate to the Southend sewage outfall. At both stations the dominant nematode *throughout the year* was the selachinematid *Richtersia inaequalis*. This species accounted for between 26 and 45% of the nematode population at Southend and between 33 and 64% at Sea



Reach No. 2 Buoy. In both cases dominance was highest during the 2nd. survey (July - September 1989). *R. inaequalis* is a non-selective deposit feeder (type 1B species; see Module IV, Appendix II) and is usually present at nominal densities in subtidal marine sediments often occurring in meiofaunal assemblages at the mouths of British estuaries (Trett, pers. obsvn.). Circumstantial evidence indicates that this elevated dominance and modified species assemblage relates to the impact of discharged sewage at these sites; sediments from both stations at different times of the year yielded high densities of fine, optically active fibres¹. Paper fibres commonly arise from pulped toilet papers, although other sources include paper mill effluents. Similar densities of paper fibres were not found at other sites in the Estuary. It is of interest to note that *R. inaequalis* was a recognised as a key species in multivariate analyses of meiofaunal assemblages in a study undertaken by Trett *et al.* (1990) of the benthos associated with the Fylde Coast sewage outfalls. In this instance it was dominant in stable muds. In the present survey, however, both Southend and Sea Reach No. 2 Buoy were characterised by muddy sands.

The modified nature of the meiofaunal communities observed at Sea Reach No. 2 Buoy is graphically illustrated by the k-dominance plots for this station (see Module VI, Figure 15). Dominance in sandy sediments is usually low. In the 2nd., 3rd. and 4th. surveys the dominance of *R. inaequalis* was exceptionally high correlating with its highest densities and a decline to less than half the total number of species observed at this station during the 1st. survey. Possible explanations include a concentration effect of effluent during the drier periods experienced between July 1989 and early 1990.

iv. Trophic Structure of Communities

Amongst meiofaunal assemblages, pollution stress is commonly reflected in a change in the trophic structure of the community. In the present study, specialist feeding types, such as selective epigrowth and diatom feeding nematodes (type 2A species; see Module IV, Appendix I, Section 5), were poorly represented in the more modified nematode assemblages. Non-selective deposit feeding species (type 1B species) predominated along with certain, lower salinity microbivorous nematodes. At Beckton (9) and Crossness (10), for example, the non-selective detritivores *Sabatieria punctata* and species of *Daptonema* were usually the dominant species and, on occasion, the only species observed in these assemblages. It was also apparent that relatively few type 1A species were present throughout the Estuary (see Module IV, Appendix I, Sections 1 and 2). These species are thought to be microbivorous or selective deposit feeders and each has a narrow, unarmed buccal cavity (Module IV, Appendix I). Evidence from studies of point source industrial discharges suggests that the populations of these species may be sensitive to changes in sediment microbiology (Trett and Forster, pers. obsvns.). Densities of microbivorous species belonging to the families Desmoscolecidae, Trefusiidae, Leptolaimidae and Oxystominidae were all lower in the Thames Estuary than those observed in other estuaries, including the Humber and Tees (Trett in Newell *et al.*, 1989a, 1989b; see also Warwick, 1971). Whether this reflects adverse changes in sediment microbial populations is uncertain. Similar factors may account for the reduced densities of interstitial ameirid and laophontid harpacticoid copepod species in

¹ The lignified structure of paper fibres from toilet papers or paper mills rotates the plane of transmitted light and between the crossed polar filters that form part of the Nomarski differential interference contrast optics produce bright coloured images in and amongst the meiofauna. Colour photomicrographs of this phenomenon can be supplied if required.



the mid- and outer estuarine sediments.

v. Nematode:Copepod Ratios

Nematode:copepod ratios (Raffaelli and Mason, 1981) have not been used as indicators of pollution status as doubt has been cast on their value in the detection of polluted sediments (Lamshead, 1984). However, they may be of some value in the detection of mechanically disturbed sediments (Trett and Feil, pers. obsvn.) and this is discussed in the Module III.

4. Further Analyses of Data

Multivariate analyses of the data sets for each of the sampling stations would enable characteristic ("key") meiofaunal species groupings to be identified within the Estuary. These could typify stressed or unstressed, mud or sand meiobenthic communities in the upper, mid- and outer estuary depending on the species present and their relative abundances. Similar studies using meiofaunal data have been used to assess the pollution status of subtidal sediments in relation to sewage and industrial outfalls (see Trett *et al.*, 1990; Newell *et al.*, 1989a, b, 1990a, b). Combined with macrofaunal data, this would provide a sensitive tool with which to detect and monitor changes in the Thames Estuary. Clustering techniques could also be used to examine correlations between physical and chemical parameters and the faunal complements present at each site. For several industries, this has proved to be an important means of documenting improvements in their effluent quality. Accurate detection and delineation of impact zones using multivariate techniques to identify groupings of sampling stations based solely on their meiofaunal assemblages is of central importance in such studies. In the Thames Estuary, the affinities of meiobenthic communities can be compared with those of preceding years to assess changes in the quality of the Tideway.

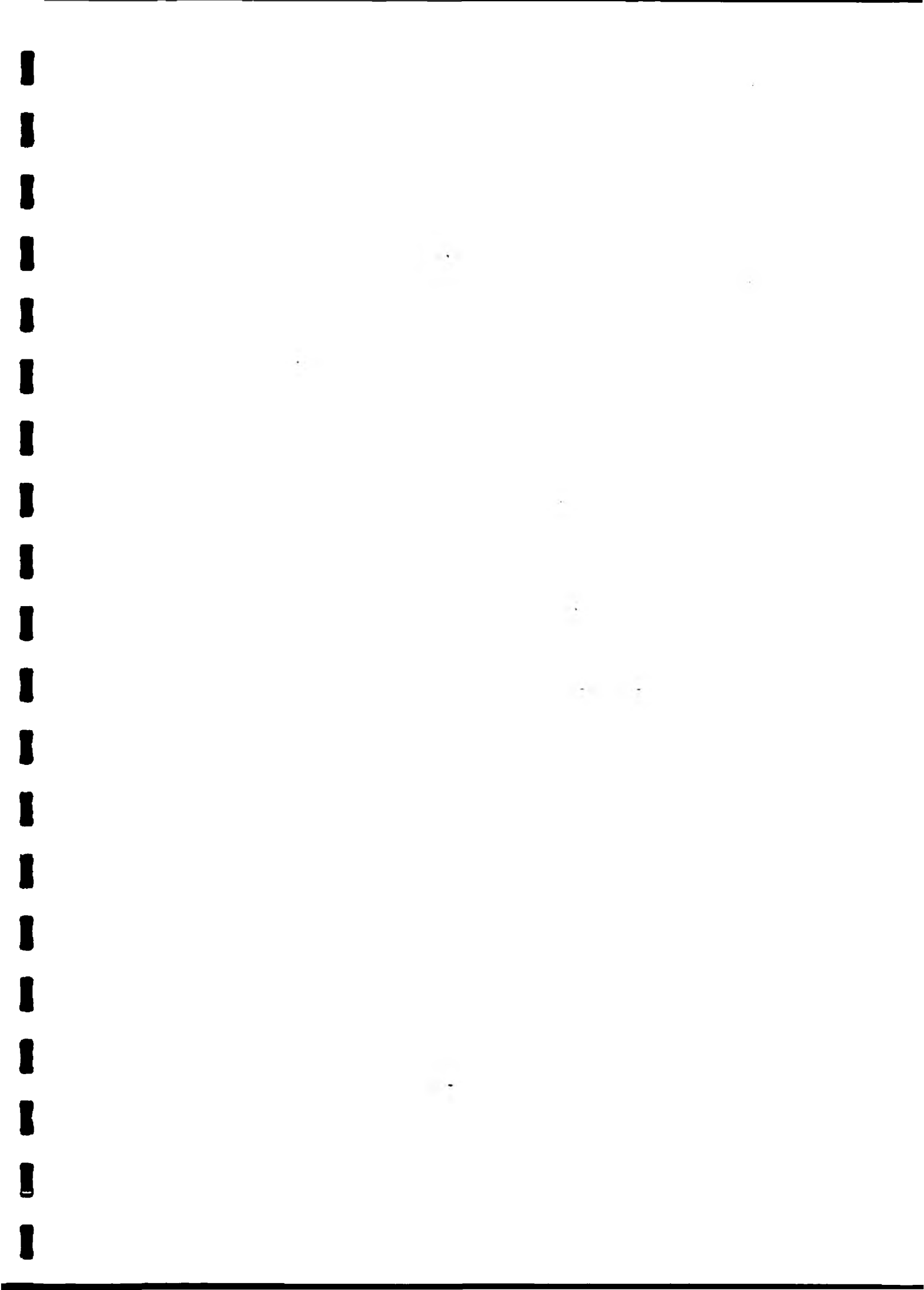
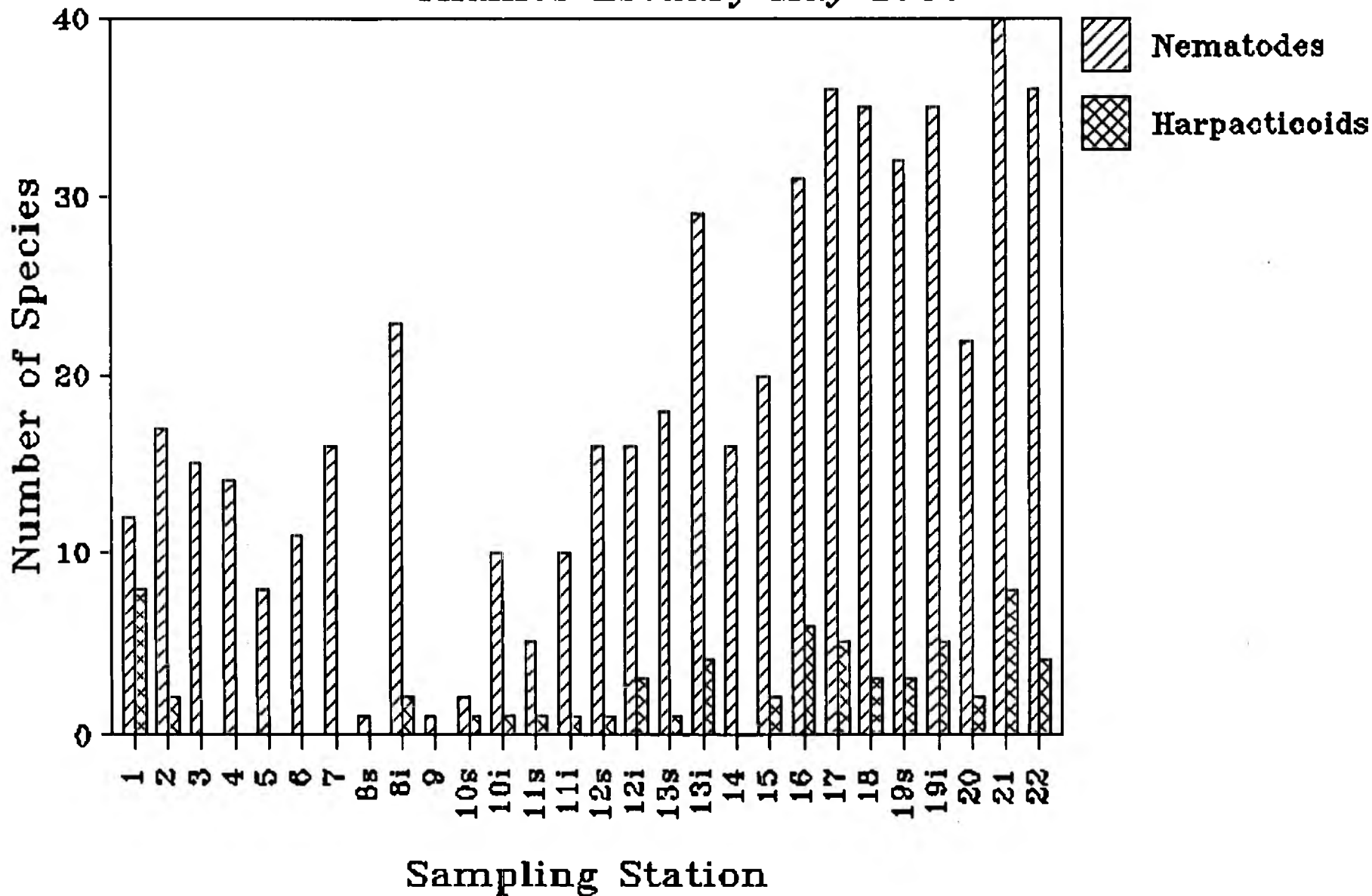


Figure 1. Total numbers of nematode and harpacticoid copepod species observed in the Thames Estuary, April - June 1989.

Nematode & Harpacticoid Copepod Species

Thames Estuary May 1989



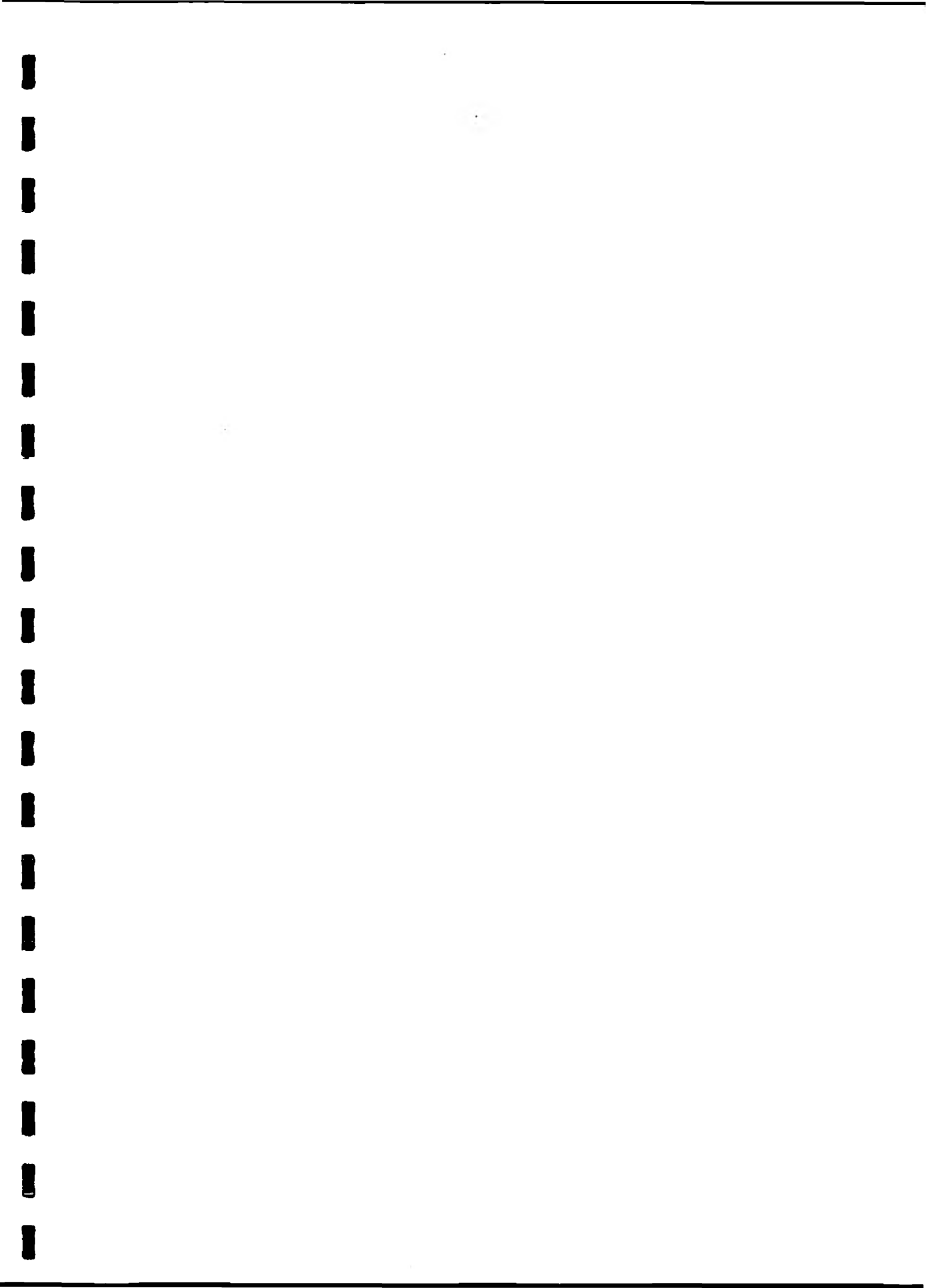


Figure 2. Total numbers of nematode and harpacticoid copepod species observed in the Thames Estuary, July - September 1989.

Nematode & Harpacticoid Copepod Species

Thames Estuary August 1989

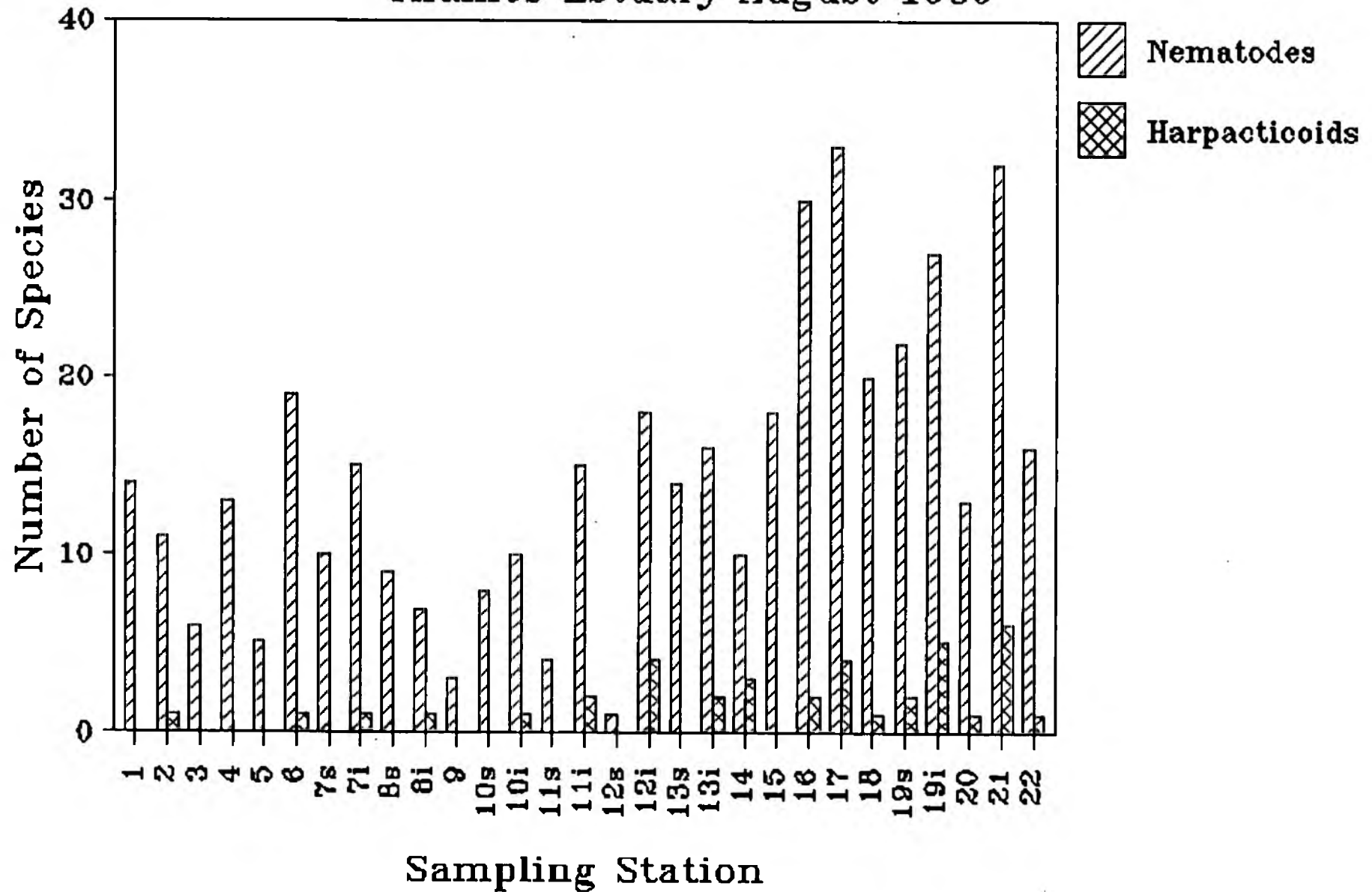
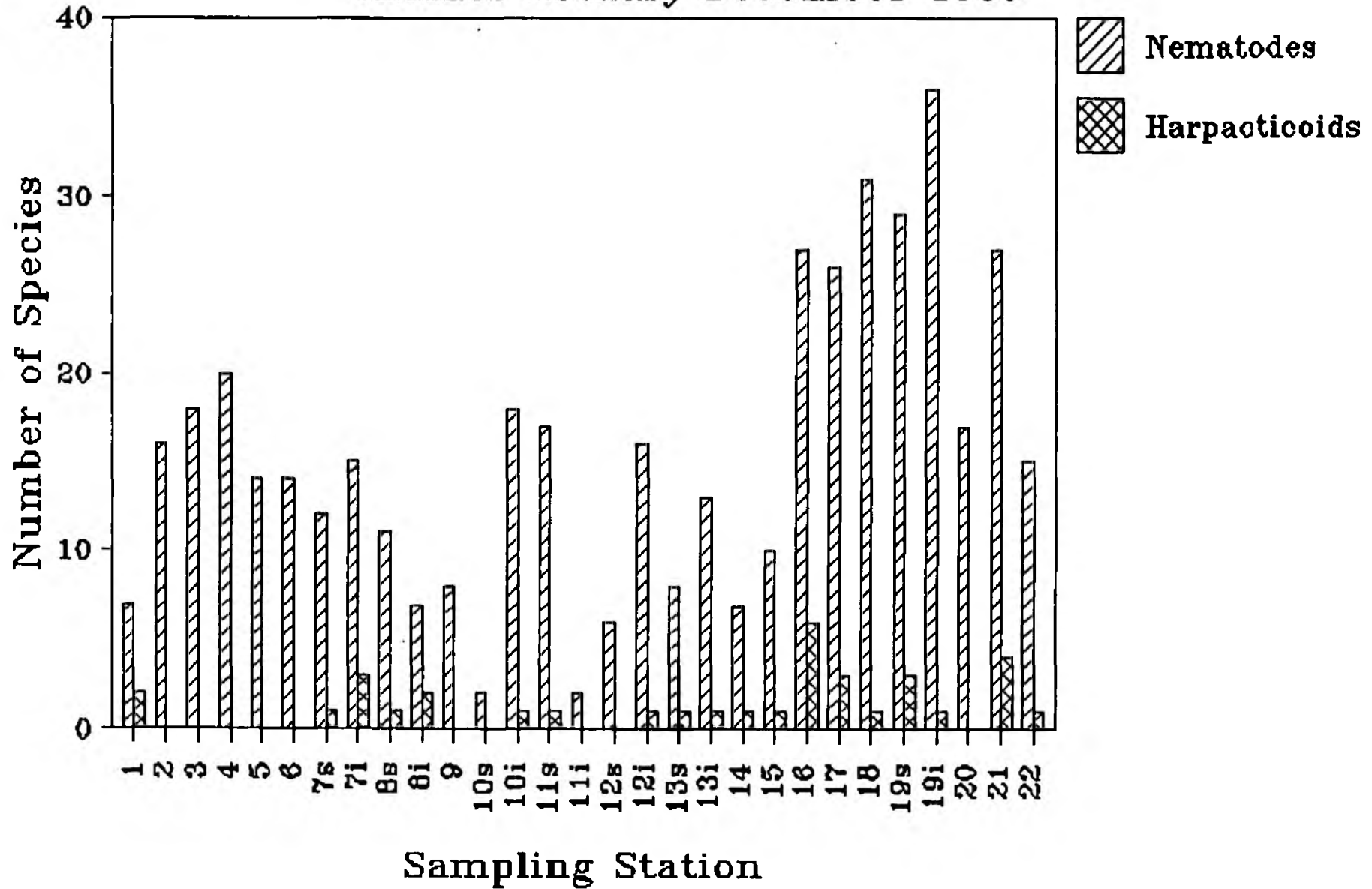




Figure 3. Total numbers of nematode and harpacticoid copepod species observed in the Thames Estuary, October - December 1989.

Nematode & Harpacticoid Copepod Species

Thames Estuary December 1989



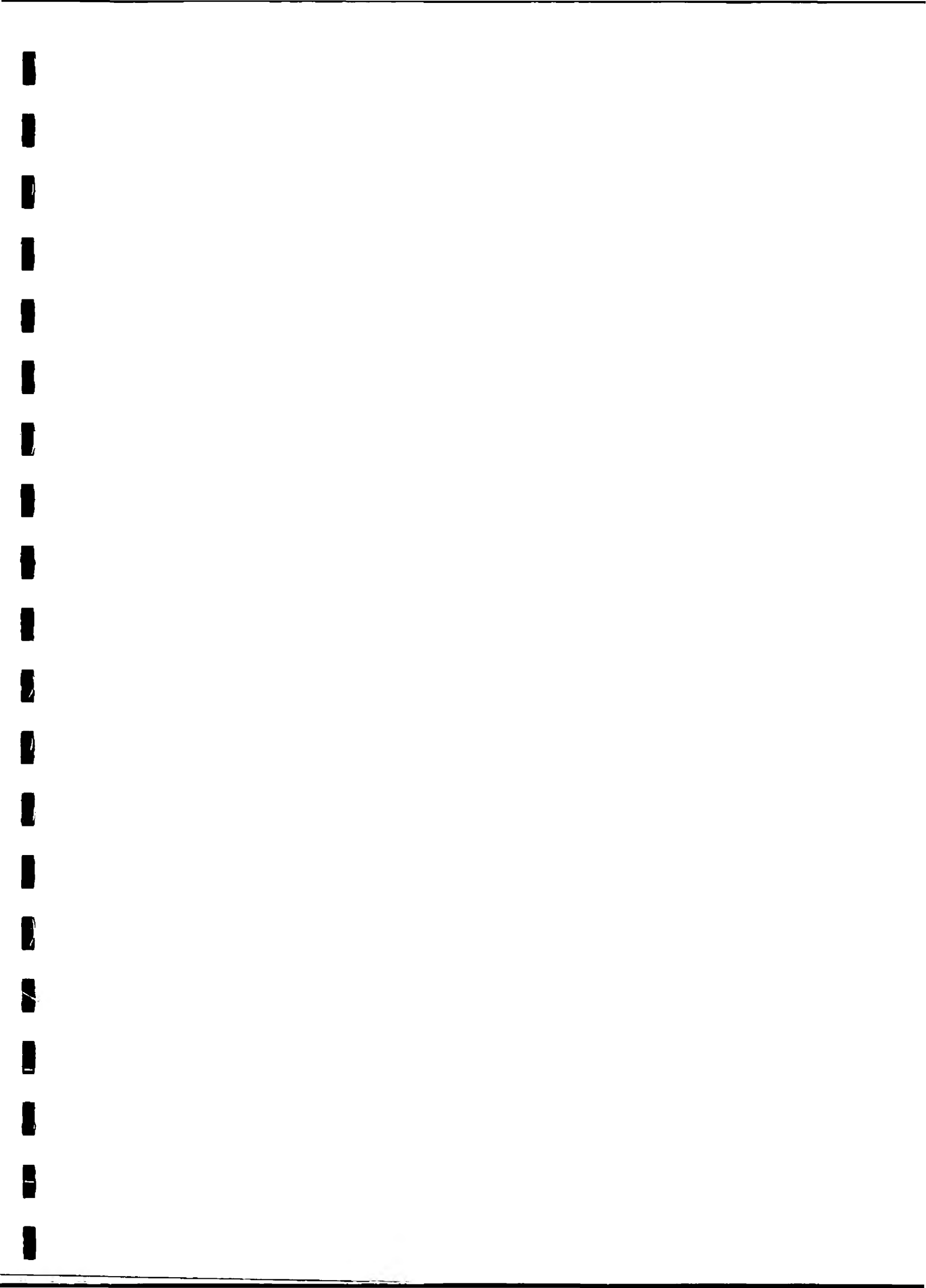
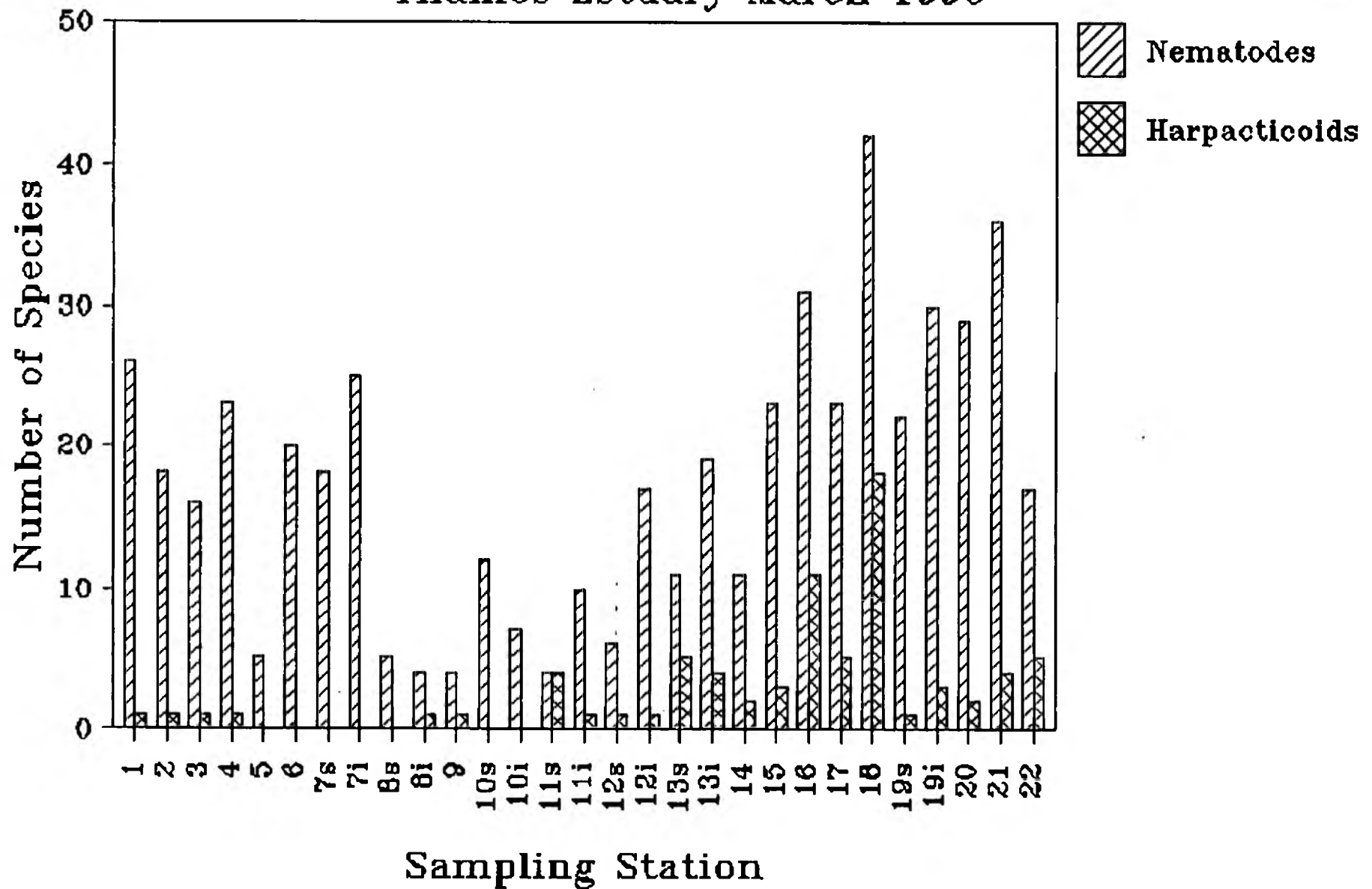


Figure 4. Total numbers of nematode and harpacticoid copepod species observed in the Thames Estuary, January - March 1990.

Nematode & Harpacticoid Copepod Species

Thames Estuary March 1990



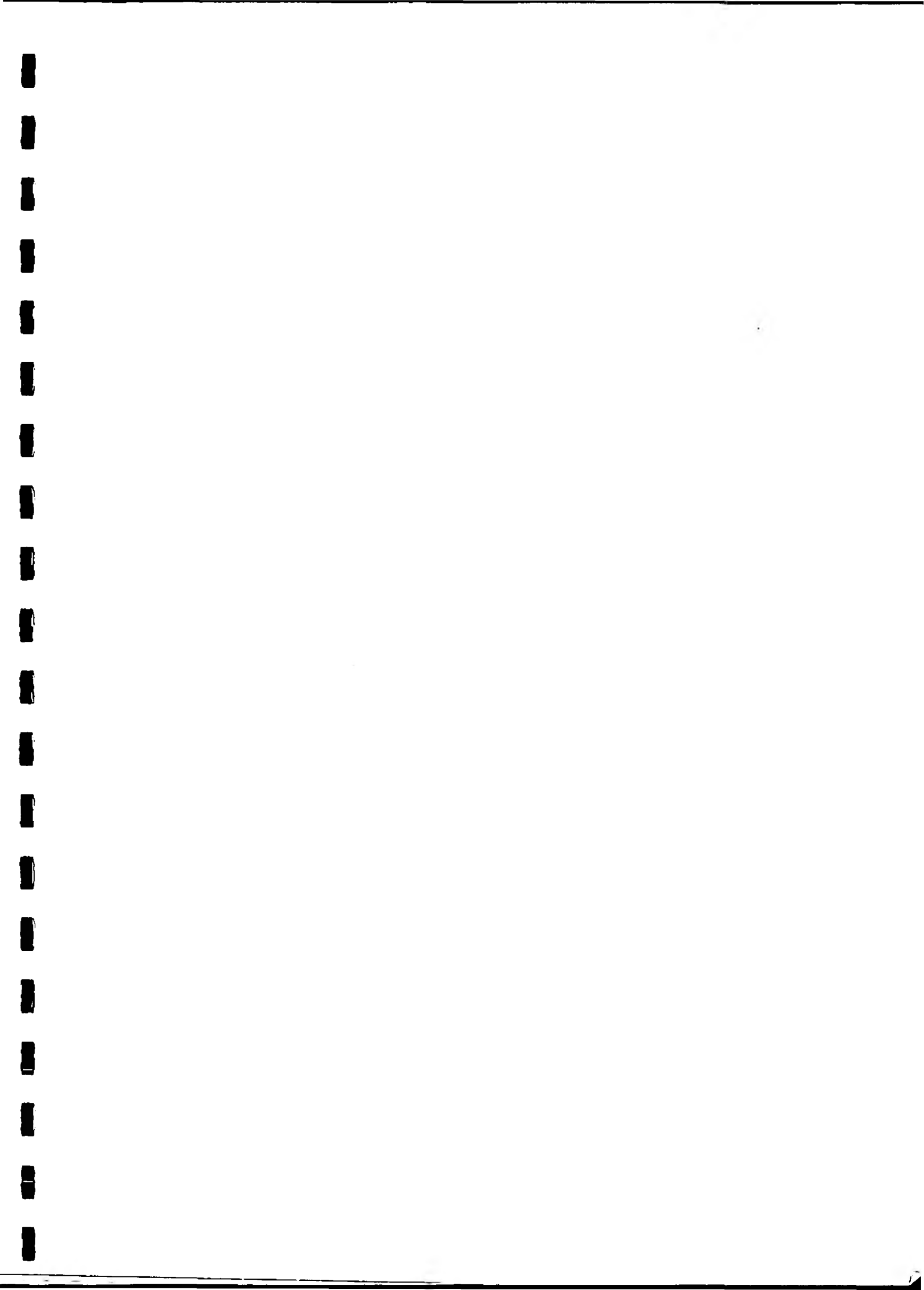
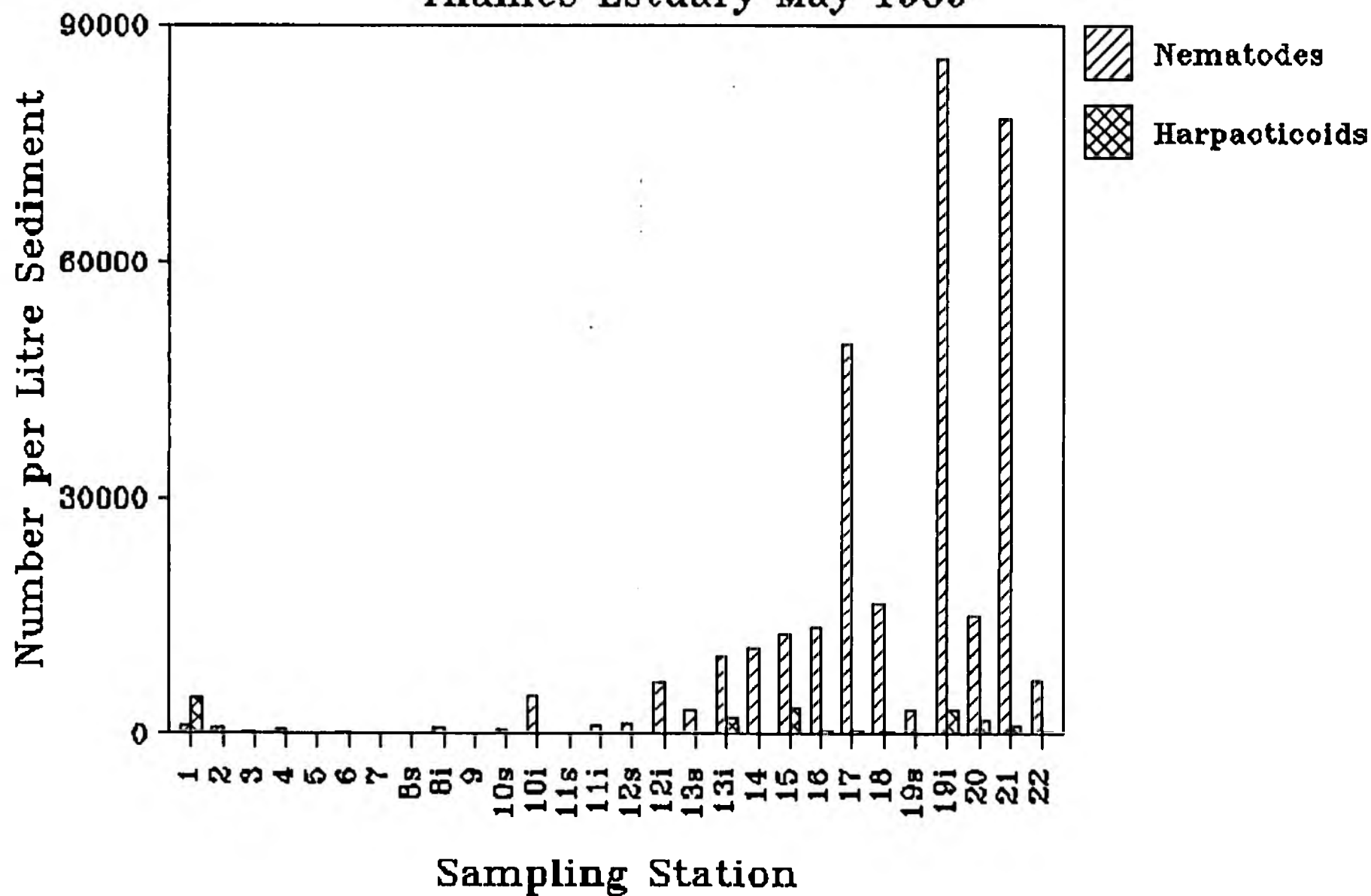


Figure 5. Total densities of nematode and harpacticoid copepod species observed in the Thames Estuary, April - June 1989.

Nematode and Copepod Densities

Thames Estuary May 1989



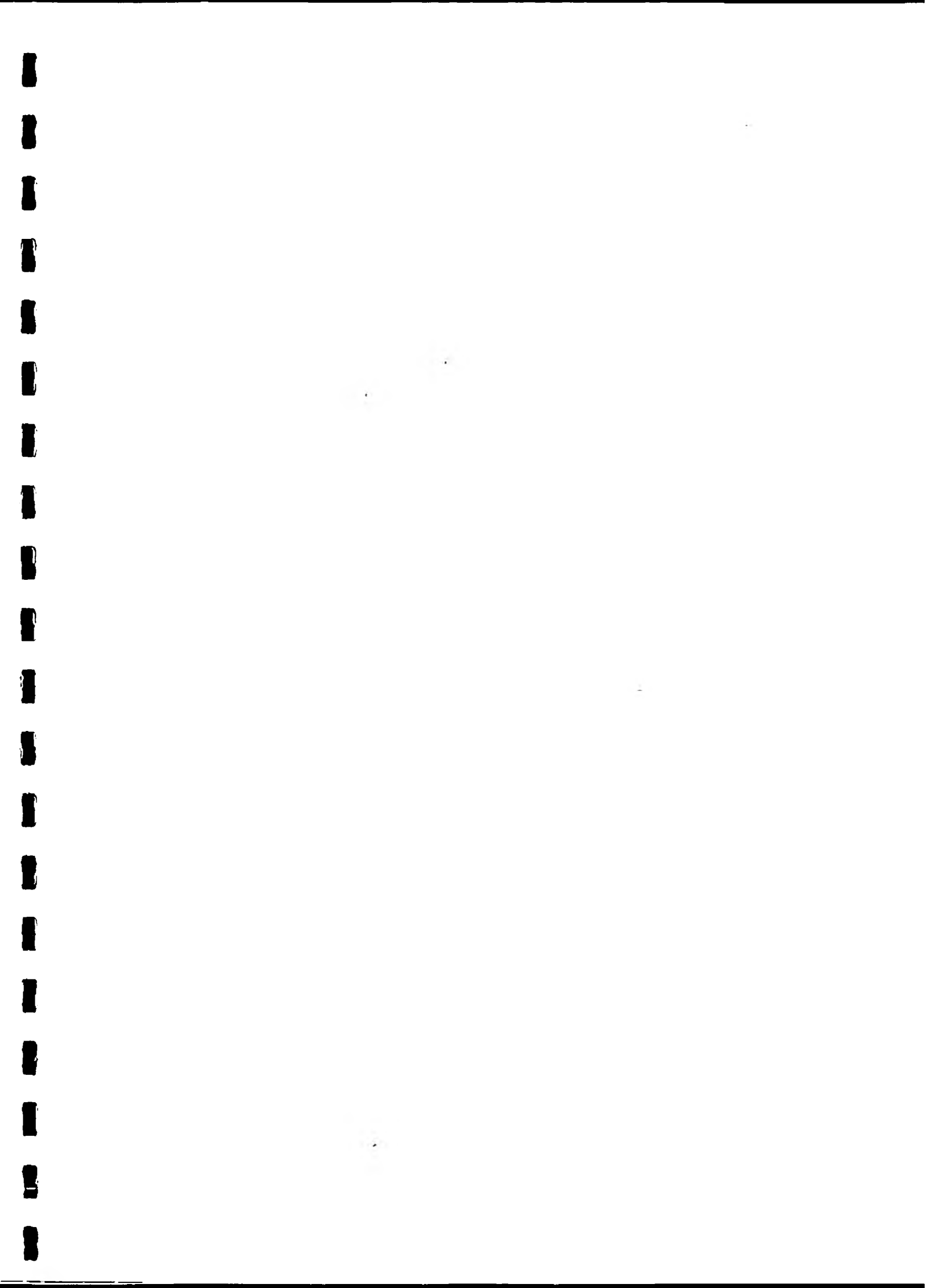
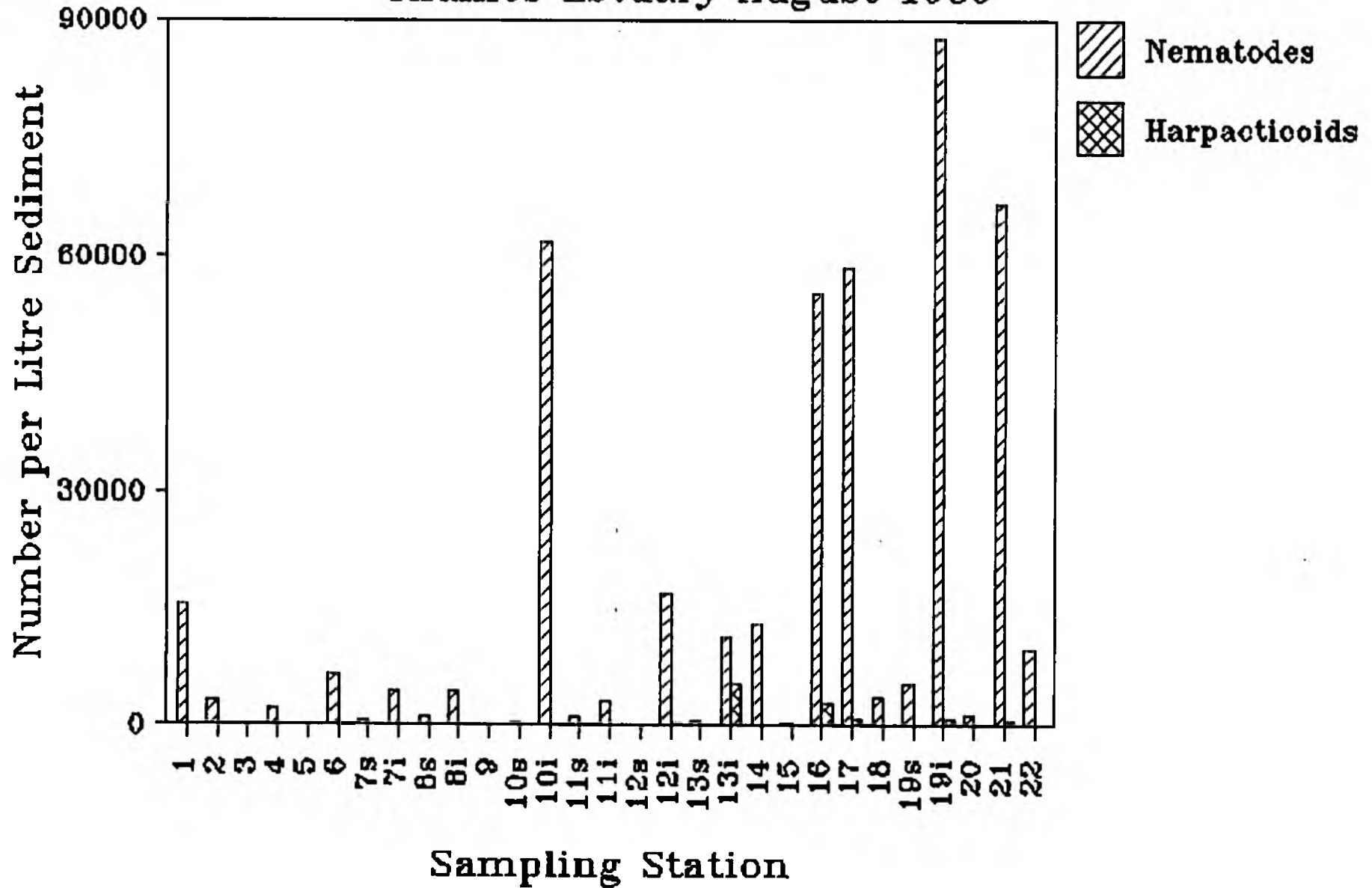


Figure 6. Total densities of nematode and harpacticoid copepod species observed in the Thames Estuary, July - September 1989.

Nematode and Copepod Densities

Thames Estuary August 1989



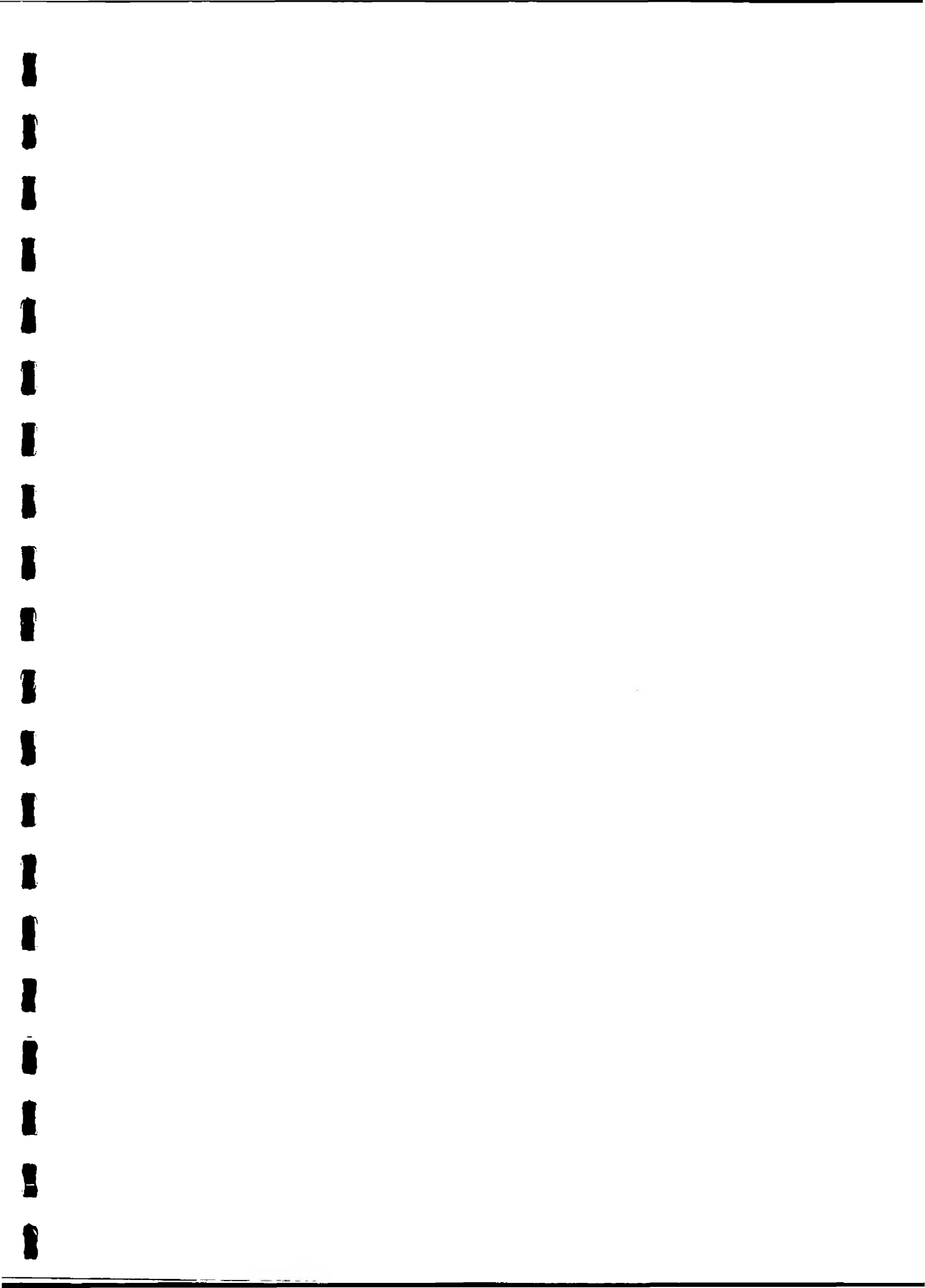
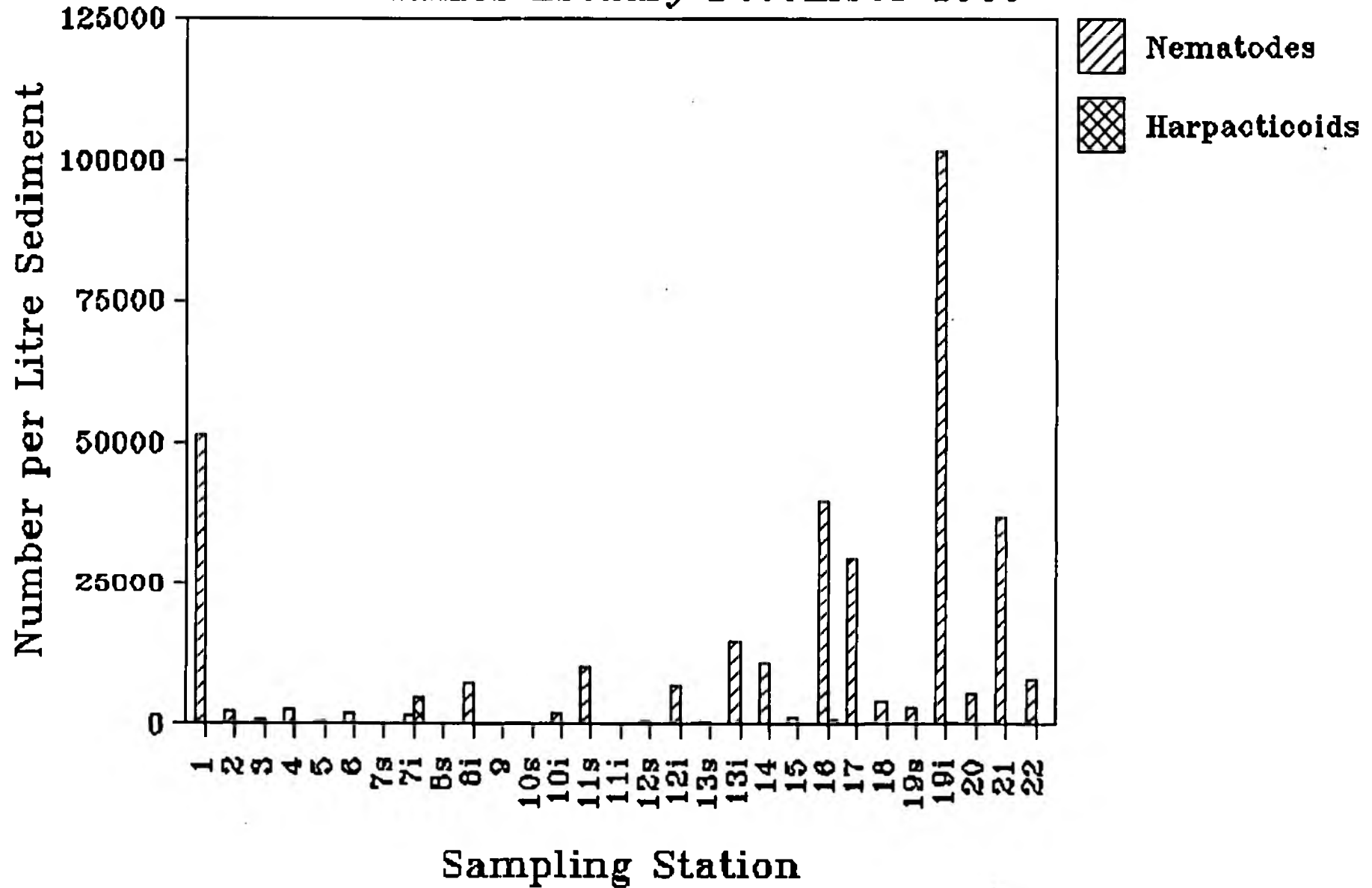


Figure 7. Total densities of nematode and harpacticoid copepod species observed in the Thames Estuary, October - December 1989.

Nematode and Copepod Densities

Thames Estuary December 1989



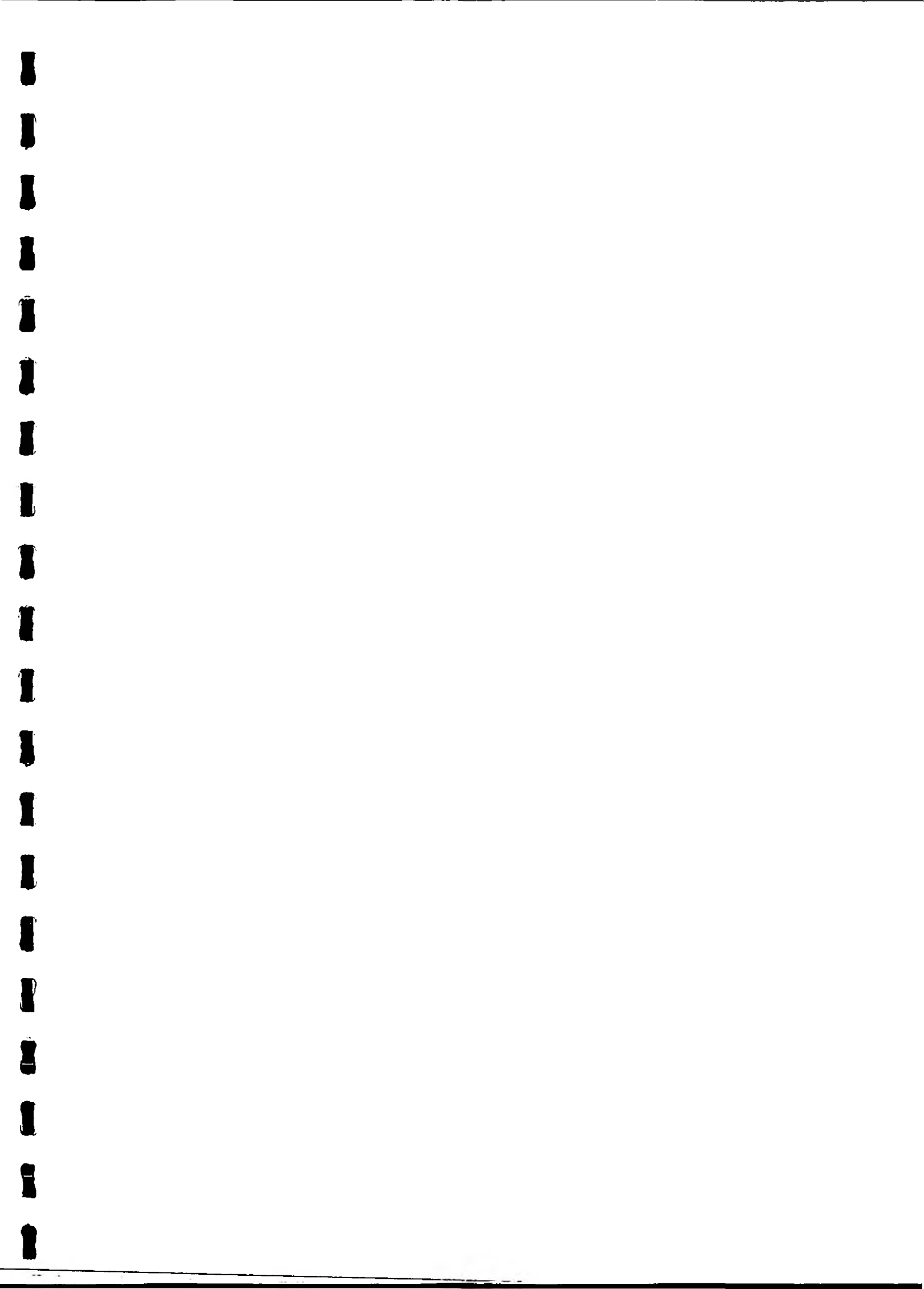
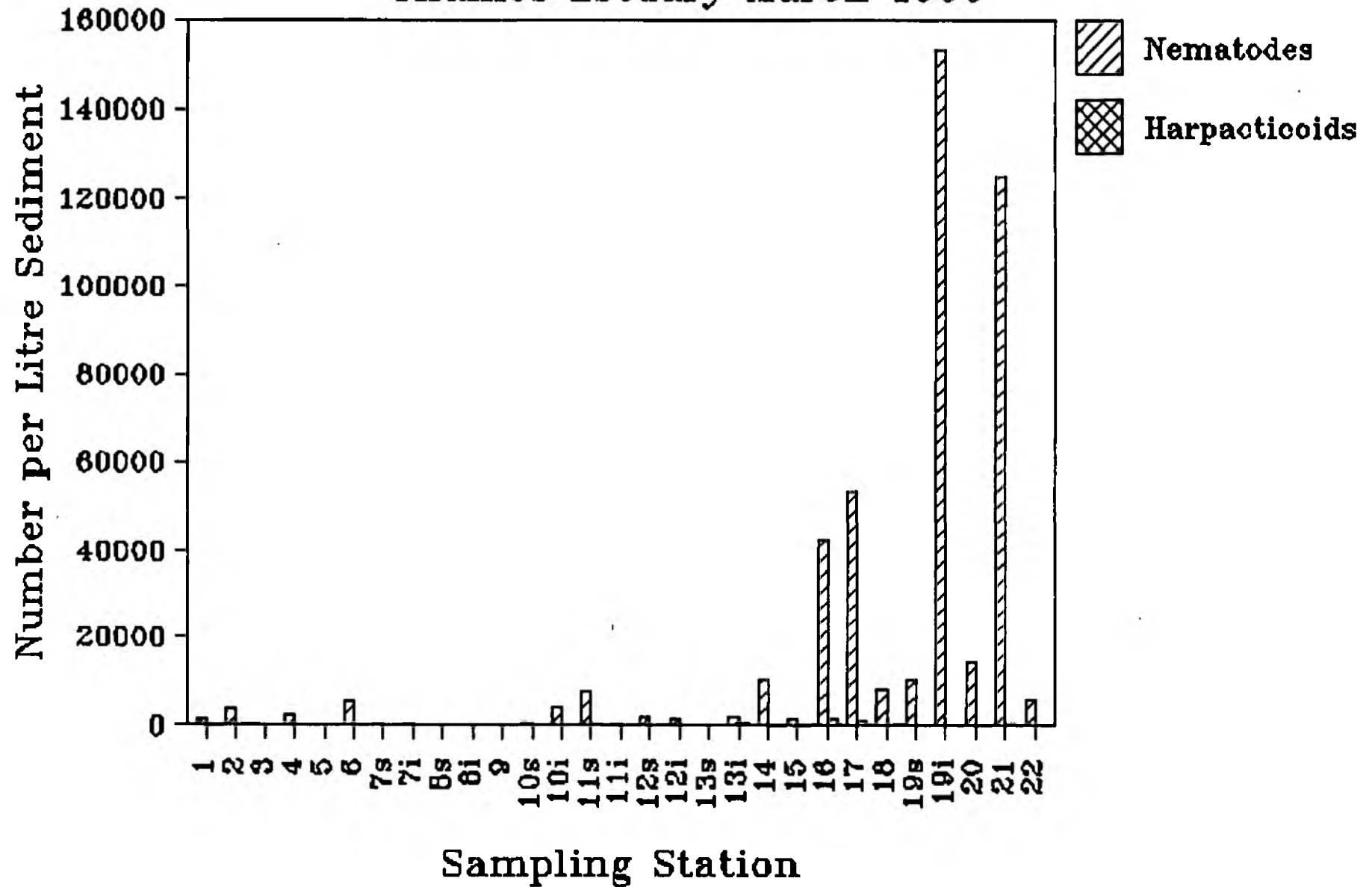


Figure 8. Total densities of nematode and harpacticoid copepod species observed in the Thames Estuary, January - March 1990.

Nematode and Copepod Densities

Thames Estuary March 1990



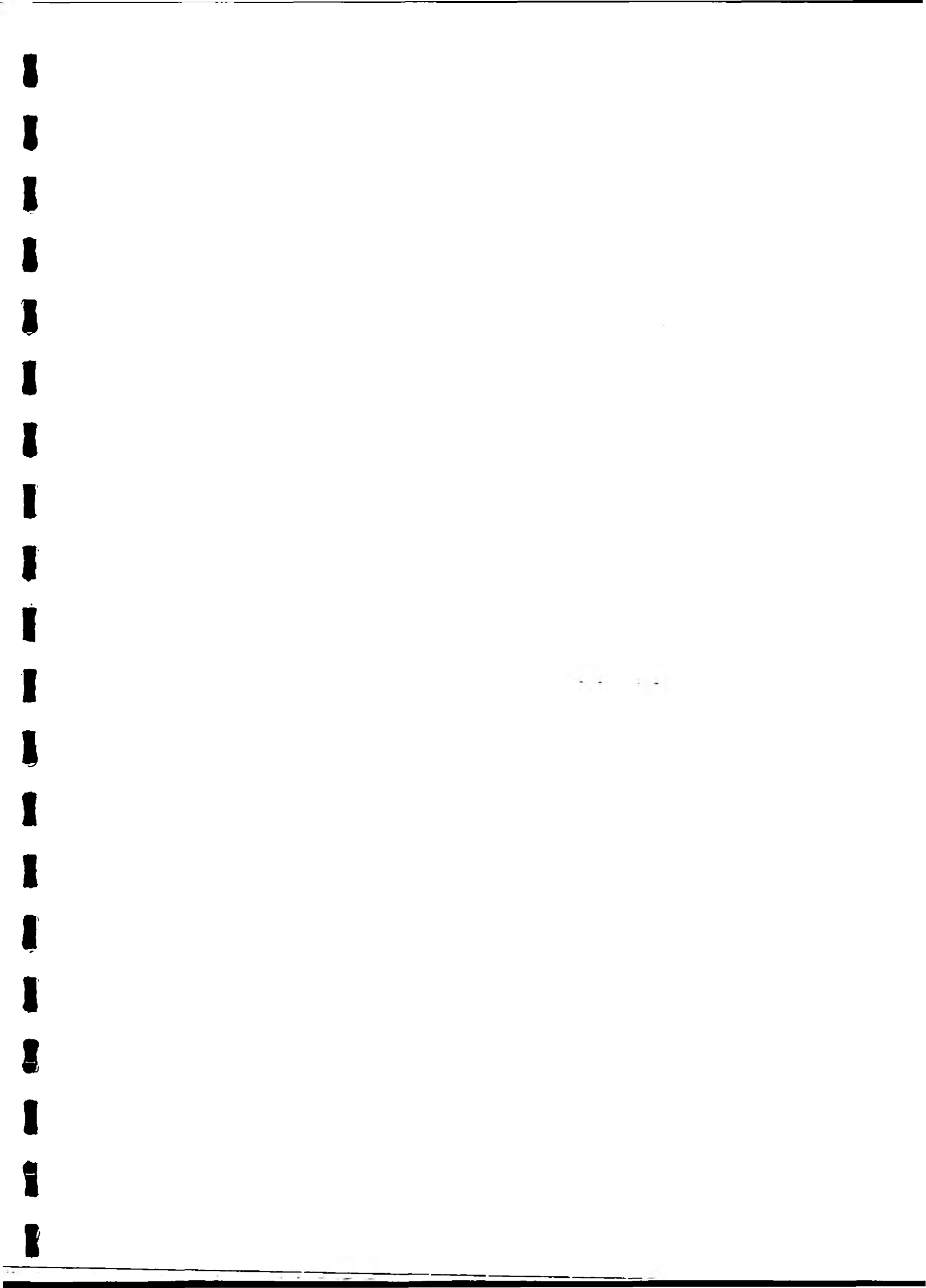


Figure 9. Maximum observed dominance in nematode assemblages, Thames Estuary, April - June 1989.

Nematode Dominance: May 1989

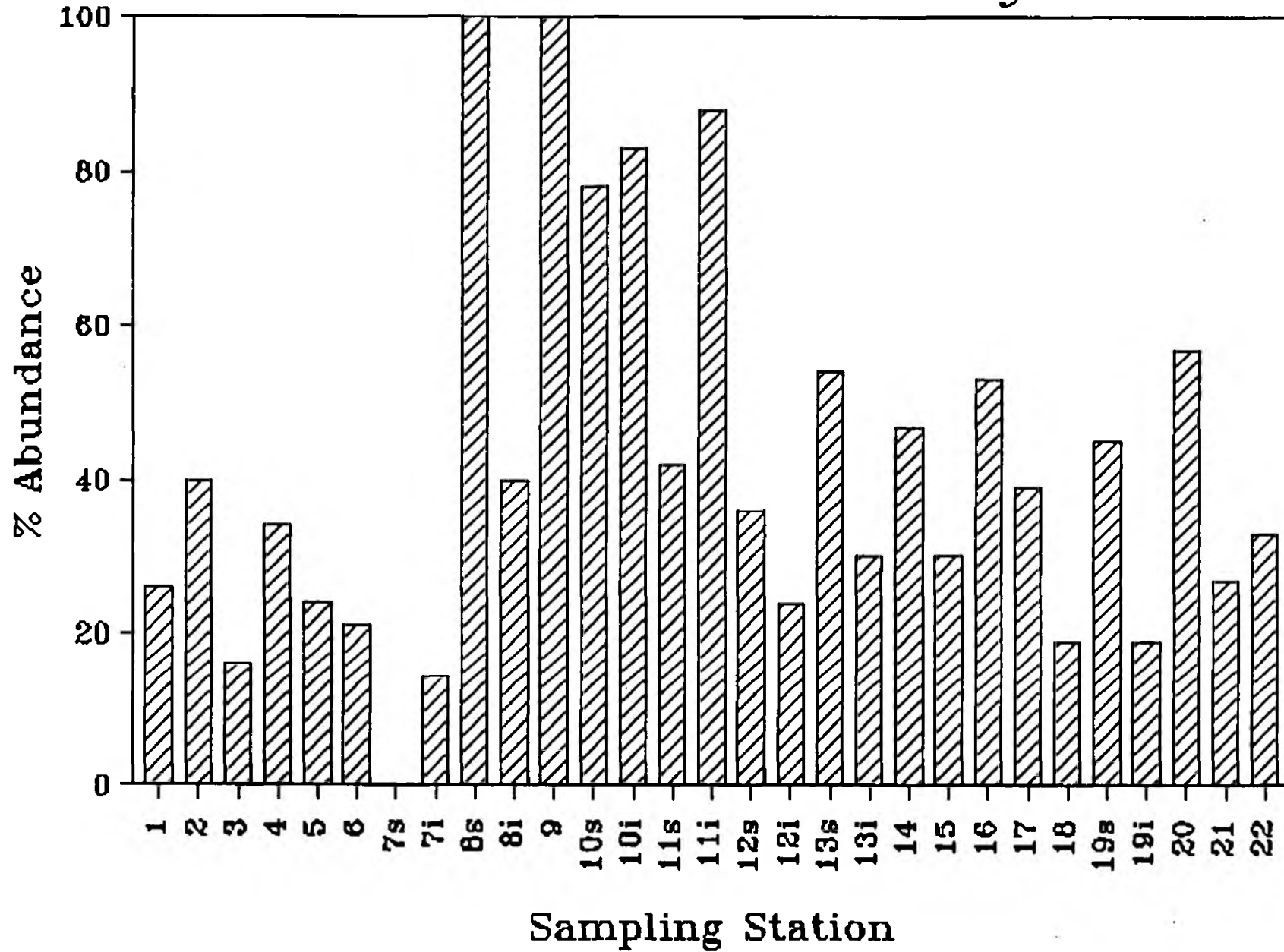
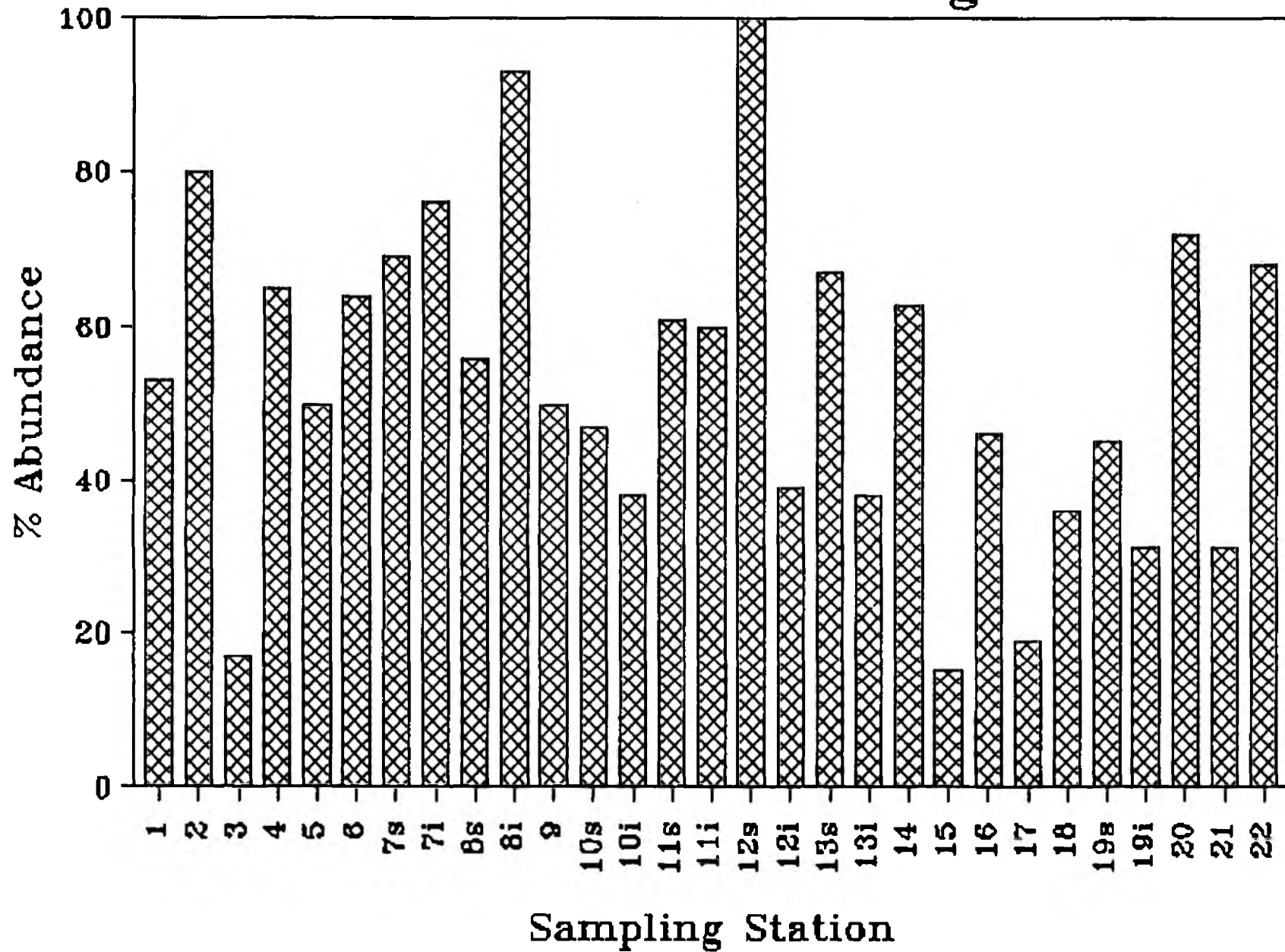




Figure 10. Maximum observed dominance in nematode assemblages, Thames Estuary, July - September 1989.

Nematode Dominance: August 1989



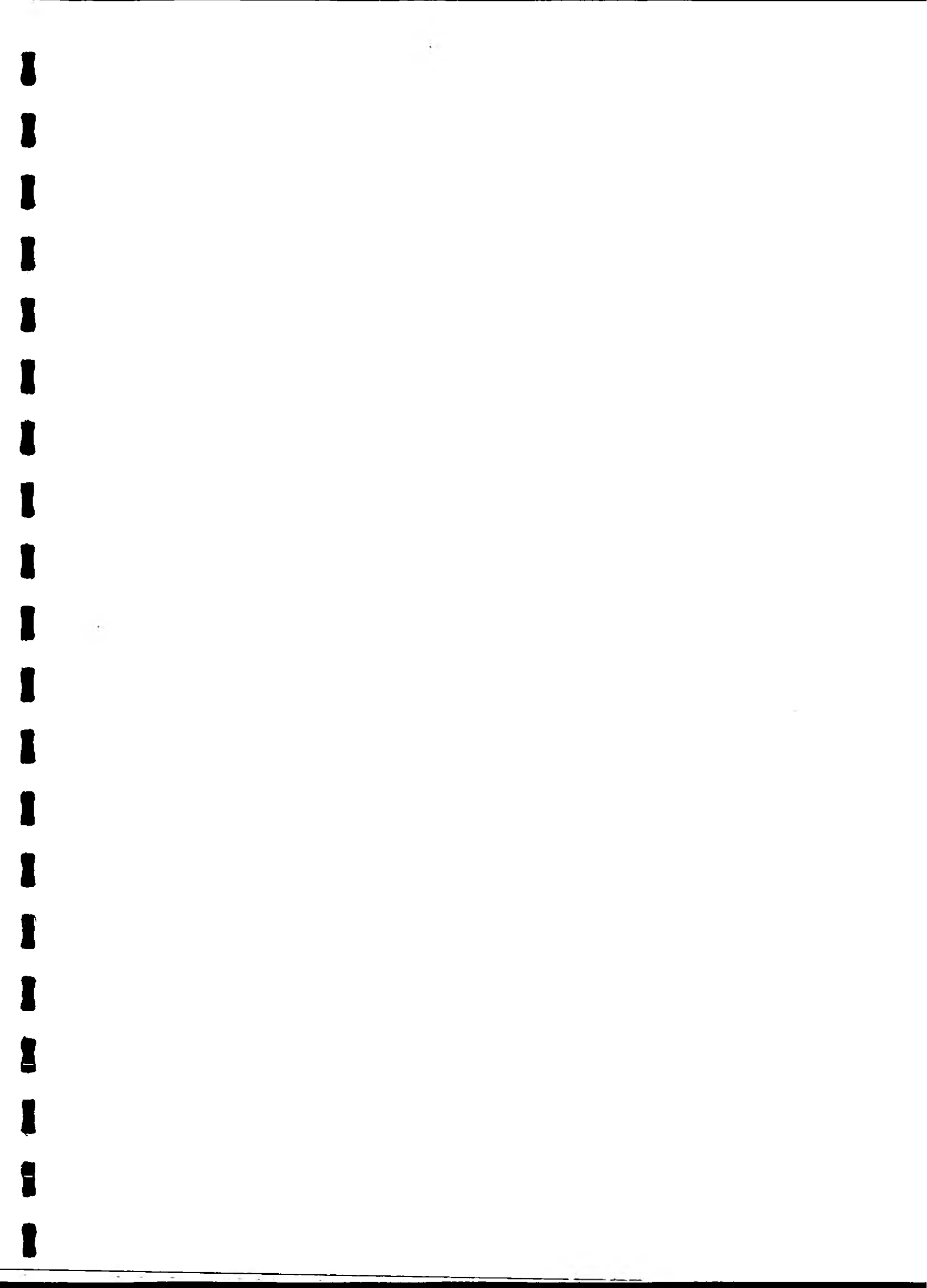
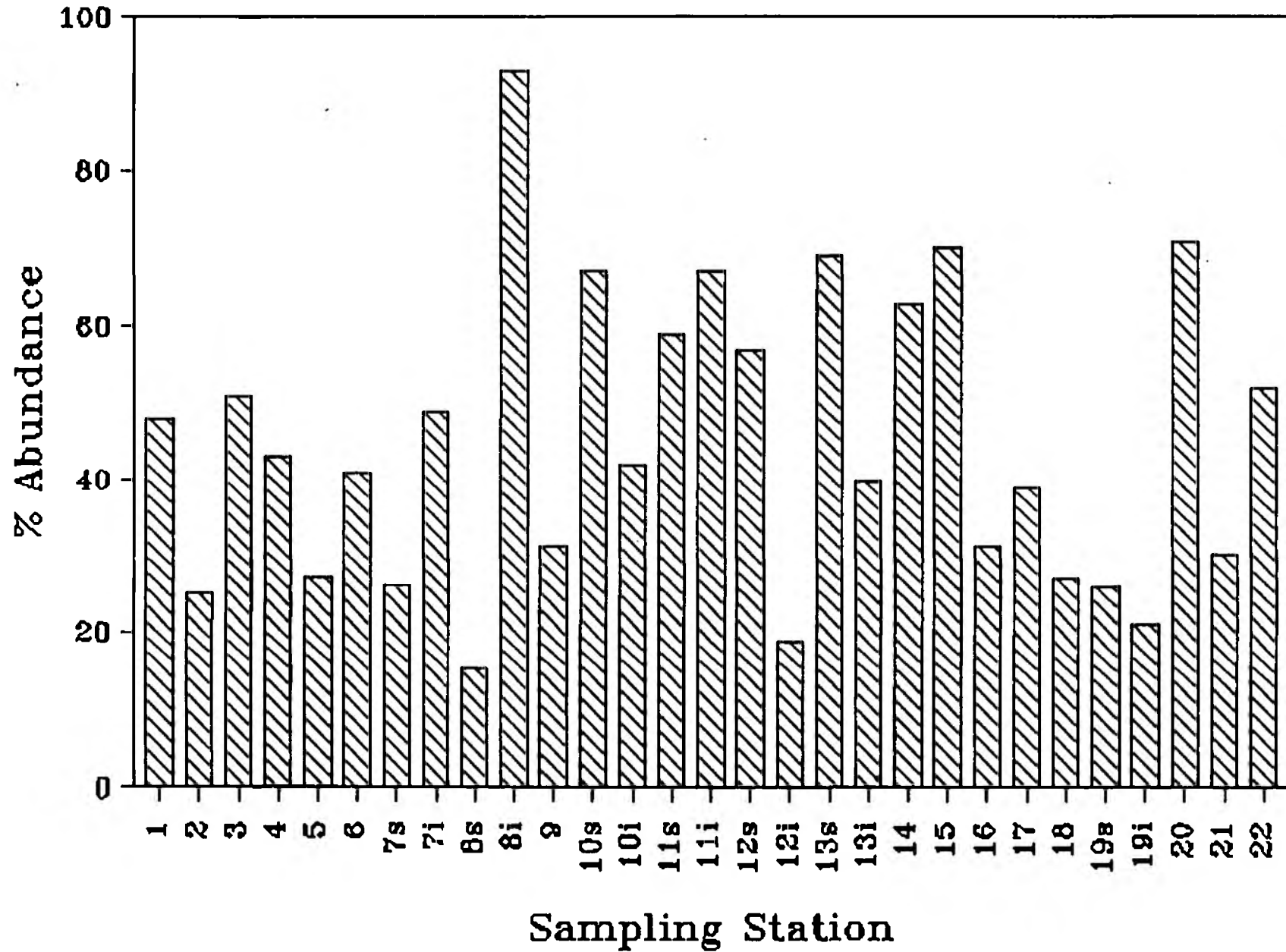


Figure 11. Maximum observed dominance in nematode assemblages, Thames Estuary, October - December 1989.

Nematode Dominance: December 1989



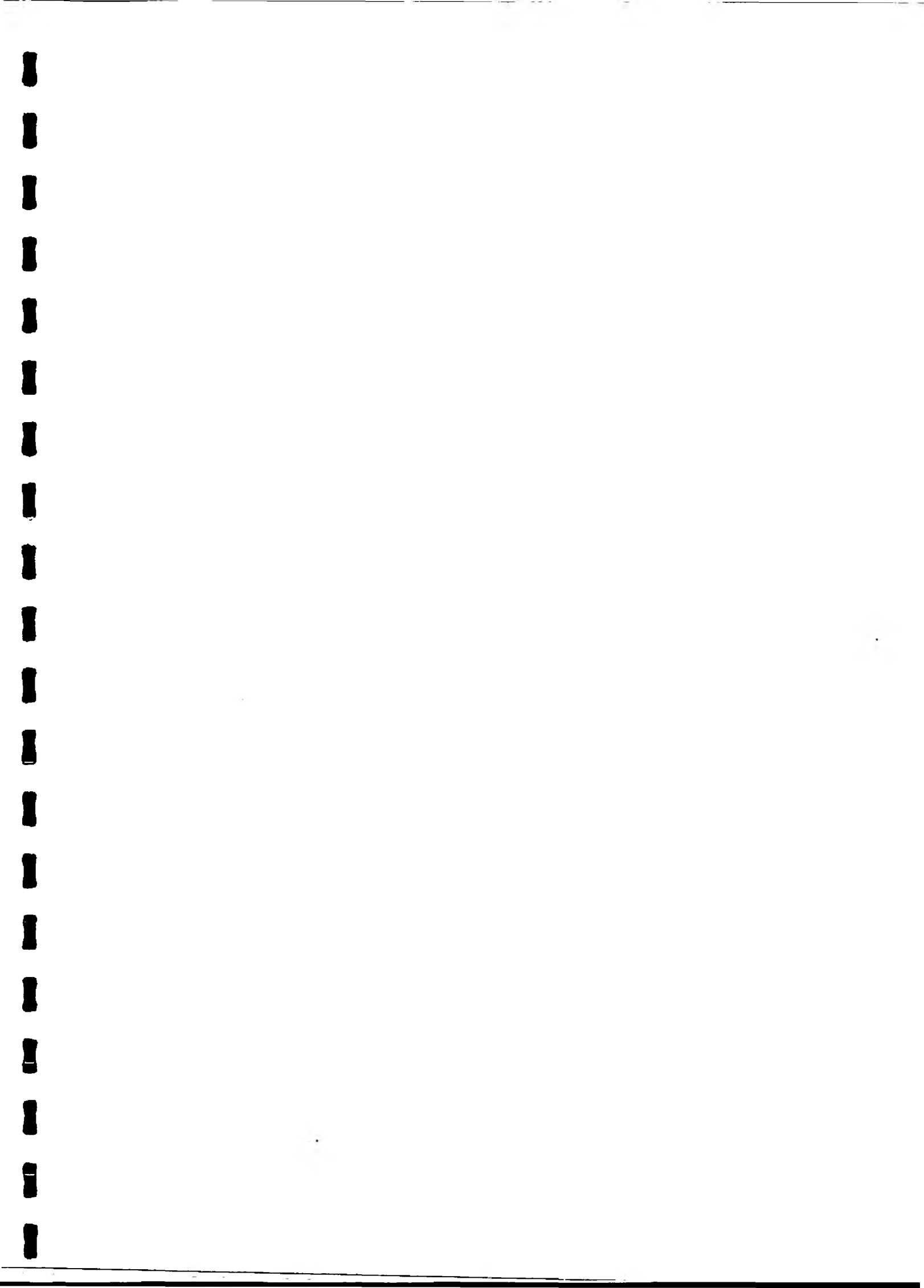
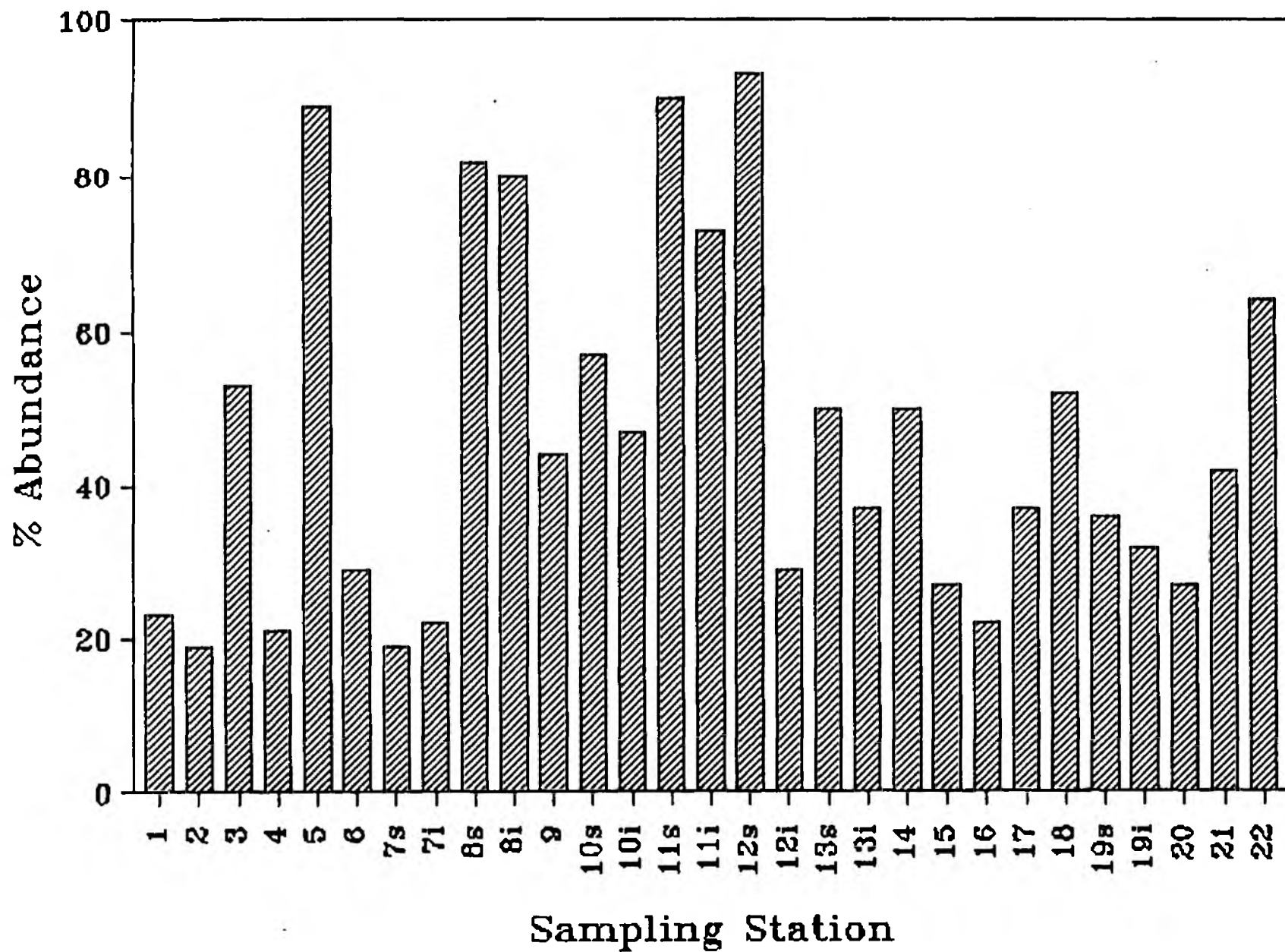


Figure 12. Maximum observed dominance in nematode assemblages, Thames Estuary, January - March 1989.

Nematode Dominance: March 1990





The Meiofaunal Assemblages of the Thames Estuary

April 1989 - March 1990

Module III

**An Assessment of the Impact of Lowflow
and Mechanical Disturbance on the
Thames Estuary as Revealed by its Meiofaunal Assemblages**

by

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1. Summary

Examination of the meiofaunal species complements at the intertidal and subtidal sampling sites within the Estuary was used to assess the effects of reduced freshwater flow from the catchment and to detect the impacts of mechanical disturbance of sediments. Reduction in densities of stenohaline meiofaunal species, intolerant of elevated salinities, at the head of the Estuary was noted following the onset of the drought period (2nd. and 3rd. quarters, 1989). This was accompanied by an extended colonisation of upstream stations by more euryhaline, mid-estuary species and the ingression of true marine taxa into the outer Estuary.

Evidence of increased sedimentation and the deposition of finer particles, possibly as a result of reduced freshwater flow, was noted throughout the mid- and upper Estuary. This exerted a fundamental effect on the meiofaunal assemblages with increased dominance of non-selective detritivore species and reduction in species richness. The changes in these communities was also reflected in the nematode 1B:2A feeding type ratios; populations of type 2A, selective epigrowth and diatom feeding species declined with the increase in fine silt-clay fraction in the sediments. There was some suggestion of increased total meiofaunal densities within the Estuary during this period which, again, is usually associated with finer sediments. However, it was not possible to differentiate this effect reliably from natural seasonal variation in this particular study.

Examples of three different classes of modified meiofaunal community were suggested to indicate mechanically disturbed conditions in the Thames Estuary. These included muddy sediments with low densities of species, residual sands with low diversities of meiofauna and sandy sediments with exceptionally low meiofaunal densities. Cases are discussed from the upper, mid- and outer Estuary. In many cases these were accompanied by a change in granulometry from finer to coarser sediments.

Whilst nematode:copepod ratios have been discounted as useful indices of pollution stress, especially in differing sediment types (see Module II), the ratios were examined in connection with mechanical disturbance effects. The absence of harpacticoid copepods from several sites at different times of the year hindered an accurate assessment of the technique. However, a predominance of low nematode:copepod ratios at certain mid- and outer estuarine sites during the first survey followed by higher ratios in subsequent surveys is suggested to indicate mechanical disturbance. Whether this related to higher freshwater flows and stronger tidal scouring in the early part of the survey could not be determined.



2. The Value of Meiofauna in the Detection of Physical Changes

i. Physical Changes and their Associated Stresses

In the present section, physical changes are taken to include altered freshwater flow rates, caused by reduced rainfall and increased abstraction of water upstream of Teddington Lock, as well as mechanical disturbance resulting natural scouring of currents and from Man's activities.

The consequences of reducing freshwater inputs into an estuarine system are far reaching and undermine the fundamental processes of an estuary. Reducing freshwater inputs will have direct effects on stenohaline freshwater species as higher salinity waters will ingress further into the estuary. Densities of these species, intolerant of the osmotic stress created by the higher salinity water, will be reduced and their distribution patterns downstream severely modified. Euryhaline species, characteristic of mid-estuarine reaches, are also directly dependent on the continuity of freshwater entering the head of the estuary. Although capable of tolerating higher salinities, their ability to survive in a given habitat may be reduced as they are out-competed by other, less euryhaline species that migrate in from the marine environment. A more direct effect on specialist feeders might be the disappearance of preferred food substrates such as single species of algae.

The oxygen carrying capacity of freshwater, at a given temperature, is higher than saline waters. At 5 °C, for example, freshwater saturated with air holds approximately 13 ppm oxygen. Full strength seawater (salinity ca. 35 ppt) at the same temperature will hold less than 10 ppm oxygen. In an estuary, such as the Thames, these seemingly small differences may be critical, especially where low oxygen tension storm waters and sewage effluents are discharged to the tideway. Species within the estuary will exhibit different tolerances to low oxygen tensions and the more sensitive species will be eliminated if deoxygenated waters are repeatedly backed up into fresher water reaches.

A further consequence of reduced freshwater flows will be altered sedimentation and deposition patterns within the estuary. Sedimentation rates in fresh and saline waters differ. Silt has a mean specific gravity of 2.65 and a given diameter particle would normally sink more slowly in seawater than freshwater. However, clumping of silt particles in saline water results in a "salting out" type process. The relative position of the zone of mixing is therefore one factor that determines silt deposition. Another relates to the fact that finer silt particles are also re-suspended on spring tides and carried seaward on the ebb. These are then transported back into the estuary and re-deposited on the surface of lower energy shores or banks. In low flow conditions this may occur further into the estuary than normal. The erosion and accretion of finer particulate sediments over a period of one or two months can severely affect the populations of estuarine species with life cycles that exceed this length of time.

This consequence of reduced freshwater flow is not unrelated to mechanical



disturbance and scouring. Where artificial re-suspension of finer sediments occurs this can lead to smothering of coarse sediment benthic communities at more distant sites. It can, of course, have catastrophic effects on the assemblages of species present in the disturbed sediments. This is commonly seen where dredging activity has occurred. A further consequence of mechanical disturbance may be the disturbance of reduced toxic materials from deeper within sediments. Depending on currents, these materials can exert effects on organisms at considerable distances from the site of the original disturbance.

ii. Meiofauna as Indicators of Physical Disturbances

As sensitive indicators of prevailing environmental conditions (see Module II), the composition, dominance-diversity characteristics and trophic structure of meiofaunal communities can be used directly to assess changes in physical conditions. Specific meiofaunal groups are associated with freshwater ecosystems and many are intolerant of elevated salinities. Successions of euryhaline estuarine species mediate between the freshwater communities and those of marine species in the outer estuary. The differing biologies and physiologies of each species in each these groups, combined with their high densities and diversities, provide a potentially sensitive method for the assessment of changes in the salinity that might be expected to occur within the estuary under conditions of reduced freshwater flow.

Mechanical disturbance can exert several different effects on meiofaunal communities. Where disturbance leads to a change in the granulometry of sediments, characteristic complements of species develop. In general, muds support comparatively few species at high densities whereas coarser grained sediments support lower numbers of individuals representing a larger number of species. Dominance is also usually higher in muddy sediments than in sands. This can be examined using k-dominance analyses. Changes also occur in the feeding guilds of species with larger numbers of specialist feeding types occurring in coarser grained sediments than in muds (see Appendix I, Module IV). In this way, it is possible to monitor physical changes that result in even the subtlest changes in sediment particle size distributions.

The nematode:copepod ratio has also proved to be of some use in the detection of mechanically disturbed sediments, although its value as a means of determining pollution status has been largely undermined (see Module II). Studies of marine sediments (Trett and Feil, pers. obsvn.) have indicated that the more mobile epibenthic harpacticoid copepods re-colonise disturbed sediments more readily than nematodes. Consequently, low nematode:copepod ratios in comparison to assemblages in similar sediment types can provide an indication of recent disturbance. This is especially true where dredging has occurred.



3. Reduced Freshwater Flow

This section deals with the direct effects on the meiobenthos of reduced freshwater input into the Thames Estuary from the catchment. Changes in granulometry of sediments within the Tideway that might result from low freshwater flow and altered sedimentation rates are considered in section 4, below, along with stability of sediments and the effects of mechanical disturbance.

i. Opportunistic Species

With reduction in the flushing rate of the upper Thames Estuary associated with the onset of low freshwater flow conditions, the potential for influence of brackish waters at the head of the Estuary is increased. This can exert profound effects on populations of stenohaline freshwater species, intolerant of increased salinities and enable more euryhaline, opportunist (*r*-selected) species to establish. Evidence for this can be seen in the meiofaunal assemblages. At the Teddington end of the estuary, meiofaunal species assemblages comprised freshwater species (see Module IV). Examination of these assemblages reveals changes in the complements present at each station during each of the surveys. The percentage faunal similarities (calculated using a presence:absence index; Jaccard) between the species present during each of the 4 surveys at the upstream stations are shown in the table below. At Teddington, similarity between consecutive surveys ranged from 23%, between the 1st. and 2nd. surveys, to 47%, between the 2nd. and 3rd.

	SURVEY				Ref. No.
	1st.	2nd.	3rd.	4th.	
Station:					
Teddington	23%	47%	30%		1
Kew	42%	45%	41%		2
Hammersmith B.	19%	8%	18%		3
Cadogan Pier	44%	18%	23%		4
South Bank C.	15%	11%	21%		5
London Bridge	13%	30%	35%		6
Greenwich	7%	41%	45%		7i

Table of Percentage Faunal Similarities Between Species Assemblages in Consecutive Surveys at Upstream Sampling Stations, Thames Estuary, April 1989 - March 1990.



Uniformly high similarity values existed between the meiofaunal assemblages observed at Kew suggesting more that conditions were more stable at this station over the year. However, communities at stations from Hammersmith Bridge to the South Bank Centre were much more variable in composition, exhibiting higher species turnover values. The meiofaunal communities present in the 2nd. and 3rd. surveys at Hammersmith Bridge, for example, showed a similarity of only 8%. If the species present during these surveys at Hammersmith are compared, it is seen that the 3rd. survey contained many more representatives of euryhaline estuarine species. Amongst the Nematoda, these opportunistic species included *Daptonema setosa*, *Leptolaimus* species, *Anoplostoma viviparum*, *Theristus* species and *Dichromadora geophila*. Higher salinity, outer estuarine species were also present including *Axonolaimus paraspinosus*, *Odontophora villoti* and *Daptonema furcata*. The same general effect can be seen between the 1st. and 2nd. surveys but the transition did not occur as dramatically and it is displaced downstream to between the South Bank Centre and London Bridge sampling stations. This is undoubtedly the direct effect of higher salinity waters extending further into the upper reaches of the estuary due to reduced flow and displacing stenohaline freshwater meiofaunal species. Beyond Greenwich other effects begin to appear that may relate to the reduced salinity centred around Beckton (9) and Crossness (10) sewage treatment works rather than reduced flow; in some instances there is evidence to suggest that the sewage treatment works may even stabilise the community structure but in a severely modified form (see Module II).

Evidence for the effects of reduced flow throughout the remainder of the estuary are difficult to separate from seasonal effects relating to increased population sizes of given species extending naturally into more marginal habitats. However, an example discussed in Module IV is worthy of note. This relates to the distribution of the *Sabatieria punctata* population (see Module IV, Section 4.1). The distribution charts show that this euryhaline species progressively extended its upstream range into the estuary from Woolwich (8) to Hammersmith Bridge (3). It did not retreat as might have been expected with the decline in its population densities. This again is seen as an effect of reduced freshwater flow favouring euryhaline species in the upper reaches of the estuary. In the outer estuary, evidence for the increased ingress of more marine species is contentious. Some examples of extended oxystominid distributions can be seen but the distributions of predominantly marine families such as the Desmodoridae extended into the estuary less during the presumed periods of low flow than at other times of the year. It might be postulated that this pollution sensitive group was responding not to increased salinity but to the increased concentrations of poorly mixed pollutants - however, there is no evidence to support this suggestion.

ii. Dominance and Species Richness

Further indications of stress that may relate to periods of reduced flow are seen in the increased maximum dominance that occurs in the nematode assemblages at stations between Teddington (1) and Greenwich intertidal (7i) observed during the second survey. These are illustrated in Module II, Figure 10. There is evidence to suggest that stressed



nematodes assemblages exhibit increased dominance followed by reduced species complements before new, more stable communities establish (pers. obsvn.). In the present study, the increased dominance indicates that reduced freshwater flow may have exerted effects between July and September before the lowest faunal similarities were observed between species assemblages in the 2nd. and 3rd. surveys. This illustrates the potential value of dominance analyses in the early detection of stress that will ultimately lead to a change in species richness of meiobenthic communities and altered abundances. It also emphasises the sensitivity of the populations of individual species to changes in prevailing conditions. It is not possible to determine the seaward limit of the effects of reduced flow in the Thames Estuary; dominance at Woolwich (8) was high throughout the year, possibly as a result of the influence of the Beckton and Crossness sewage treatment works (see Module II). In this connection, there is no evidence to indicate whether the sphere of influence of the sewage works extended further into the estuary during the periods of low freshwater flow.

4. Stability of Sediments and Mechanical Disturbance

i. K- and r-selected Species

A broad spectrum of K- and r-selected species of meiofauna were recorded in the Thames Estuary over the year (see Module IV). The low densities and patchy distribution of the larger predatory and omnivorous nematode species (type 2B) were notable, especially amongst the order Enoplida (e.g. families Enoplidae, Thoracostomopsidae, Oncholaimidae and Enchelidiidae). Populations of these species are slow to establish and may take several years to reach equilibrium. Many species employ sticky eggshells and caudal cement glands to prevent displacement from the sediments. Where these groups were present, they were usually found in littoral muds and muddy sands and, in one case (Chapman Buoy (18)), in sublittoral sands. In each instance, the sediments that supported these species can be classified as "mechanically stable". Examples of this include intertidal sites at Gravesend (13i) and Southend (19i).

By default, sediments that do not support well established populations of these species might be classified as unstable. Indeed, in the Thames many of these sediments were colonised by populations of opportunist, r-selected species that have short life-cycles and produce large numbers of eggs/offspring. Amongst the Nematoda, these include members of the families Comesomatidae, Xyalidae and Monhysteridae that are predominantly non-specialist feeding types. Examples of stations at which this type of meiofaunal community have been observed include the South Bank Centre (5) and several of the subtidal stations in the outer estuary such as Grain Flats (20). These are discussed in more detail below.

ii. Dominance and Species Richness

In general, coarse grained sediments support higher numbers of meiofaunal



species at lower densities than muds at equivalent salinities and vice versa. Mechanical disturbance caused, for example, by scouring currents will often reduce meiofaunal densities as finer sediment fractions are re-suspended and transported up or downstream. However, species richness may remain relatively unaffected by this process. Consequently, muds with low densities of meiofauna, residual sands with low diversities of meiofauna and sandy sediments with exceptionally low meiofaunal densities may all be indicative of recent mechanical disturbance. In the Thames Estuary there were examples of all of these categories. Throughout the year, for example, the sands at the South Bank Centre (5) supported lower numbers of meiofaunal species than might have been expected (see Module II and Module IV) and the subtidal muds at West Thurrock (12s) yielded one species of meiofauna at low density ($<40 \text{ litre}^{-1}$) in the 2nd. survey and comparatively low densities of meiofauna in the following survey. The difficulty lies in determining whether these observations relate to pollution incidents or physical disturbance. The presence of certain more specialist species at the South Bank Centre might undermine the pollution explanation, adding strength to the mechanical disturbance hypothesis. Diatomivorous and microbivorous nematodes present at low densities at this site might not be expected to tolerate pollutants that would affect their food sources. A similar argument could be advanced for the species present at the subtidal site at West Thurrock during the 3rd. survey with *Monhystera* and *Leptolaimus* species (type 1A, microbivorous species) occurring at low densities. However, the observation of a single species, *S. punctata*, at low density in the previous survey is less easy to interpret.

Changes in sediment type were noted between consecutive surveys that might indicate mechanical activity and deposition. Grain Flats (20) is a good example of this (see Modules II and IV). The increased fine fractions in the sediments correlated directly with a decrease in meiofaunal species along the lines that would be predicted for an outer estuarine site. These changes were accompanied by an increase in dominance during the first 3 surveys (Module II, Figure 9 to 11). Elsewhere, changes in sediment types were also noted during the survey (see for example Cadogan Pier) which might relate to reduced flow (see also Module IV).

Meiofaunal dominance-diversity curves for stable communities examined at different times of the year exhibit a high degree of congruence. Once equilibrium is reached, the habitat will support a climax community consisting of a finite number of permanent species with a reduced turnover. This, in turn, reflects stability of the substrate. Examples of this were seen at the intertidal site at West Thurrock (12i), Allhallows (17) and Shoeburyness East (21).

iii. Nematode:Copepod Ratio

Whilst doubts have been raised concerning the value of nematode:copepod ratios in detecting pollution stress (Lambhead, 1984), they may be of some value in the detection of mechanically disturbed sediments (Trett and Feil, pers. obsvn.). Exceptionally low ratios can result where there has been an influx of mobile epibenthic harpacticoid copepods before nematode populations have been able to re-establish. The failure to observe copepods at certain stations was a serious drawback to this approach. However, the lowest ratios (those less than 10.0) are given in the table below.



The values might indicate greater general instability of sediments during the early part of 1989, possibly linked to higher flow levels. However, this is more than a little conjectural. The value for Grain Flats is of some interest given the discussions above. The lowest ratio observed (Greenwich intertidal, 3rd. survey) reflects an exceptionally high density of harpacticoids present during this survey - epibenthic species dominated by a *Microarthridion* species. However, there is little evidence to support the suggestion that this habitat had been physically disturbed. Indeed, considerably lower densities of nematodes were observed in the 1st. and last surveys at this site than in the 3rd. survey. Unfortunately, copepods were not observed at The South Bank Centre throughout the year or at the subtidal West Thurrock site during the 2nd. and 3rd. surveys preventing comparison of the nematode:copepod ratios with sites for which some evidence for disturbance might already exist.

	SURVEY			
	1st.	2nd.	3rd.	4th.
Station:				
London Bridge	6.8			
Greenwich (i)			0.3	
Woolwich (i)				2.9
Gravesend (s)				1.6
Gravesend (i)	4.6			3.7
Blythe Sands	3.8			
Grain Flats	7.9			

Table of lowest (<10.0) nematode:copepod ratios observed in the Thames Estuary, April 1989 - March 1990. (s - subtidal site; i - intertidal).

In summary, the nematode:copepod ratio probably provides less information about the stability of sediments in the Thames Estuary than meiofaunal similarity studies owing principally to the patchy occurrence of harpacticoids and their low densities. On the basis of faunal similarities, higher salinity sites such as Purfleet (13) become of interest where 0% similarity exists between certain consecutive meiofaunal surveys (3rd. and 4th. surveys, station 11i). Copepods were also absent from the muds at this site during the 3rd. survey.



5. Future Studies

A particular area of interest is the extent to which seasonally expanding populations of marine species of meiofauna will penetrate into the Thames Estuary under normal conditions of freshwater flow. This may be difficult to assess with trends towards drier summer months and ever increasing demands on water supplies in the Thames catchment. A sampling programme along the lines of that undertaken in the present study would provide valuable information about this aspect of estuarine recruitment if rainfall in future summers was sufficiently different from that of 1989. In terms of mechanical disturbance, increased freshwater flow during the winter months might increase the scouring action of tides on the ebb. Examination of changes in subtidal meiobenthic communities could confirm this and add considerably to our understanding of the dynamics of meiofaunal populations in estuarine systems. Attention might also be focused on sites at which constructional activity affects the Tideway. In such instances, it would be of interest to determine whether disturbance of toxic materials, formerly buried in sediments, exerted direct effects on meiofaunal communities that could be detected and monitored. Dredging activities in Dutch estuaries have already been implicated in this type of impact on macro- and meiofaunal assemblages (Trett, pers. obsvn.).



**THE MEIOFAUNAL ASSEMBLAGES
OF THE THAMES ESTUARY**

April 1989 - March 1990

Module IV

by

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1. Summary

Meiofauna was examined by staff at *Physalia Ltd.* in 28 sediment samples collected by the National Rivers Authority, Thames Region, from 22 intertidal and subtidal sites in the Thames Estuary. Samples were collected in the second, third and fourth quarters of 1989 and the first quarter of 1990. From east to west, the sites sampled were located at Teddington, Kew, Hammersmith Bridge, Cadogan Pier, The South Bank Centre, London Bridge, Greenwich, Woolwich, Beckton and Crossness sewage treatment works, Purfleet, West Thurrock, Gravesend, Mucking, Blythe Sands, Canvey Beach, Allhallows, Chapman Buoy, Southend, Grain Flats, Shoeburyness East and Sea Reach Number 2 Buoy. The principal meiofaunal groups present were nematodes, harpacticoid copepods and aquatic mites. Other meiofaunal groups were present at lower densities. Nematoda dominated all the assemblages examined. In total, 207 nematode taxa from 43 families were observed, although not all were present in any one survey. Total densities of nematodes ranged from 8 litre⁻¹ sediment at Beckton during the second quarter of 1989 to over 153,000 litre⁻¹ sediment at the Southend intertidal sampling site in the first quarter of 1990.

A total of 18 harpacticoid copepod taxa representing 12 families were observed. These were most abundant at the Greenwich intertidal site during the last quarter 1989 where 4,549 litre⁻¹ sediment were recorded. Although present throughout the estuary, copepods were not observed at several of the stations examined. Acari were represented by 6 taxa sparsely distributed throughout the Estuary. These comprised bdellid, oribatid, hydracarine and halacarid species. The highest densities observed were 44 litre⁻¹ at the Crossness intertidal site in the 4th. quarter 1989.

The other meiofaunal groups observed included rhizopod amoebae, larger ciliated Protozoa, Kinorhyncha (*Pycnophyes* species), Gastrotricha (?*Chaetonotus* species), Tardigrada (*Macrobionus dispar* and *Batillipes mirus*), planispiral and multilocular Foraminifera (e.g. *Elphidium* and *Braziliana* species) and several species of Turbellaria, Gnathostomulida and Ostracoda. The occurrence and distribution of representatives of several non-meiofauna groups also present in the sediment samples are also described. These include nemerteans, rotifers, annelids, molluscs, Crustacea and larval Diptera (principally Chironomidae).

The high diversity of the meiofauna reported is unique amongst British estuarine studies and appears to reflect the large range of habitats encompassed by the sampling programme. In general, meiofaunal densities and species richness conform to those described for macrofaunal species with diverse, high density assemblages occurring towards each end of the estuary. High species richness correlated well with the higher salinity sites and those with heterogeneous, coarse grained sediments. Lower meiofaunal species richness and high densities were largely associated with the finer grained or high silt/clay fraction sediments.



2. Introduction

Meiofauna are sediment-dwelling species that measure, in general, less than 1 mm in length. They are of fundamental importance in aquatic ecosystems forming food for macrofaunal invertebrates, fish and possibly certain avifauna (e.g. waders). They offer many distinct advantages in the study of prevailing environmental conditions. Unlike other animals, the less mobile meiofauna are subjected continuously to the effects and constraints of any "foreign" materials that enter their environment and this is reflected in the composition of their communities (Newell *et al.*, 1990a and b). Being nearer to the base of food webs than macrofaunal organisms and having shorter generation times, meiofauna respond more rapidly to changes in the quality and nature of their food supply. This can be used to provide early indications of improved or worsening conditions.

In addition to their sensitivity to environmental changes, meiofauna comprise exceptionally diverse groups, such as the Nematoda ("roundworms"). Combined with their high densities, this high diversity makes statistically valid sampling easier than with macrofaunal organisms and, in the case of point source discharges, greatly improves the resolution of studies aimed at detecting impact zones. This has proved to be a valuable tool for monitoring the improvements in the quality of effluents (Trett *et al.*, 1990; Newell *et al.*, 1990a and b) and has been employed by several industries and authorities as part of their environmental audit programme.

The disadvantages of using meiofauna in the assessment of prevailing environmental conditions is the requirement for expert taxonomists skilled in the identification of "difficult" groups such as the Nematoda, harpacticoid Copepoda and halacarid Acari. This has been overcome by organisations such as *Physalia* who provide routine meiofaunal services to environmental consultancies, authorities and industrial concerns throughout the world.

The aim of the survey reported here is to complement the existing Thames Estuary Benthic Programme undertaken by the National Rivers Authority (Thames Region) (see Attrill, 1990a and b). A further aim was to critically evaluate the use of meiofaunal studies in the examination of conditions in the Thames Estuary with especial attention to the pollution status and the possible effects of reduced freshwater flow resulting from the drought period. These aspects are reported elsewhere. Staff at *Physalia* were retained through the Centre for Research in Aquatic Biology, University of London, by the National Rivers Authority, Thames Region (NRA) to examine the meiofaunal assemblages present in 4 sets of sediment samples collected over a period of one year from the Thames Estuary. This module describes the methods used and discusses the meiofaunal assemblages observed. Samples were collected from Teddington to Shoeburyness between April 1989 and March 1990 by NRA staff. Further details of the sites sampled are given in the NRA reports (Attrill, 1990a; 1990b).



3. Materials and Methods

120 - 200 ml subsamples were taken by staff at *Physalia* from formalin-fixed sediments collected by NRA from 29 sites at 22 sampling stations in the Thames Estuary (Figure 1). Initial separation was carried out using a modified Boisseau apparatus (after Macintyre and Warwick, 1984) and fractions collected at increasing water velocities onto 38, 50, 75, 100 and 150 μm sieves immersed in flowing tapwater (see Flegg and Hooper, 1970). Pooled meiofauna/silt-clay fractions for each sample were further separated using density separation techniques and meiofauna collected onto 38 μm sieves. Residual materials were examined to confirm complete elution of infauna. Harpacticoid and calanoid copepods were removed by hand using mounted needles and dissected for identification by means of 5th. limb setotaxy. Acari were also removed at this stage and mounted in polyvinyl lactophenol. After clearing (approximately 3 days at room temperature) specimens were identified and enumerated.

Remaining fauna, principally Nematoda, was processed to glycerol using the Seinhorst method (Seinhorst, 1959) at 40°C in a vacuum oven and mounted on slides for identification and enumeration. All microscopic examination was carried out using Zeiss Nomarski and Nikon differential interference contrast microscopes (DIC). For Nematoda, the first 100 specimens encountered in a standardised "box scan" (excluding unidentified specimens) were identified and counted. The presence of other nematode species observed during subsequent counting (consequently accounting for less than 1% of the nematode population) was also recorded.

Where necessary, identification of species was confirmed by comparison with the meiofaunal reference and specimen collection maintained by taxonomists at *Physalia*. Accurate drawings and morphometric measurements were made for each unidentified species within a genus or, rarely, family to maintain continuity between surveys; essential should subsequent multivariate analyses of species assemblages be required. Data were transformed to numbers of individuals per species per litre sediment sampled enabling direct comparisons to be made between stations and surveys.

Particle size determinations were performed by dry sieving techniques as described by Buchanan (1984) using standard phi range sieves (Endecotts Ltd.) between 2 mm ($\phi = -1$) to 63 μm ($\phi = +4$) and a dry bed shaker.

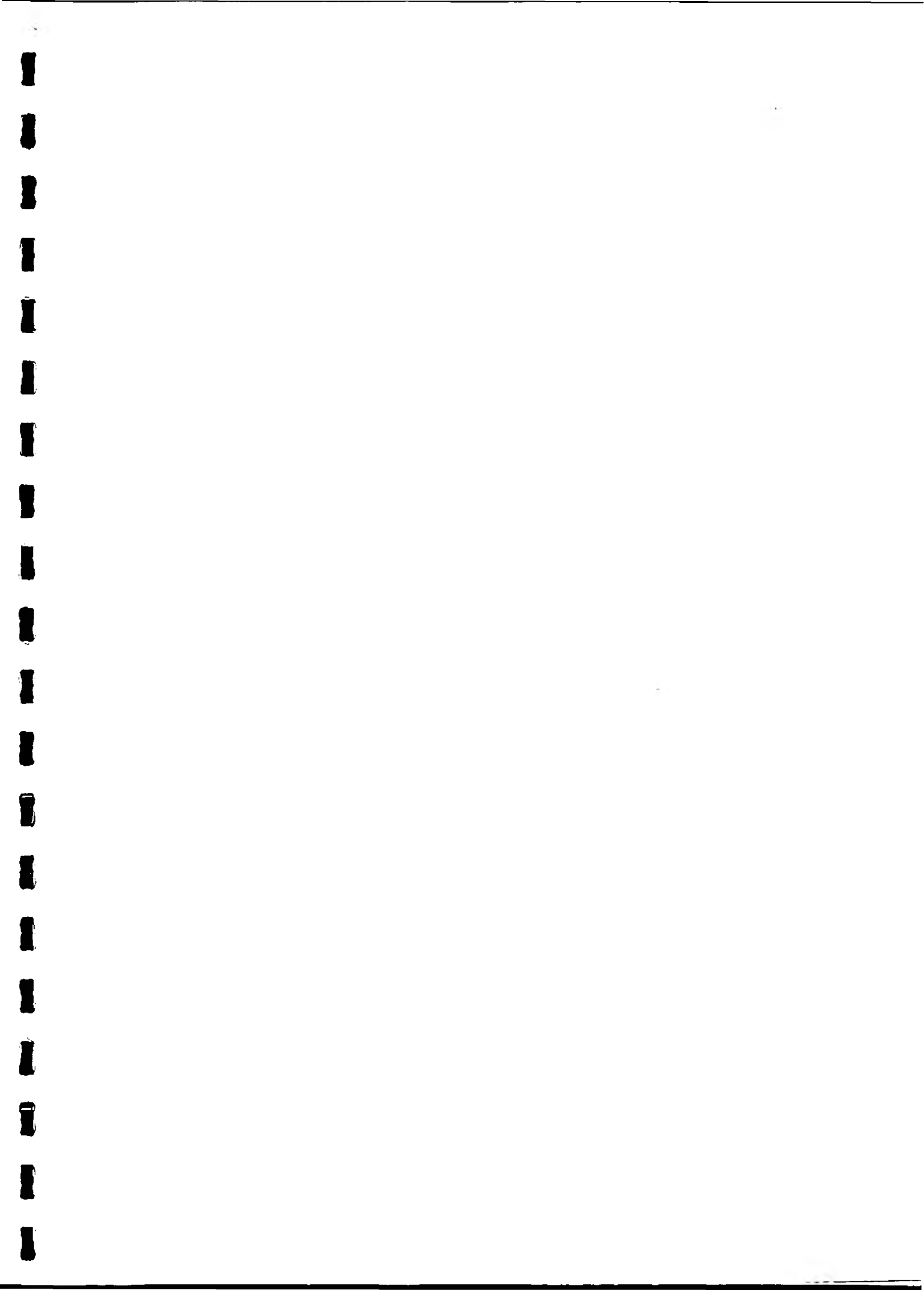


Figure 1. Location map to show positions of 22 principal sampling stations in the Thames Estuary meiofaunal survey, April 1989 - March 1990.



In the order Monhysterida, *Sphaerolaimus gracilis* and *Desmolaimus zeelandicus* are both typical of lower salinity habitats and, as in the example above, are progressively replaced by other, closely related species towards the mouth of the estuary. Although said to be a non-selective deposit feeder (type 1B species) by some authors, *D. zeelandicus* may feed selectively on diatoms (pers. obsvn.). *S. gracilis* is a type 2B species and specimens were seen in the process of ingesting juvenile *Sabatieria* species (?*S. punctata*). Little is known of the behaviour used by this group of nematodes to capture prey and almost nothing is known of the dynamics of nematode predator-prey interactions. This emphasises the need for further study of the biology of aquatic Nematoda. In samples from the seaward end of the estuary, the more euryhaline *Sphaerolaimus balticus* was also present.

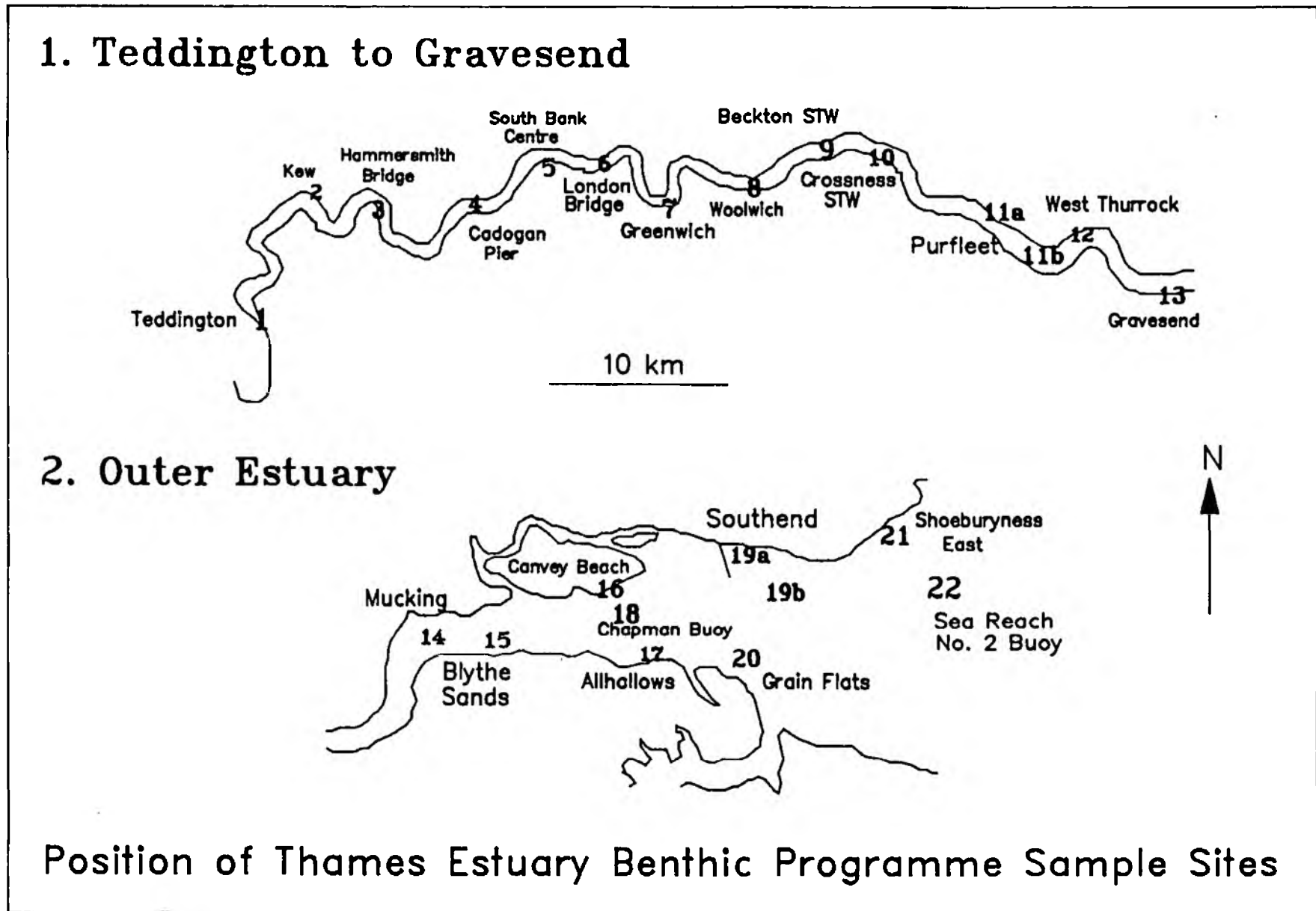
With the exception of a few almost ubiquitous species (e.g. *Daptonema setosa*), the bulk of the nematodes seen in the mid-estuary sediments are euryhaline 'marine' species at the extremes of their ranges. The distributions of these species represent realised niches as opposed to their preferred niches reflecting resource competition. The result is a typical estuarine gradation with extended species overlaps in areas of suitable sediment types (see Green, 1968).

An interesting example of species distribution is given by *Sabatieria punctata*. This type 1B species is an exceptionally common comesomatid in British estuaries where it can occur at extremely high densities (up to 25,488 litre⁻¹ sediment were recorded intertidally in the Tees Estuary in 1988; Trett in Newell *et al.*, 1989b). It is also has a widespread occurrence at lower densities in full marine sublittoral habitats. In the present study, densities of *S. punctata* increased over the summer months reaching a maximum of 24,757 litre⁻¹ sediment at Canvey Beach (16) between July and September (Appendix I; Section 2). Mean densities then declined with the onset of autumn but remained above the pre-survey levels recorded in the 1st. survey. Throughout, the muds at Canvey Beach retained the highest densities of *S. punctata* observed in the Thames. The distribution of the population also changed during the survey period. In the 1st. survey, *S. punctata* was confined to stations between Woolwich and Shoeburyness East with low densities occurring at each end of its range (Figure 2). In the 2nd. and 3rd. surveys, the population extended upstream to Greenwich (7) and the South Bank Centre (5), respectively (Figures 3 and 4). In the final survey, this range was extended still further, despite the decline in mean numbers, with a low density of *S. punctata* (< 10 litre⁻¹) occurring at Hammersmith Bridge (3; Figure 5). This extended distribution coincided with a marked increase in densities of *S. punctata* at the subtidal site at Purfleet. These changes may have related to increased salinity in the upper reaches of the estuary and/or changes in sediment types resulting from reduced flow following the onset of drier weather.

ii. Harpacticoid Copepoda

The Thames Estuary supports a variable and diverse complement of harpacticoid Copepoda. Between April 1989 and March 1990 a total of 47 species of harpacticoid copepod were observed representing 12 families (see Appendix I; Section 1). Numerous unidentified copepodites (principally belonging to the family Diosaccidae) were also present

Figure 1.





4. Results and Discussion

I. General Observations

Samples ranged in granulometry from compacted clay muds and flocculant, high silt-clay fraction sediments to semi-clean sands. Sediment from Teddington, Kew and Cadogan Pier comprised muds with some large stones which made accurate sub-sampling difficult. The particle size distributions of sediments collected in the 4th. quarter 1989 are given in Appendix II. Sediments at either end of the estuary were predominantly coarse grained (usually sands with differing detritus contents). The middle reach sediments comprised mostly muds which were occasionally cohesive. There was some evidence for changes in the sediment types during the course of the survey, possibly as a result of changes in flow of the Thames during the drier summer months. This was most apparent at Cadogan Pier (4), Grain Flats (20) and Sea Reach No. 2 Buoy (22; see Section II below). With the exception of Kew (2), which contained a large proportion of macrophyte detritus, the organisms were separated cleanly from the sediments. Partially decomposed plant material is often of a similar density to the meiofauna and can hinder density gradient separation. However, in all cases, examination of residues showed that loss through inefficient separation was minimal. The biological material was well fixed and showed no signs of deterioration post-collection. "Oily" films were noticed in association with samples collected from Woolwich (8), Crossness (10s), Purfleet (11s) and Southend (19s).

a). Principal Meiofauna Groups

i. Nematoda

The Thames Estuary contains a rich assemblage of nematode species. In total, 207 nematode species belonging to 43 families were recorded in the sediment samples examined over the four surveys (see Appendix I; Section 1). The assemblages present at each station are discussed in detail in Section II below. The nematodes observed during the survey included species usually found in association with semi-terrestrial and freshwater habitats, brackish water species, true estuarine species and full marine species. The range of habitats encompassed in the survey means that the observed numbers of species is almost unique amongst the estuaries studied in Northern Europe (see for example, Capstick, 1959; Riemann, 1966; Warwick, 1971; Bouwman, 1983). In general, intertidal habitats yielded higher densities and diversity of nematodes than equivalent subtidal habitats (see Appendix I; Section 2). This was as expected. Similarly, with few exceptions, the high salinity sites towards the mouth of the Estuary supported larger numbers of species at higher densities than sediments collected further inland.

The Tylenchida and Dorylaimidae examined were all plant parasitic species commonly found in association with the root systems of plants. These frequently occur in freshwater sediments where the rhizosphere of marginal plants enters the littoral zone. Others are also known to feed on the tissues of freshwater euhydrophytes. In both groups, feeding is by means of a hollow stylet, the shape and size of which is of considerable taxonomic importance. These species, along with the microbivorous rhabditids, are often



transported into more saline reaches of estuaries which may account for their sporadic occurrence at stations neighbouring freshwater inputs (e.g. *Rhabditis* species at Beckton sewage treatment works (9), second quarter 1989, and *Criconemoides* species at Mucking (14), second quarter 1989). With one exception, none is known to occur naturally in saline or marine conditions.

Several other freshwater species were observed. These include detritivores and diatomivorous species such as *Plectus granulosus*, *Cryptonchus* species, *Prismatolaimus verrucosus*, *Tobrilus gracilis*, *Tripyla affinis*, *Monhystera stagnalis* and the *Mononchoides* species as well as the predatory or omnivorous species *Ironus ignavus*, *Mononchus aquaticus* and members of the family Diplogasteridae (see Appendix I; Section 5). The above species are typical of the upper, low salinity reaches of estuaries supplied by rivers from mixed chalk and acid catchments and occur throughout the south east of England and East Anglia (Trett, pers. obsvn.). From a biological standpoint, it would be of interest to know how closely these reflect the relative abundances of these species in the freshwater reaches of the Thames immediately upstream of Teddington and at sites further inland.

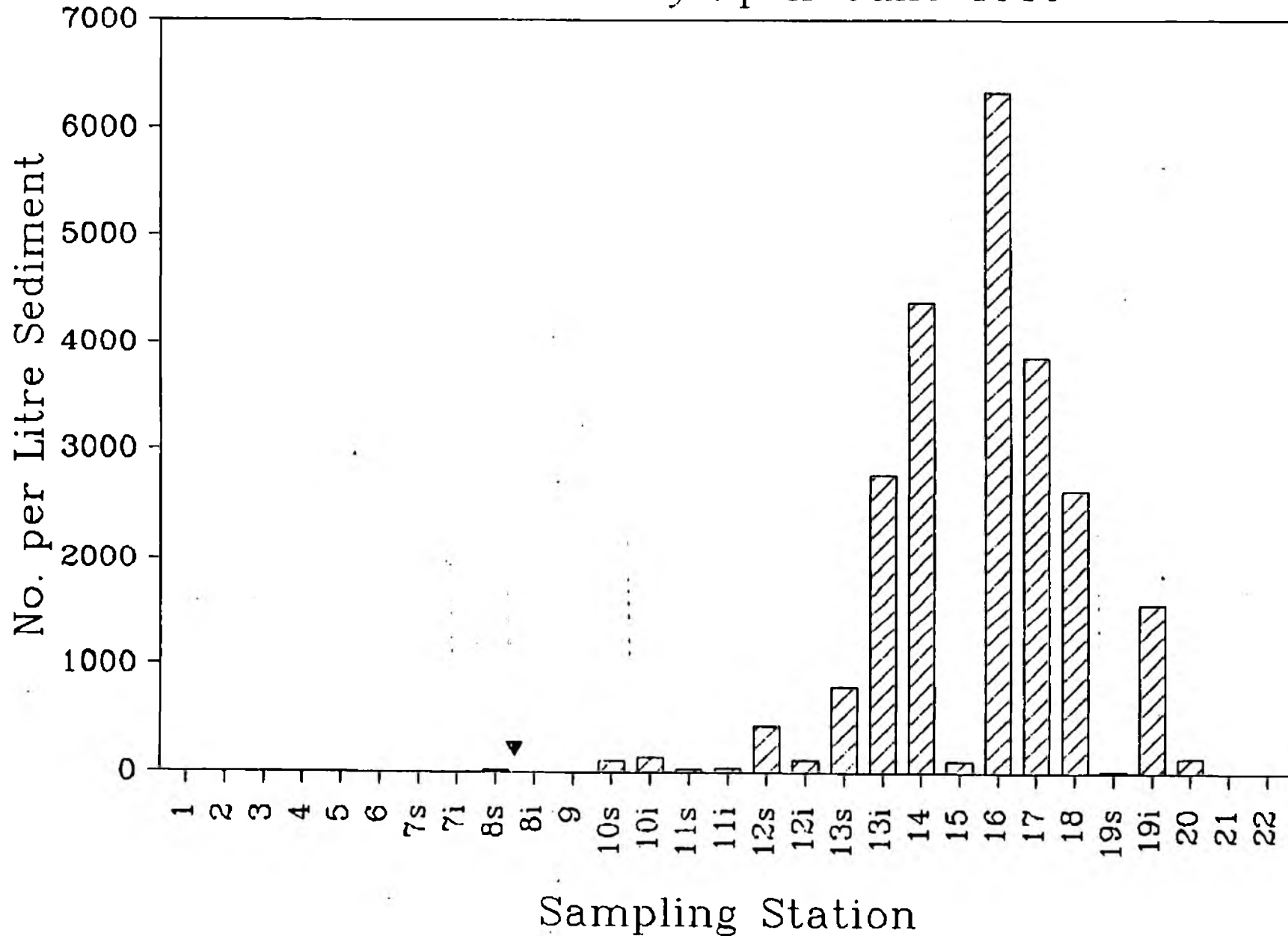
Low salinity and estuarine nematode species were well represented; again the species were typical of estuaries of southern England (Warwick, 1971; see also Capstick, 1959). Amongst the Enoplida, the larger predatory species, *Adoncholaimus thalassophygas* and its less common relative, *A. fuscus*, are archetypal estuarine nematodes and are commonly found in low salinity littoral sediments. These are type 2B species (see Appendix I; Section 5) with a large buccal cavity armed with three teeth of differing sizes. Along with many other species belonging to the family Oncholaimidae, they are probably K-strategists with populations taking several years to establish fully. *Anoplostoma viviparum* has been recorded from several estuaries in Britain (Platt and Warwick, 1983; Trett, pers. obsvn.), mostly from intertidal sites, although this might reflect the ease of sampling of these localities rather than preferred habitats. In the present survey, whilst highest densities of *A. viviparum* were recorded at intertidal sites, specimens were found in subtidal samples. *A. viviparum* is a type 1B species (see Appendix I; Section 5) and possesses a large unarmed buccal cavity. As such, it is held to be a non-selective detritivore although Thun (reported in Bouwman, 1983) suggests that they feed selectively on Protozoa.

Amongst the Chromadorida, estuarine/low salinity species include *Dichromadora geophila*. This species was largely replaced by another estuarine species in the same genus, *Dichromadora cephalata*, towards the mouth of the estuary. The latter is usually found in higher salinity habitats whereas *D. geophila* is capable of exploiting exceptionally low salinity conditions including barely brackish pools. Both are type 2A species (see Appendix I; Section 5) and are either diatomivorous or selective epigrowth feeders browsing microflora from the surfaces of sand grains. Densities of *D. geophila* were generally low (ca. 100 litre⁻¹). However, exceptional densities were occasionally observed (e.g. 28,732 litre⁻¹ at the intertidal Crossness sewage treatment works sampling site (10i), 3rd. quarter 1989). The species was also uniformly distributed throughout the upper estuary sampling sites. In contrast, densities of *D. cephalata* were more variable, ranging from 9 litre⁻¹ (3rd quarter 1989; Gravesend subtidal site (13s)) to 8,932 litre⁻¹ (4th. quarter 1989; Canvey Beach (16)). This is in keeping with observations made in other European estuaries (Trett pers. obsvn.; see also Riemann, 1966; Bouwman, 1983; Warwick, 1971; Newell *et al.*, 1989a).

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Figure 2. The percentage distribution of the population of *Sabatieria punctata* in the Thames Estuary, April - June 1989. Arrow denotes furthest upstream station at which *S. punctata* was observed.

Densities of *Sabatieria punctata* Thames Estuary April-June 1989



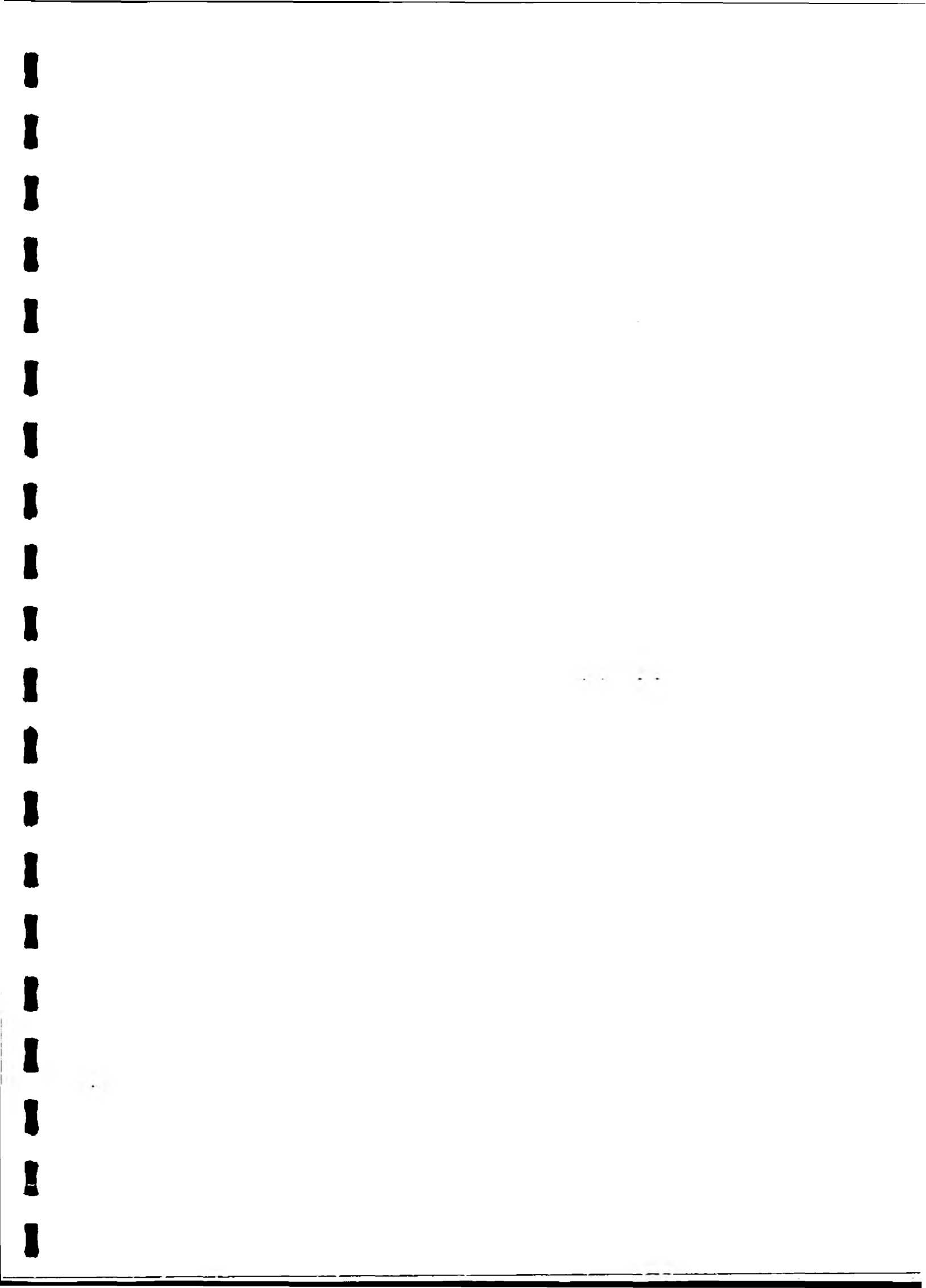
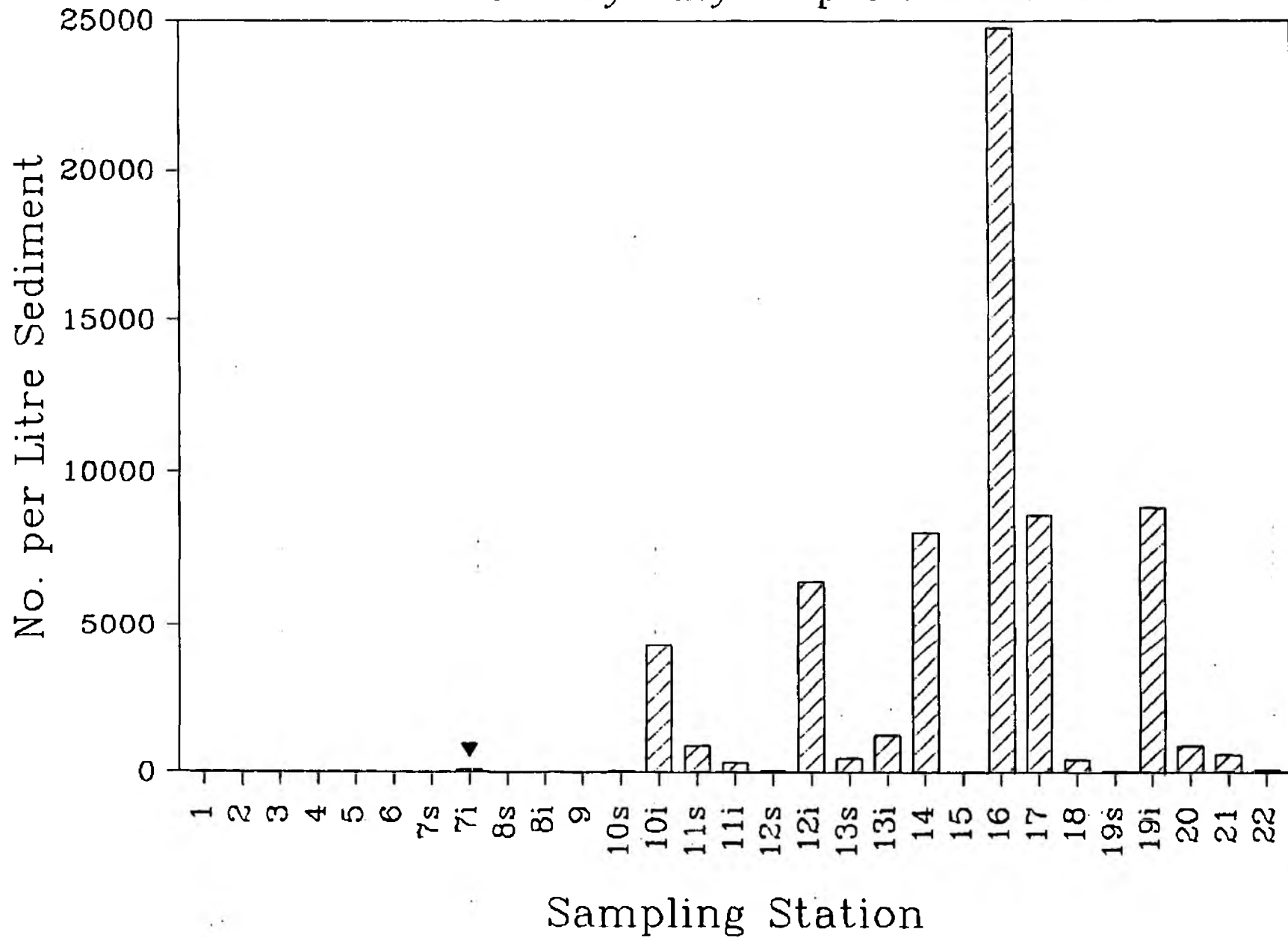


Figure 3. The percentage distribution of the population of *Sabatieria punctata* in the Thames Estuary, July - September 1989. Arrow denotes furthest upstream station at which *S. punctata* was observed.

Densities of *Sabatieria punctata* Thames Estuary July-September 1989



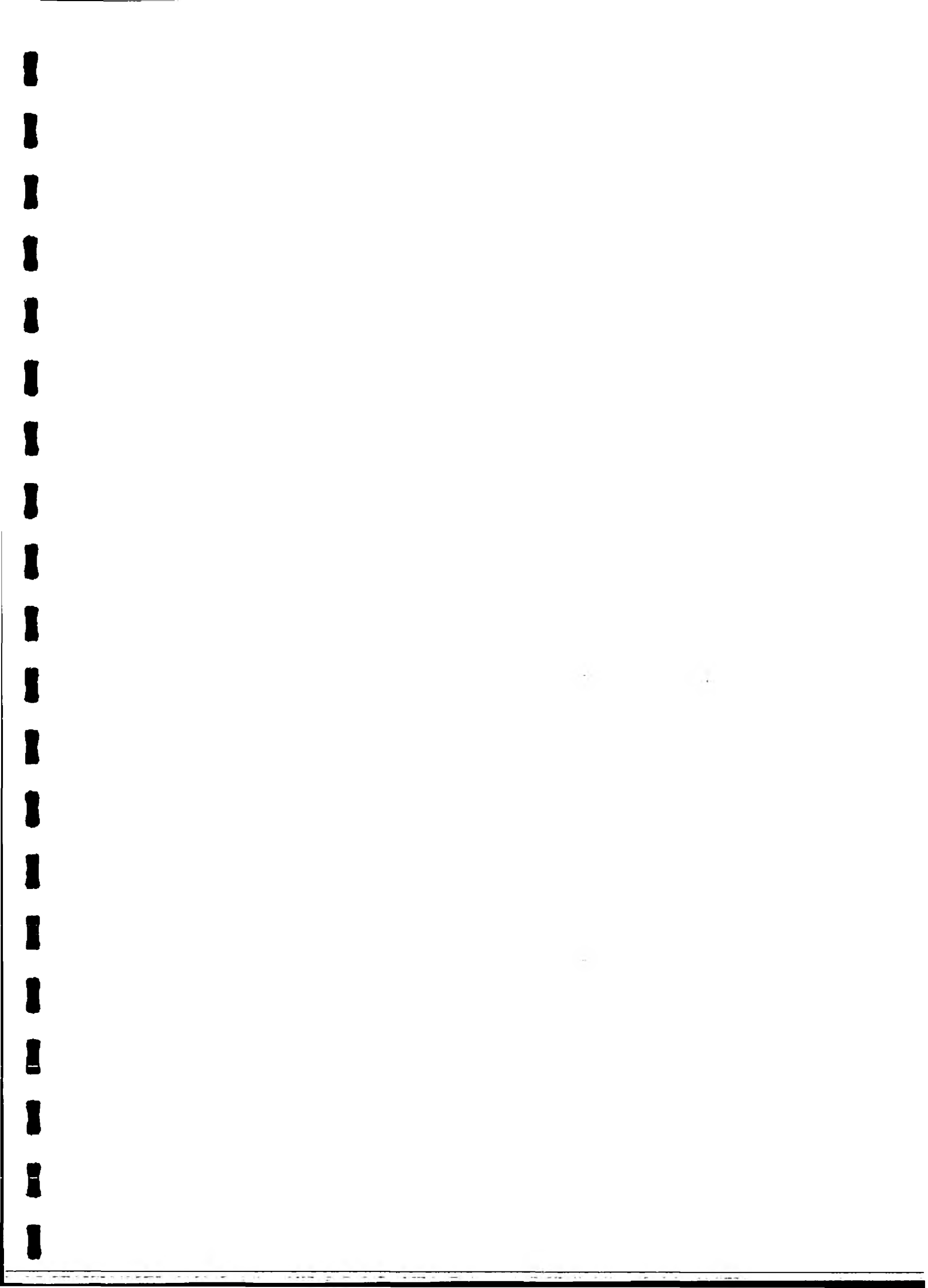


Figure 4. The percentage distribution of the population of *Sabatieria punctata* in the Thames Estuary, October - December 1989. Arrow denotes furthest upstream station at which *S. punctata* was observed.

Densities of *Sabatieria punctata*

Thames Estuary October–December 1989

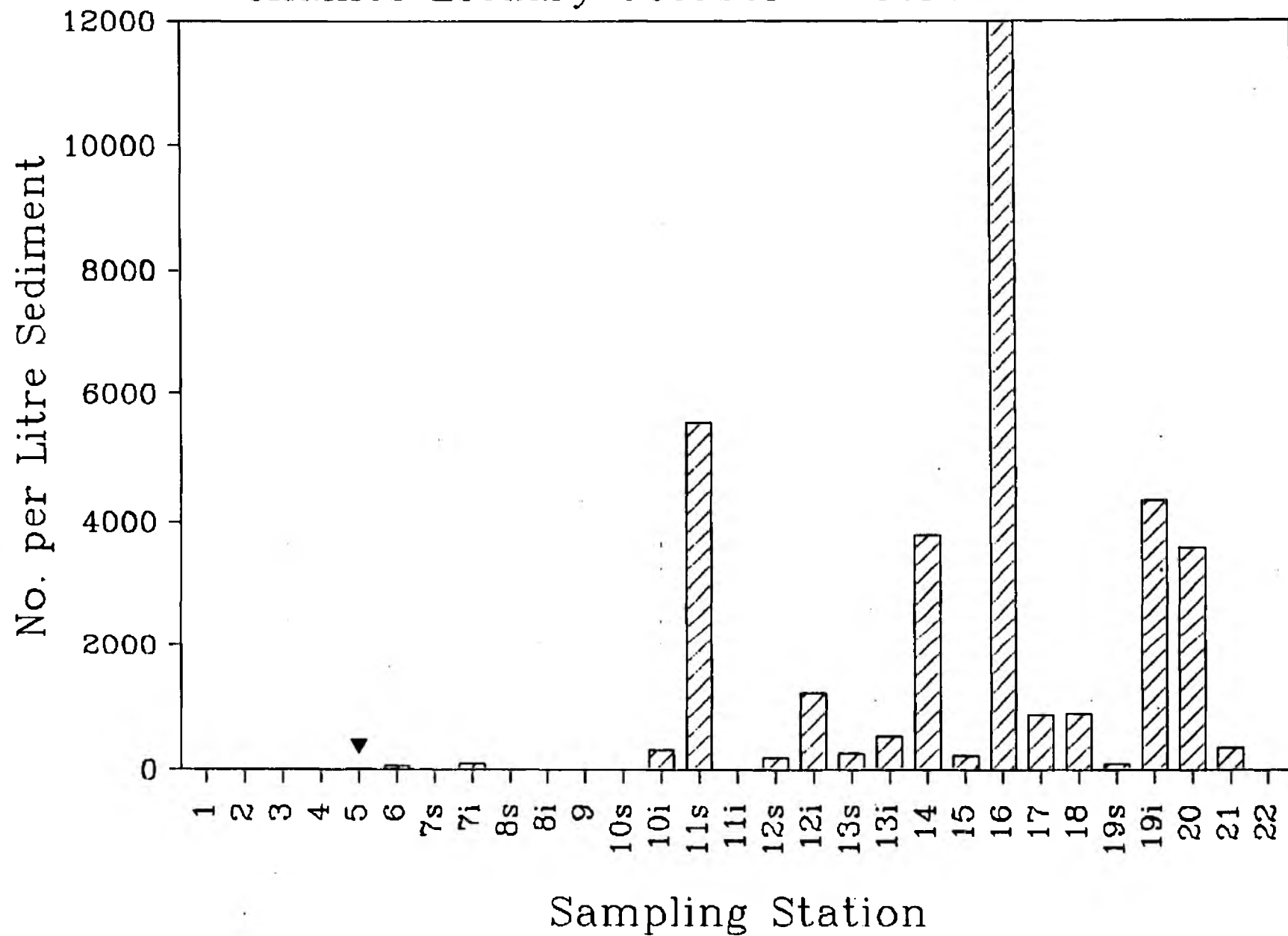
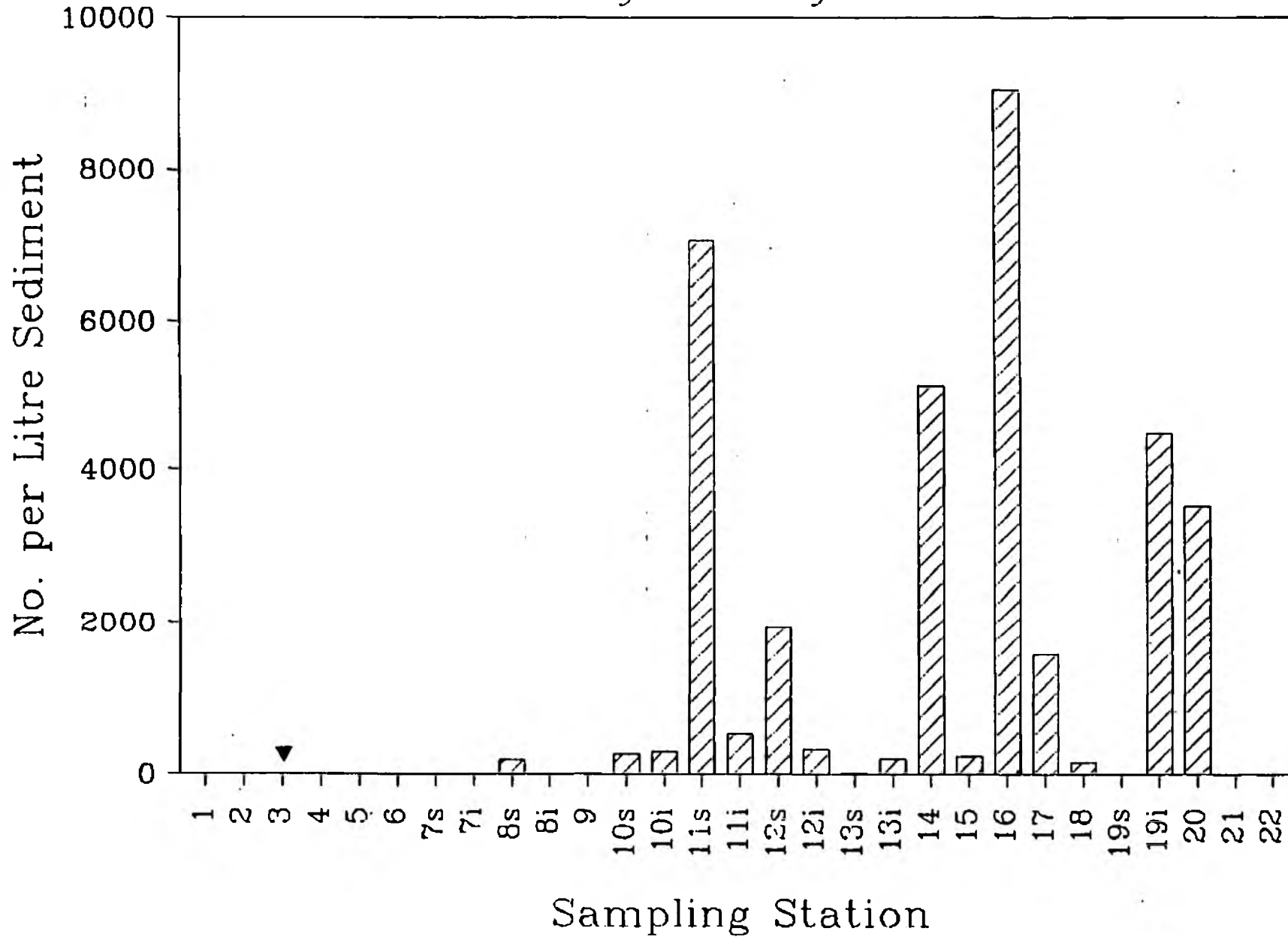




Figure 5. The percentage distribution of the population of *Sabatieria punctata* in the Thames Estuary, January - March 1990. Arrow denotes furthest upstream station at which *S. punctata* was observed.

Densities of *Sabatieria punctata* Thames Estuary January–March 1990





in each of the surveys. As with the Nematoda, the high species richness probably reflects the large area encompassed by the survey and the range of salinity regimes to which the sites sampled are subjected. The assemblages present at each station are considered in Section II below.

Apart from their distribution, comparatively little is known of the biology of the harpacticoid species found in the Thames samples. Several are known to be freshwater species accounting for their dominance at the Teddington end of the Estuary in the first survey (2nd. quarter 1989; see Appendix I; Section 2). For example, *Epactophanes richardi* is usually found in freshwater and semi-terrestrial habitats such as *Sphagnum* bogs or permanently wet leaf litter along banks of rivers. Similarly, *Bryocamptus ?praegeri*, *Canthocamptus* species, *Elaphoidella gracilis* and *Moraria* species are all usually associated with freshwater habitats such as lakes or slower flowing rivers. It is of interest to note that few freshwater species were found in the 2nd., 3rd. and 4th. surveys (see III below). Most of these species are epibenthic and feed in the upper layers of the sediment. They are probably detritivores but this is not known for all the species named above. In nearly all cases they are highly mobile and, in the case of lake and still water species, may undertake short dispersal swims up into the water column at night.

The same may be true of the diosaccid harpacticoids which are mostly marine and estuarine species although there are some notable exceptions. *Schizopera* species, including *S. clandestina*, prefer brackish waters and some species have been reported from freshwater localities (Lang, 1948). Earlier studies of the Thames revealed *Schizopera compacta* in and amongst fine red-brown algae and in the byssal holdfasts of *Mytilus edulis* (Feil, pers. obsvn.). *Stenhelia palustris* is a common epibenthic diosaccid of estuaries (Green, 1968) and has characteristic nauplius larvae which were observed at low densities in many of the samples examined. *S. palustris* is known to occur in muddy intertidal sites where there may be considerable variations in temperature and salinity. We have found that this species often builds small temporary tubes, up to 4 mm long, into the sediment. This suggests that it might feed in a similar manner to certain species of *Corophium* that also construct burrows, namely a combination of filter and deposit feeding (Hughes, pers. comm.).

Diosaccids and canthocamptids have well developed, protruding swimming limbs. In contrast the body shape of ectinosomatids is streamline. The antennae are reduced and the limbs are directed posteriorly and held against the body. This appears to facilitate burrowing. In the present survey their distribution demonstrates their preference for higher salinity estuarine sediments and correlates with coarser grained sediment types. Of the Ectinosomatidae, *Halectinosoma* species are commonly found in subtidal marine sediments. The furthest into the estuary that *Halectinosoma* species penetrated was the subtidal sampling site at Gravesend (13s) in the 1st. quarter 1990 (*H. curticorne*). This probably represents the extreme end of its salinity tolerance range. Densities of this species increased towards the mouth of the estuary and were accompanied by the appearance of other ectinosomatid species such as *Ectinosoma melaniceps*.

In the Ameiridae and Laophontidae, the limbs are reduced and the species are almost vermiform. Members of these families are much less mobile than the other harpacticoid species and adopt a truly interstitial existence. In *Leptomesochra macintoshi* the reduction



in limbs is more pronounced than in *Laophonte ?denticornis*. The interstitial species are more sensitive to environmental conditions than epibenthic species and their distribution within an estuary usually reflects this fact. It is of note that the two species were largely restricted to the coarser grained sediments at the mouth of the estuary (see Southend (19s), Shoeburyness East (21) and Sea Reach No. 2 Buoy (22)).

iii. Acari

The total acarine community of the Thames Estuary observed over the 4 surveys was found to comprise 6 taxa from 4 families (Appendix I; Section 1). The different assemblages of Acari at each station are considered in Section II below. Of the species recorded, an oribatid species was the most commonly observed, occurring at 16 of the 29 stations sampled over the 12 month period sampled (Appendix I; Section 2). Oribatid mites are commonly present in sediment samples from estuarine and nearshore marine habitats. They prefer terrestrial rather than aquatic habitats and probably originate from decaying organic material along the strandline. Accordingly their distribution within a survey area such as the estuary may be of little direct biological significance. However, their densities do appear to vary in relation to the amounts of macrophyte or macroalgal detritus that is present. The single bdellid species is not thought to be an aquatic species (Green, pers. comm.) and has probably arisen from leaf litter in the Thames catchment. In contrast the cryptostigmatid mites observed are all true aquatic species. The hydracarine found at Teddington (1), 1st. quarter 1989, is a true freshwater species and is indicative of clean, unpolluted conditions. Halacaridae can occur in freshwater, but the species observed are marine species that will happily tolerate reduced salinities. *Copidognathus rhodostigma* is a common species from around the British coast. It has been recorded in samples down to a depth of 60 m but occurs in highest densities in algae just below the low water springtide levels. Personal observations (Trett and Feil) suggest that it is also common in the byssal attachments of *Mytilus edulis* and amongst attached weed on piers in the outer Thames Estuary.

Copidognathus dentatus is an interesting species. Until recently its known distribution was the Adriatic coast of Yugoslavia only. Specimens were then found in a meiofaunal survey of the Humber Estuary, off Grimsby, and at Westgate-on-Sea on the North Kent coast (Green and McQuitty, 1987). It is usually found in the lower intertidal zone and shallow subtidal sediments amongst stones and algae. In the present surveys it was found at low densities at Blythe Sands (15), Canvey Beach (16), Chapman Buoy (18), Southend subtidal (19s) and Sea Reach No. 2 Buoy (22). From the biological point of view, it would be of interest to know something of the macrofauna recorded at these stations.

Remaining unidentified species comprised mostly halacarid nymphs which might well belong to *C. rhodostigma* or *C. dentatus*. The densities of halacarids in the middle reaches of the Estuary are smaller than those reported in other estuaries. Trett (in Newell *et al.*, 1989a), for example, reported 6 acarine species at densities of up to 109 litre⁻¹ sediment in the Humber Estuary.



b. Other Meiofaunal Groups

Qualitative data on the occurrence of other groups of meiofauna observed in the Thames sediment samples are summarised in Appendix I; Section 3, Tables 1 to 8 and considered at each sampling station in Section II below. The tests of rhizopod amoebae (predominantly *Centropyxis* species) were very abundant at stations from Teddington (1) to Cadogan Pier (4). Other records seaward of the pier relate to observations of single tests. Measuring up to 400 μm in length, the amoebae prefer freshwater and are intolerant of polluted conditions. Marine ciliate species were observed in comparatively large numbers at 3 stations at the mouth of the estuary at densities up to 500 litre⁻¹ sediment (4th. quarter, 1989). In common with the Rhizopoda, these do not tolerate polluted conditions. The distribution of tests of Foraminifera reflects their preference for higher salinity waters with high densities of planispiral *Elphidium* species and multilocular *Brazilliana* species present at stations in the outer Estuary but relatively few specimens occurring upstream of West Thurrock. Single specimens of the large, unilocular marine genus *Lagena* were occasionally found at sampling stations near the mouth of the estuary.

Tardigrada were represented by three species. *Macrobotus dispar* and *Echiniscus* species were present at stations between Teddington (1) and Woolwich (8). These are predominantly freshwater species that often occur in fresh-brackish water sediment samples. Stations between Canvey Beach (16) and Sea Reach No. 2 Buoy (22) possessed a 'primitive' marine species, *Batillipes mirus*. Each of these species of tardigrade is commonly found in British estuaries in southern England (Morgan and King, 1976) and *B. mirus* may be distributed widely throughout British coastal waters (Forster and Trett, pers. obsvn.).

Kinorhyncha and Gastrotricha have been recorded in European estuaries but are usually restricted to clean, unpolluted sites and the higher salinity sediments. In the present surveys, both kinorhynchs and gastrotrichs were observed, represented by a *Pycnophyes* species and a *Chaetonotus* species, respectively. However, these were present at the mouth of the estuary (e.g. Grain Flats (20), Shoeburyness East (21) and Sea Reach No. 2 Buoy (22)). In contrast, Turbellaria were recorded at several stations in the estuary and, as many of these are stenohaline it is likely that several species are present. Taxonomy of this group is particularly difficult and requires fresh, anaesthetised specimens of sexually mature adults. The same applies to the closely related Gnathostomulida. However, these were much rarer and were observed at Shoeburyness East (21) in the 2nd quarter 1989 and Sea Reach No. 2 Buoy (22) in the 1st. quarter 1990 only.

c. Non-meiofaunal Groups

Qualitative observations made on the presence/absence of non-meiofaunal groups present in the sediment samples examined are presented in Appendix I; Section 4, Tables 1 to 8. Groups present at individual stations are described in Section II below. Diatoms and their empty frustules were almost ubiquitous in the Thames Estuary sediments. Towards Teddington, these were predominantly freshwater pennate species which were replaced by



increasing densities of marine and estuarine centric species (*Trigonium* and *Coscinodiscus* species). *Nitzschia* species were also present in many of the higher salinity sediments (stations to the east of West Thurrock). In sediments from many of the upper estuarine sampling stations, loricae of rotifers were present (mostly *Keratella* species). Many of these may have been transported to the higher salinity sites from the upper reaches of the Thames Tideway and from the catchment although some specimens of *Brachionus* species were seen. The latter are tolerant of higher salinity conditions and can become abundant in estuarine systems (Green pers. comm.). The chironomid larvae observed were not identified to species. These were found in samples from Teddington (1) and Kew (2) only during the 2nd. quarter 1989 but were more widespread in subsequent surveys, extending as far as London Bridge (5) in the 1st. quarter, 1990. At Teddington and Kew the numbers of larvae were high with densities of up to 200 litre⁻¹ occurring in the 3rd. quarter 1989. Cladocerans showed a similar distribution to the chironomids with *Bosmina* and *Daphnia* species present at low densities at both Teddington (1) and Kew (2). Unidentified chydorid cladocerans were also commonly observed in the sediments from these stations.

The distribution of oligochaete annelid juveniles was much as expected. Naidid and tubificid species were abundant towards Teddington. These became increasingly rare towards the mouth of the Estuary. However, the distribution of polychaete neochaete larvae was much wider than expected and extended almost throughout the estuary in the 2nd. quarter 1989 and the 1st. quarter 1990. The neochaete larvae resembled those of spionids and, to the seaward end, nereid species. *Polydora* species are known to occur in large numbers in the Estuary, usually in mud tubes around the bases of epilithic hydroids and bryozoans (Hughes, pers. comm.; observations made on epifauna on rocks collected with a Baird dredge). It is possible that the neochaetes and newly metamorphosed individuals at the westerly end of the estuary belong to the freshwater polychaete genus *Manayunkia* although British records of this are uncertain.

Comparatively few newly settled mollusc larvae were observed in any of the surveys. Low densities of littorinid gastropod egg capsules were observed at Canvey Beach (16), Allhallows (17) and Southend (19i) in the 2nd quarter 1989. Whether these had arisen from local gastropod populations is uncertain. A survey of Thames shores undertaken in 1987 (Feil and Newell pers. comm.) failed to find littorinids for use in silver determinations between Dartford and Cliffe Creek. Low densities of newly settled bivalve larvae were also observed in sediments downstream of Gravesend. These are thought to belong to tellinid species.

The calanoid copepod, *Eurytemora affinis* is not a benthic species and is usually a member of the zooplankton present in the water column. It is a typical estuarine species and commonly present in considerable numbers in plankton samples taken from middle reaches of the Thames by CRAB staff. Care has to be taken to distinguish this species from *Eurytemora velox* which occurs in the lower salinity waters of the London docks but can enter the estuary during operation of locks (e.g. at St. Katherine's Dock (Green and Hutchinson, pers. comm.)). Despite its widespread occurrence within the estuary, its presence at Teddington (1) and Shoeburyness East (21) is somewhat extreme and may have resulted from tidal or avian transportation.



II. Meiofaunal Summaries

This section provides a more detailed description of the fauna present at each sampling site reported in Appendix I; Section 2 (q.v.) with additional comments on seasonal variation.

Station 1, Teddington

The muddy sediment between the coarse stones at this station supported a mixed community of predominantly freshwater meiofaunal species, typical of those that we have observed in surveys of freshwater ecosystems in lower course rivers in southern England. The complement of nematode species present varied in composition between the surveys and, although a cumulative total of 37 species was observed, a maximum of 26 species was observed in any one survey (4th. survey). The lowest number of nematode species were recorded in the 3rd. survey 1989 (7 species). Plant-parasitic members of the Tylenchida and Dorylaimida were commonly observed at this site although their relative abundance declined in the samples collected after the 1st. survey. These are a common component of lake and riverine meiofaunal assemblages and probably feed on the root systems of marginal plants. The 1B:2A feeding-type ratios indicate that detritivorous species were relatively more important at this site in the latter half of the year than the selective epigrowth and diatomivorous species. This results from a decrease in type 2A species rather than an increase in type 1B species and might reflect a change in the diatom populations or a change in the quality or nature of bacterial and algal epigrowth communities.

Dominance-diversity characteristics of the nematode assemblages at this site are illustrated in Figure 7. Between April and June, *Tobrilus gracilis* was the dominant species accounting for just less than 26% of the individuals in an assemblage of 12 species. In the following 2 surveys the dominance of *T. gracilis* increased to approximately 50% and was highest in the 3rd. survey (53%). In the final survey species richness had risen from 7 in the previous quarter to 26 taxa with a concomitant decline in dominance as *Paracyatholaimus intermedius*, a low salinity type 2A cyatholaimid, superseded *T. gracilis* as the dominant species.

The most diverse harpacticoid copepod assemblage was observed during the 1st. survey (April - June 1989). This assemblage comprised 8 species most of which are associated with freshwater habitats. Harpacticoids were not observed in the 2nd. survey and few individuals only were present in the remaining surveys. This significant change in the harpacticoid community may have resulted from changes in the flow rates upstream of the lock owing to drier weather and/or changes in the ambient salinity. Acarina were observed only in the 1st. quarter 1990 and included an oribatid and bdellid species.

In each of the surveys, other invertebrate species observed in the meiofaunal samples from this station were typical of low salinity or freshwater conditions. Densities of chironomid larvae were highest in the late summer and winter samples and lowest in the spring. This most probably relates to the emergence of adults in spring and progressive larval recruitment through the summer months.

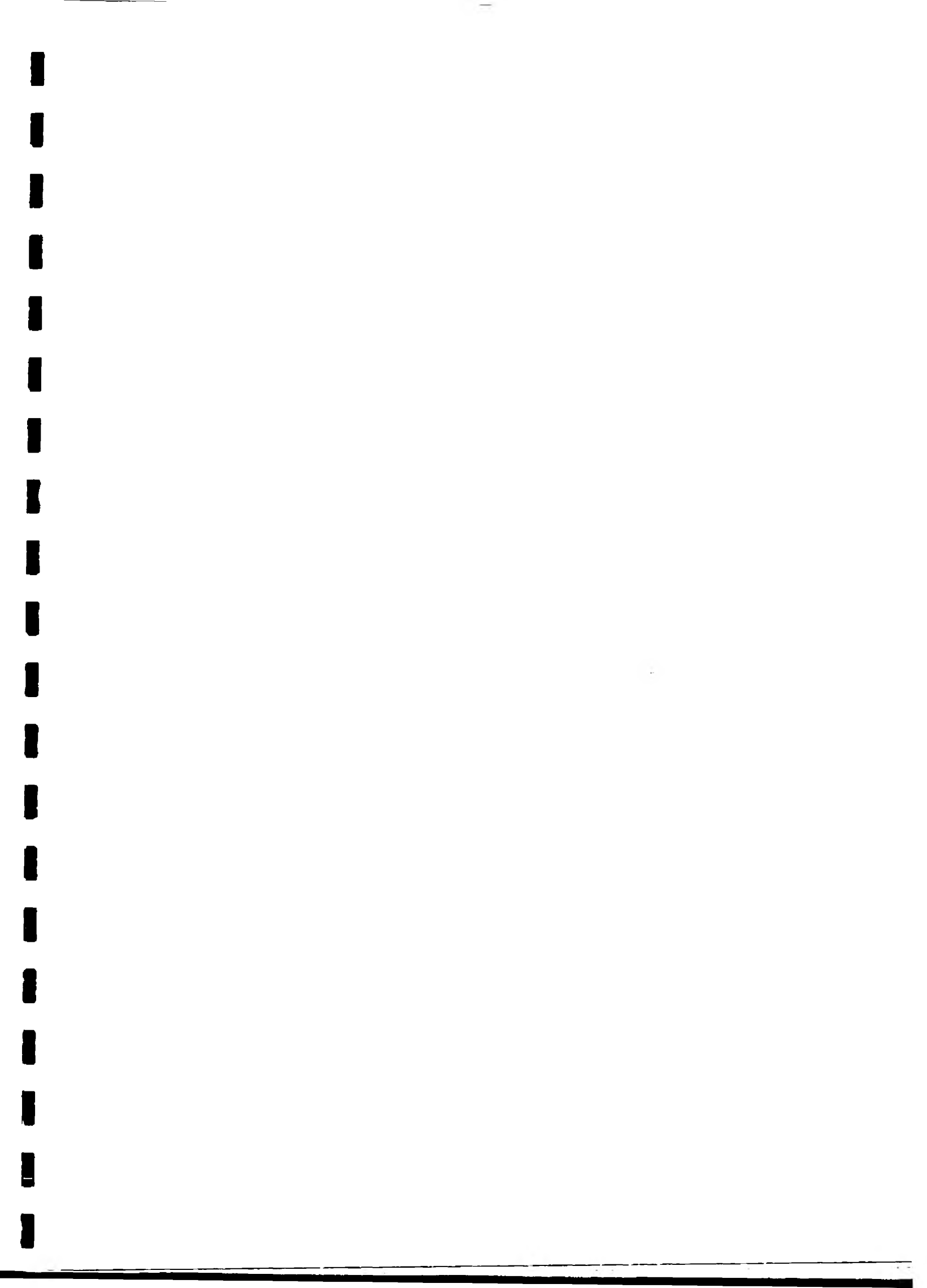
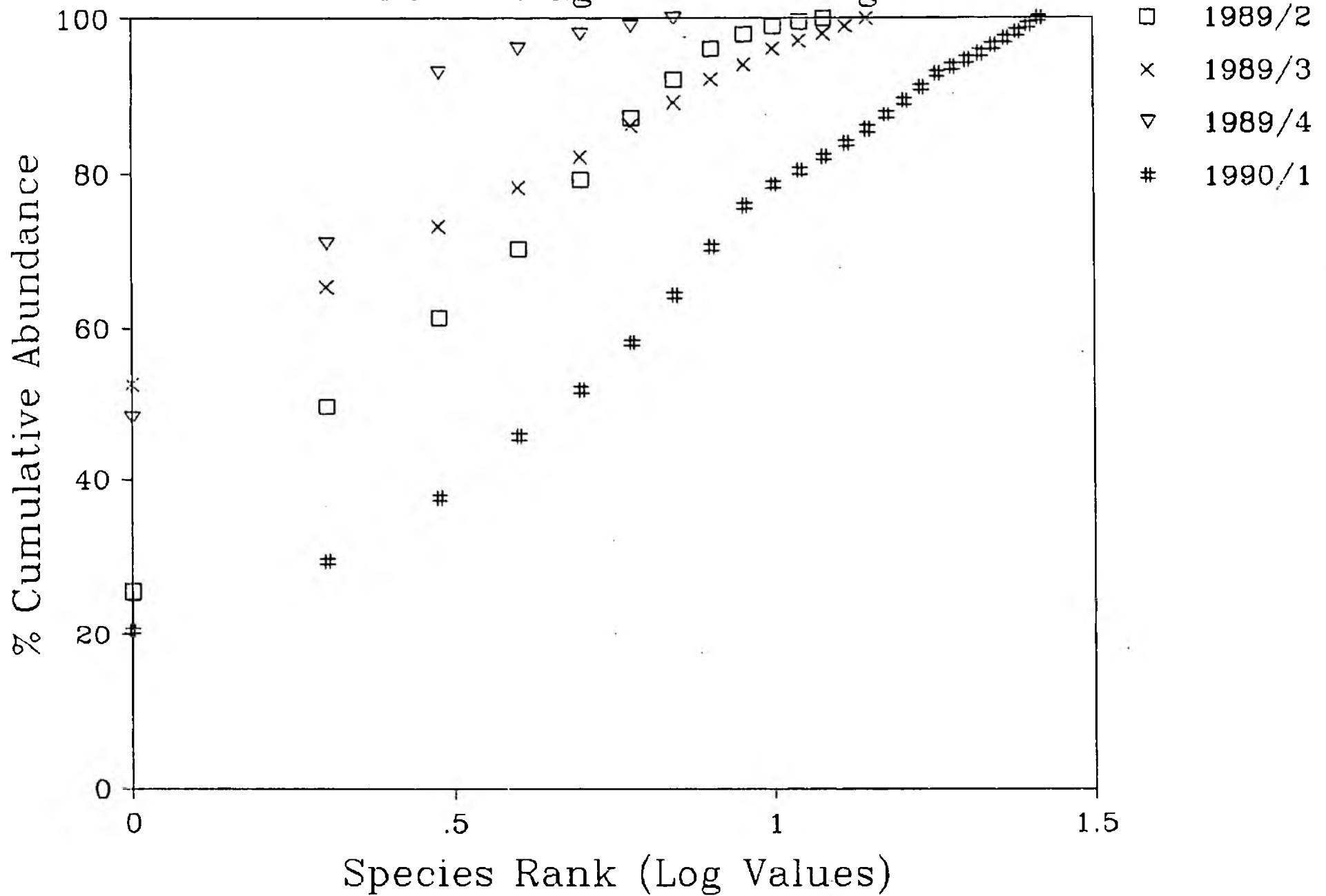


Figure 7. The dominance:diversity characteristics of the nematode assemblages observed at the Teddington sampling station between April 1989 and March 1990.

k-Dominance Curves for Nematode Assemblages at Teddington





Station 2, Kew

Meiofauna at Kew was similar in many respects to that observed at Teddington. *Tobrilus gracilis* was a key component of the nematode assemblages which, again, comprised freshwater nematodes including several plant-parasitic tylenchid and dorylaimid species. Variability in composition of the nematode assemblages was also high and of a total of 37 species observed throughout the year, 18 was the maximum number observed in any one survey (4th. survey). The number of species and the densities observed, however, were more uniform than at Teddington. Dominance-diversity plots (Figure 8) also showed that dominance in the 2nd. survey exceeded that recorded in any of the other surveys (ca. 80%; *T. gracilis*). This correlated with the lowest number of species observed (11 taxa). Feeding type ratios at this site indicated a balance in favour of non-selective detritivores.

Fewer harpacticoid species were observed at Kew than at Teddington and none was observed in the October-December samples. The species that were present included freshwater groups and an epibenthic estuarine species as well as diosaccid copepodites. As at Teddington, Acarina were observed only in the January - March samples and comprised oribatid and bdellid species.

Ostracod, tardigrade and turbellarian species along with the other invertebrates present in the samples were all in keeping with a low salinity-freshwater locality

Station 3, Hammersmith Bridge

A total of 44 nematode species were observed at this site. These included several estuarine and low salinity species such as *Oncholaimus campyloceroides*, *Dichromadora geophila*, *Axonolaimus paraspinosus* and a *Metachromadora* species. Hammersmith Bridge also marked the most easterly record of the common estuarine species *Sabatieria punctata* which was observed as a single specimen in the January - March 1990 samples. Several freshwater species persisted at this station including certain plant-parasitic species (e.g. Dorylaimid species 1, *Hirschmanniella* species and *Criconemoides* species) and several rhabditids (e.g. *Mononchoides striatus*, *Butlerius butleri* and the *Rhabditis* and rhabditid species). Again the feeding type ratios indicated a predominance of non-selective deposit feeders but the lower ratios throughout the year confirmed that the relative densities of diatomivorous and epigrowth feeding species were higher than at either Teddington or Kew. Inspection of the species lists shows these type 2A species to be principally chromadorids and cyatholaimids more commonly found in coarse grained, low to moderately low salinity sediments.

Very few harpacticoid copepods were observed at this station. Indeed, 9 *Attheyella* species litre⁻¹ were the only record (4th. survey). The genus *Attheyella* is more usually associated with the margins of lakes and rivers and, in estuaries, is usually restricted to the upper reaches. We have also observed this species in a survey of the upper tidal reaches of the Medway (Feil and Trett, pers. obsvn.). Amongst the other invertebrate groups

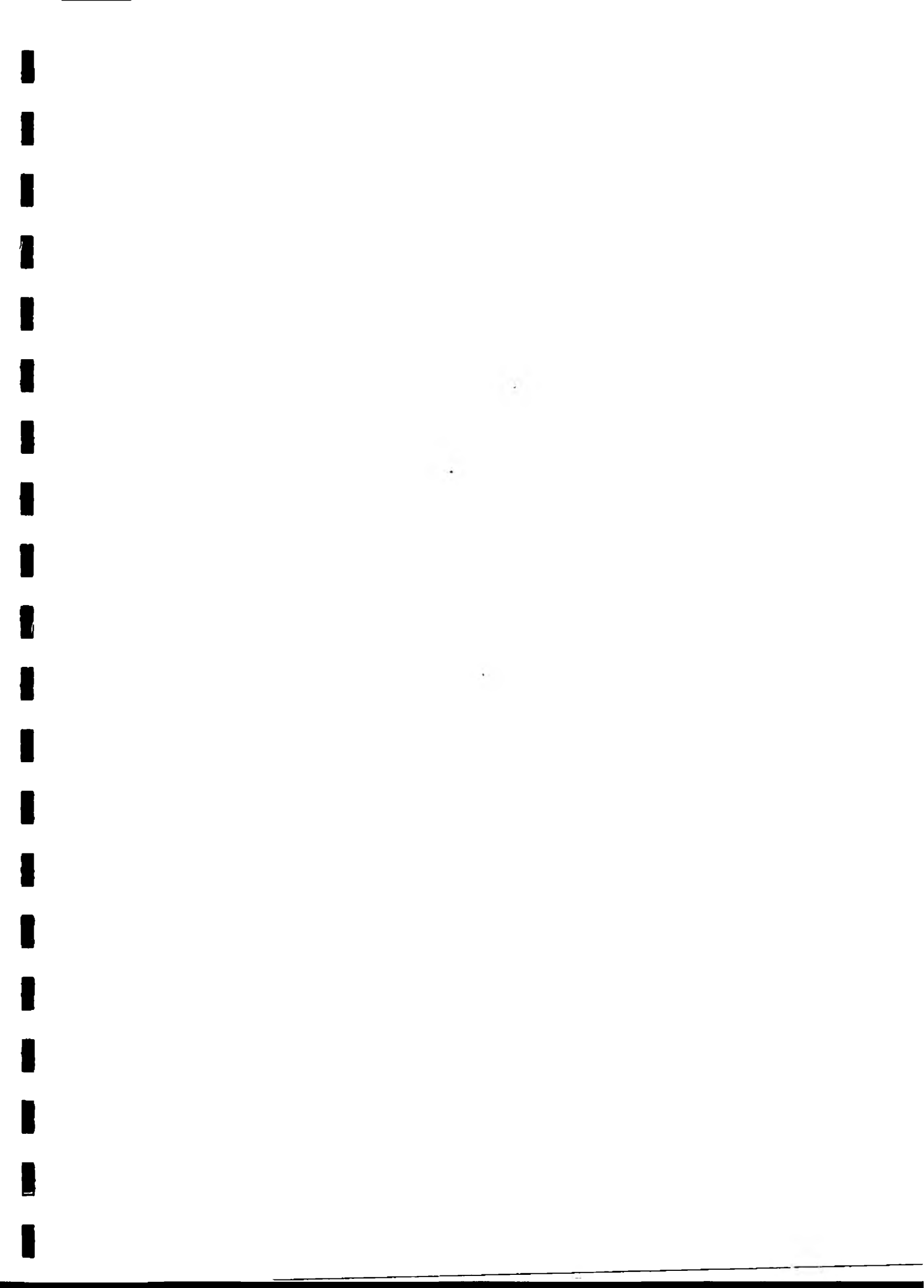
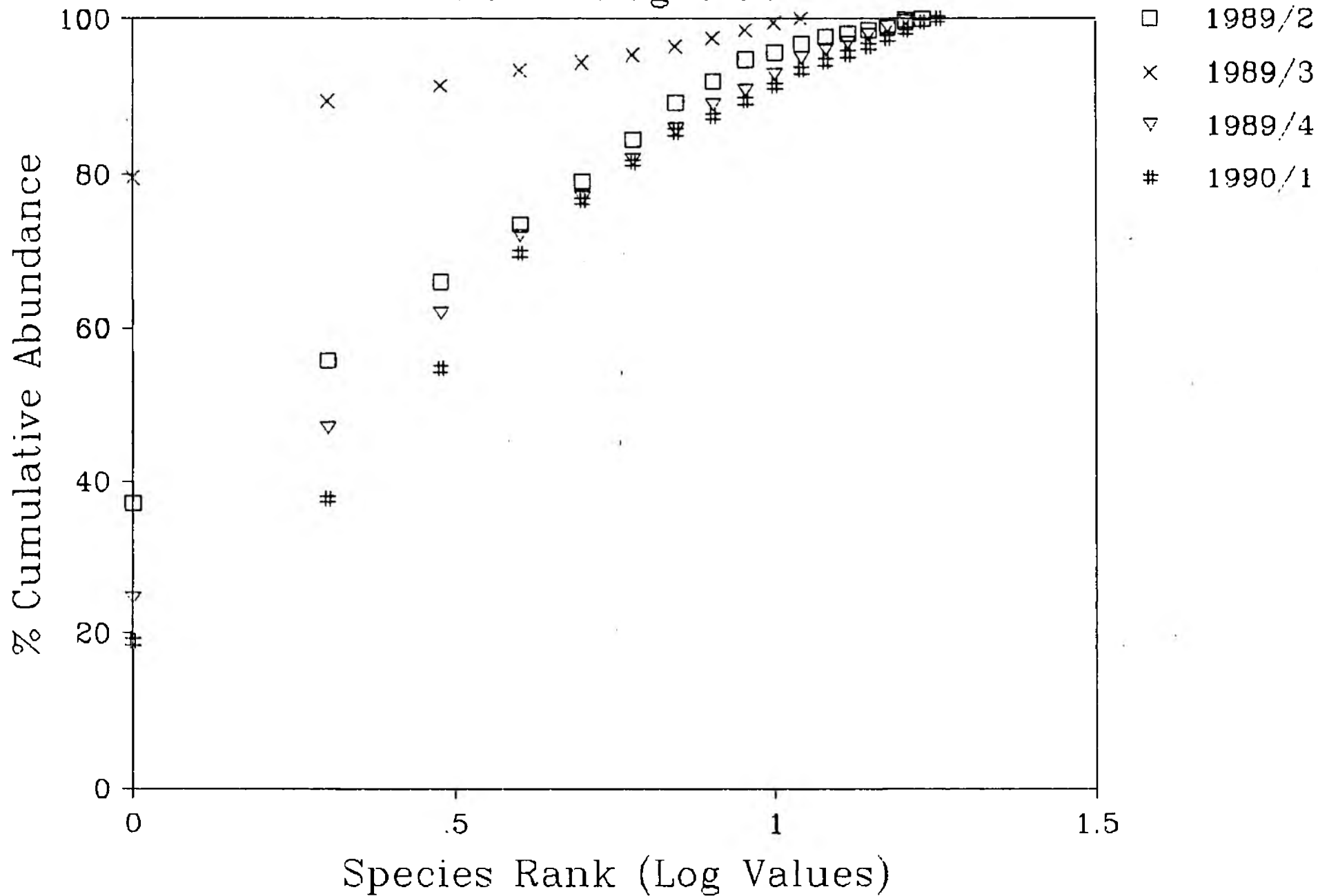


Figure 8. The dominance:diversity characteristics of the nematode assemblages observed at the Kew sampling station between April 1989 and March 1990.

k-Dominance Curves for Nematode

Assemblages at Kew





present in the samples examined, oligochaetes and rotifers were found throughout the year. Chironomid larvae were abundant in each of the surveys except the 1st. survey (April - June) when none was observed. The frustules of marine centric diatoms were present amongst those of freshwater pennate species indicating the proximity of the upper estuary limit of transportation of finer particulate material.

Station 4, Cadogan Pier

The 47 species of nematode observed at this site during the year April 1989 - March 1990 were a mixture of fresher water and true estuarine species. There was some evidence of colonisation by lower estuary and marine species such as *Innocuonema* species and *Tripylodes gracilis* during the middle surveys (July - December). This correlated with the observation of muddier sediments at Cadogan Pier and may have related to reduced flow during these periods (see also Stations 20 and 22). In the April 1989 and January 1990 surveys the sediment contained noticeable sand fractions. The 1B:2A feeding type ratio was highest in the second survey (July - September 1989) largely as a result of the *Daptonema* species (*D. furcata*), a non-selective detritivore, that became dominant (ca. 65%) in the finer sediments. This is another species that would be expected to occur in higher salinity conditions and its presence here probably reflects a realised as opposed to a preferred niche.

Harpacticoid species were observed in the final survey only and even here were restricted to a single species of *Microarthridion* at low density (46 litre⁻¹). This genus comprises small epibenthic species that appear to be "completely euryhaline" (Gurney in Lang, 1948). Acari were not observed.

The most diverse assemblage of other invertebrate phyla present in the Cadogan Pier samples was observed in the final survey. Oligochaetes were the only invertebrate group to be observed in each of the 4 surveys as were the diatoms amongst the algae.

Station 5, South Bank Centre

This station was characterised essentially by sandy sediments and sandy sediment infauna. As at Cadogan Pier, the mid-survey samples (July - December 1989) contained higher silt-clay fractions than in either the first or last samples. In general, sandy sediment nematode assemblages would be expected to be richer in species than those present in fine sediments. However, a total of 27 species only was observed in the 4 surveys, all at comparatively low densities (range: 102 to 354 litre⁻¹). The relatively high densities of cyatholaimid and/or chromadorid species and low abundances of non-selective detritivores accounted for the low feeding type ratios in the 1st. and 3rd. surveys. However, selective epigrowth (epipsammic) and diatomivorous species (type 2A) were not observed in the 2nd. or final surveys, undermining comparison of the trophic assemblages.



The trend for reduction in fresh and brackish water species with increasing distance down the estuary continued to be observed and relative densities of true estuarine were higher than at any of the preceding sampling stations. Single individuals of *Ptycholaimellus ponticus*, *Odontophora setosa* and *Chromadora macrolaima* were observed at this site in the 3rd. survey marking the most westerly occurrence of these essentially outer estuarine, type 2A species.

Harpacticoid copepods were not observed in any of the samples from the South Bank Centre sampling station. Acari were limited to observations of single individuals of an oribatid species (1st. survey) and *Copidognathus dentatus* (final survey). It is of interest to note that the South Bank Centre may represent one of the lowest salinity sites at which *C. dentatus* has ever been recorded. This species usually inhabits much higher salinity sites either in the outer estuary or in coastal waters.

Turbellaria (indet.) were present in each sample taken from this site. Other groups, however, were comparatively poorly represented.

Station 6, London Bridge

In contrast to the South Bank Centre sampling site, the London Bridge site yielded a high nematode species count (cumulative total of 46 species). Total numbers of nematodes observed litre⁻¹ sediment were, with the exception of the 1st. survey, high with a maximum of 6,640 litre⁻¹ in the July - September 1989 sample. The species recorded here are commonly found in muddy substrates that contain at least some coarse-grained material in a low to moderate salinity environment. The freshwater catchment and low salinity upper reaches of the estuary were indicated by the presence of species such as *Plectus granulatus*, *Criconema* species, Dorylaimid species 2, *Tobrilus gracilis* and *Mononchoides striatus*. The marine influence is indicated by the presence of trace *Oxystomina asetosa*, *Deontolaimus* species, *Paracanthochus heterodontus*, and, possibly, *Axonolaimus paraspinosus*. The remaining species are all commonly observed in European mid- to upper estuarine samples. As in the Cadogan Pier samples, the 1B:2A feeding type ratio was highest in the second survey again as a direct result of increased densities of xyalid species (*Daptonema* species).

The harpacticoids were represented by a single specimen of a species of the freshwater genus *Bryocamptus*. These are commonly found in more acid waters associated with decaying leaf litter in woodland streams, *Sphagnum* bogs or, occasionally, subterranean streams. As such, it is likely that this species was transported to the London Bridge site from the Thames catchment. Specimens were also recorded at Teddington and Kew in the 3rd. and 4th. surveys, respectively.

Acari were not observed in any of the London Bridge samples. Other fauna included specimens of the calanoid copepod, *Eurytemora affinis*; this is a common species in the plankton of the Thames Estuary (Green pers. comm.) and frequently occurs as aberrant specimens in meiofaunal samples from European estuaries (Trett, Feil and Forster, pers.



obsvn.). It occurs in brackish waters only, unlike its close relative *Eurytemora velox* which can inhabit fresh and brackish water; *E. velox* occurs widely in the lower salinity Dockland waters and is an occasional inhabitant of the Thames Estuary zooplankton. Rotifers observed at London Bridge included a species of *Brachionus* which is again usually planktonic. Some species of this herbivorous rotifer are able to exploit low salinity waters and may be a characteristic component of certain estuaries in southern England.

Station 7, Greenwich

Subtidal and intertidal sampling sites were established at Greenwich, although the subtidal site was not assessed during the 1st. survey. In all cases, larger numbers of meiofaunal species were observed at the intertidal rather than the subtidal site; the cumulative species totals for nematodes being 48 and 29 species, respectively and, for harpacticoid copepods, 5 and 1 species, respectively. An oribatid species was recorded in the 1st. and final surveys of the intertidal site only. Similar differences between the inter- and subtidal meiofaunal assemblages have been noted in surveys of the Yare, Bure and Waveney (East Anglia; Trett pers. obsvn.) and studies of the Elbe (Germany; Riemann, 1966) and Ems Estuary (Netherlands/Germany; Bouwman, 1983). Factors that might be responsible for these differences include greater heterogeneity of the littoral habitat, elevated primary production, especially of benthic diatoms, increased oxygenation of sediments and mechanical disturbance by wave action.

Of the 29 species observed in the subtidal samples; 8 species only were not observed in the intertidal zone. With the exception of *Paroigrolaimellus bernensis* and a plectid species, which are common in low salinity or freshwater habitats, those unique to the subtidal site were either marine or outer estuary species. Densities of nematodes in the intertidal site were not always higher than the subtidal populations. However, intertidal nematode densities did rise to between 5 and 7 times those of the subtidal site in the mid-survey samples (July - December 1989).

Sufficient species were present at the intertidal site to enable dominance-diversity characteristics to be examined graphically (see Figure 9). Dominance was highest during the 2nd. survey (July - September 1989) when *Daptonema setosa* accounted for 76% of the nematode population. The dominance of this species declined to approximately 49% in the 4th. survey (October - December 1989) and 22% in the final survey (January - March 1990). This is in keeping with the biology *D. setosa* which is a euryhaline r-selected opportunist with a high biotic potential. Whilst it is usually considered to be a non-selective deposit feeder, feeding on decaying organic material, several specimens in the Thames were observed with intact pennate diatoms in their intestines. Several other nematode species appear to infill as the dominance of *D. setosa* declines and species richness increased from 15 taxa between July and December to 25 taxa between January and March.

The richest assemblage of harpacticoids at Greenwich was observed at the intertidal site in the 3rd. survey when 4 taxa were recorded at a total density of 4,549 litre⁻¹. This coincided with the occurrence of the only harpacticoid specimen at the subtidal site

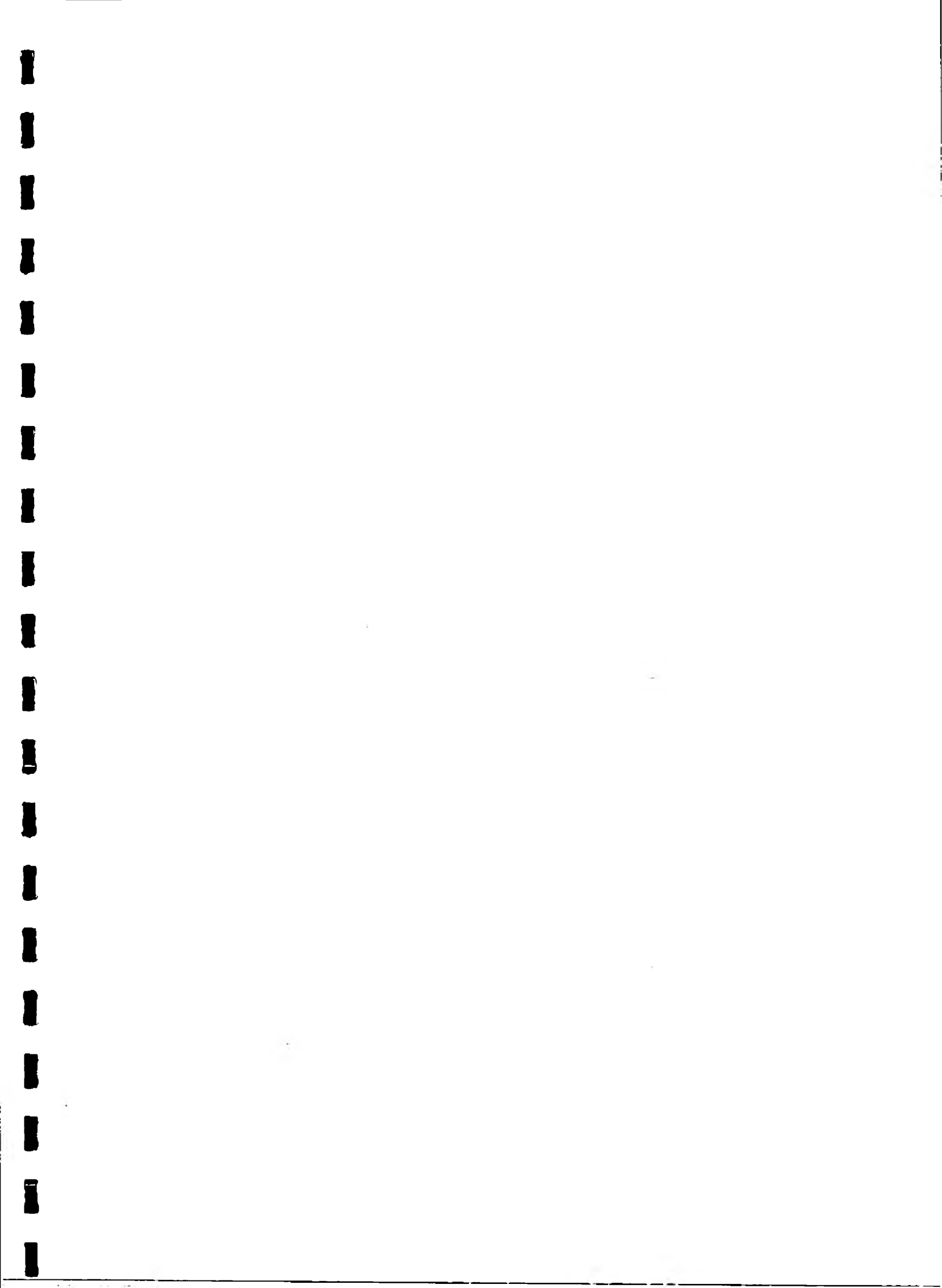
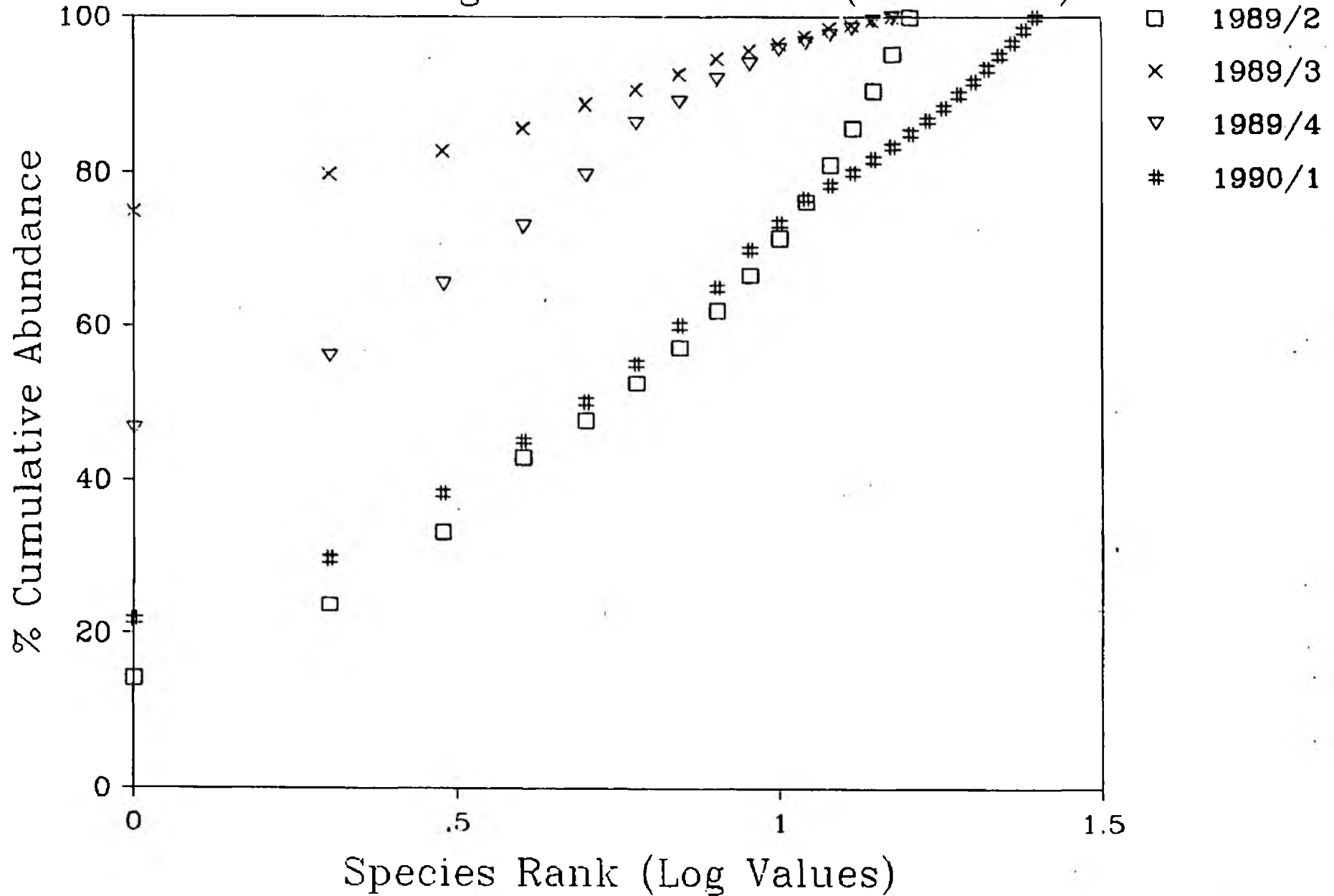


Figure 9. The dominance:diversity characteristics of the nematode assemblages observed at the Greenwich intertidal sampling station between April 1989 and March 1990.

k-Dominance Curves for Nematode Assemblages at Greenwich (Intertidal)





(*Microarthridion* species - also present in the intertidal samples). The high density of harpacticoids supported by the intertidal sediments at this time of year might relate to the high primary productivity of the Greenwich mudflats (Hughes pers. comm.).

With the exception of the 1st. intertidal sample (April - June), juvenile oligochaetes were observed in all the samples examined from Greenwich (subtidal and intertidal). Newly metamorphosed gastropod larvae (probably hydrobiid species) were present in the intertidal samples between October 1989 and March 1990 and the subtidal sample for October - December. In *Hydrobia* and *Potamopyrgus* species, breeding in the East of England usually commences between May and June. Egg maturation may take up to 3 weeks and the veligers may remain in the plankton for up to 1 month which suggests that peak recruitment might be in mid- to late summer. Meiofaunal sampling in the Thames may have missed this peak as the juveniles develop rapidly once settled.

Station 8, Woolwich

The subtidal and intertidal meiofaunal communities at Woolwich are very different from each other. As at Greenwich, the total number of meiofaunal species observed at the subtidal site was lower than at the intertidal. However, this was solely due to the large number of nematode species encountered during the 1st. survey (23 species; see Appendix I; Section 2). In subsequent surveys higher numbers of species were observed at the subtidal site. The reason for this is unknown. Sediment descriptions suggest that the sediment types remained similar with compacted muds occurring sublittorally and fine soft muds in the littoral zone. *Sabatieria* species were present in each of the subtidal samples (*S. punctata* was the only species observed at this site in the 1st. survey). Apart from a single *S. punctata* in the intertidal sample from the 1st. survey, none was observed in any of the later samples. This would seem to suggest that the intertidal and subtidal sites experience unusually different environmental conditions and might merit closer investigation.

Daptonema setosa populations dominated the intertidal nematode assemblages in the first 3 surveys (April - December 1989). In common with the Greenwich intertidal site, the densities of *D. setosa* rose markedly between the April and July sampling programmes. However, peak densities did not occur until the 3rd. survey (6,574 litre⁻¹). This was reflected in the elevated 1B:2A feeding type ratios of the 2nd. and 3rd. surveys. In contrast, in the subtidal assemblages, *S. punctata*, *Adoncholaimus thalassophygas* and *Dichromadora geophila* were more important components of the meiofauna than *D. setosa* which was present at low densities only in the 3rd. and 4th. surveys.

Harpacticoid species were present in each of the samples from the intertidal site and comprised marine and estuarine species. However, adult harpacticoid copepods were not observed in the subtidal samples. Copepodites (indet.) were present in the samples collected in the 1st. and 3rd. surveys. Of the Acari, oribatid species were found at low densities in some of the samples at both sites.

Comparatively few other invertebrate groups were observed in the subtidal samples.



Sections of perisarc and detached hydranths of thecate hydroids were found in the July - September sample indicating the proximity of more stable substrata. Our own observations on the benthos of this reach imply that these might derive from epifaunal colonies on large cobblestones. Hydranths were also present in the April - June sample from the intertidal site.

Station 9, Beckton

The observed meiofaunal assemblages at Beckton included a reduced number of nematode and harpacticoid species. The cumulative totals for these groups were 14 and 1 species, respectively. Amongst the nematodes, several fresh and brackish water species were present. Many of these were microbivorous. The remaining nematodes comprised euryhaline, non-selective detritivores, such as *S. punctata* and *D. setosa*, and selective deposit feeders/microbivores, such as the leptolaimids, *Leptolaimus papilliger* and *Leptolaimoides* species. The diatomivorous species, *Ptycholaimellus ponticus* was also present in the muddy Beckton sediments collected between October and December along with low densities of the predatory marine species *Enoplolaimus vulgaris*. The latter 2 species are generally found in higher salinity habitats. The cletodid harpacticoid copepod species observed as a single specimen in the January - March samples may have been an aberrant record; it resembled closely species more usually found in well oxygenated sandy sediments. Similarly, the Acari were also represented by a solitary specimen of an oribatid species present in the July-September survey.

Densities of other faunal groups were low and comprised mostly single individuals. Tests of marine Foraminifera were present in the first 3 surveys at densities of up to 200 litre⁻¹ (mostly planispiral *Elphidium*-type species - possibly *E. striato-punctata*). However, it is likely that these were transported to the site from more saline reaches as they are relatively intolerant of reduced salinities likely to be encountered near the Beckton sewage treatment works.

Station 10, Crossness

The subtidal meiofauna at Crossness comprised a species-poor assemblage dominated by the euryhaline nematode species, *Sabatieria punctata* (1st., 2nd. and 4th. surveys) and the predator *Adoncholaimus thalassophygas* (3rd. survey). Where type 2A nematodes were present, their relative importance was outweighed by non-selective detritivore species. This was reflected in the 1B:2A ratios (> 1.00). Throughout, the species present were essentially marine although some lower salinity species persisted in the January - March survey. These included *Ironus ignavus*, a predatory freshwater ironid, and a diplogasterid species - a fresh/brackish water species belonging to a family that includes predators as well as microbivorous nematodes. This station also marked the upstream limit of *Richtersia inaequalis*, a species that thrives in organically rich sediments but is more commonly associated with higher salinity conditions. A single specimen of *R. inaequalis* was observed in the January - March survey. This species became an important component of the

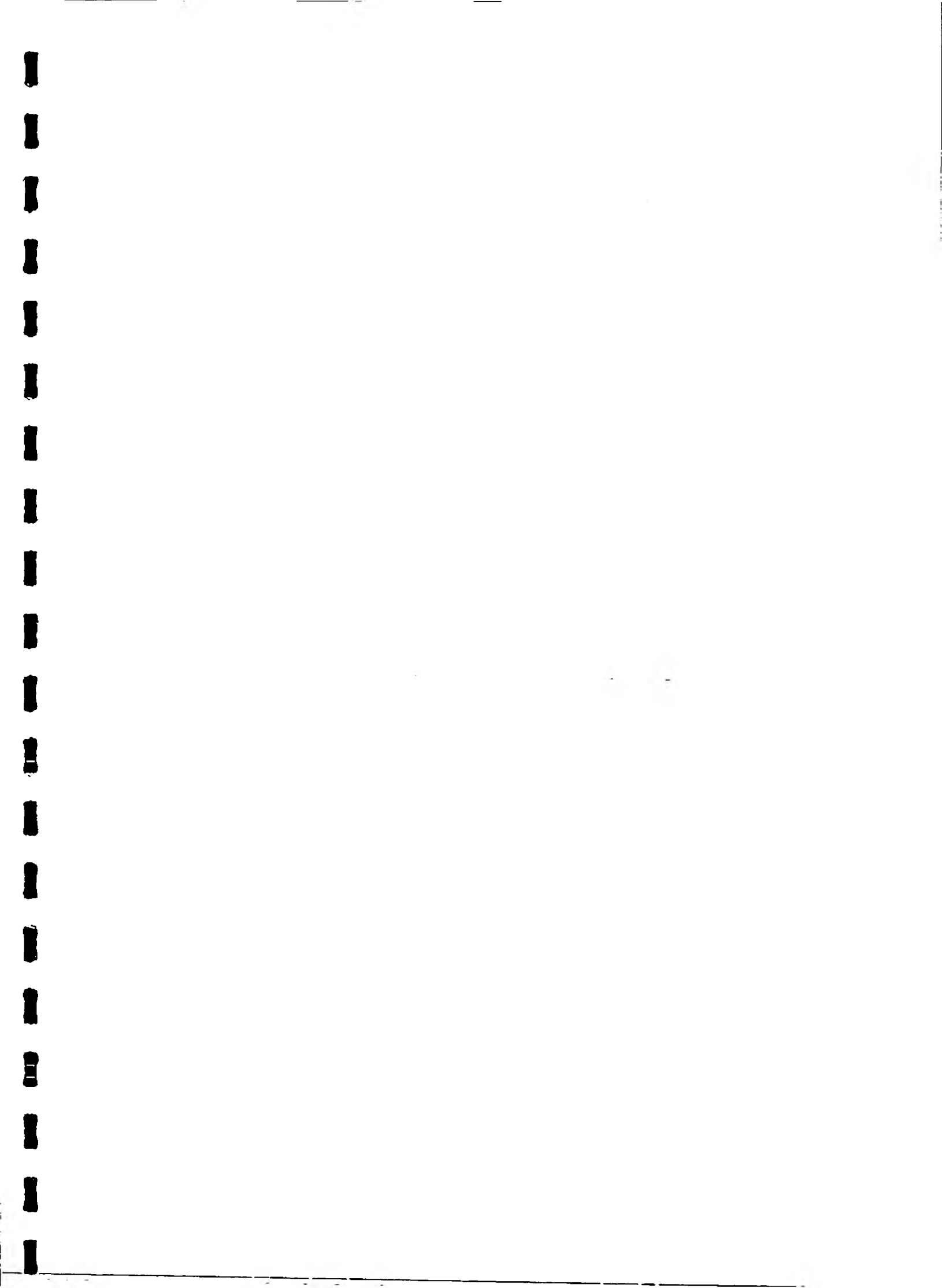
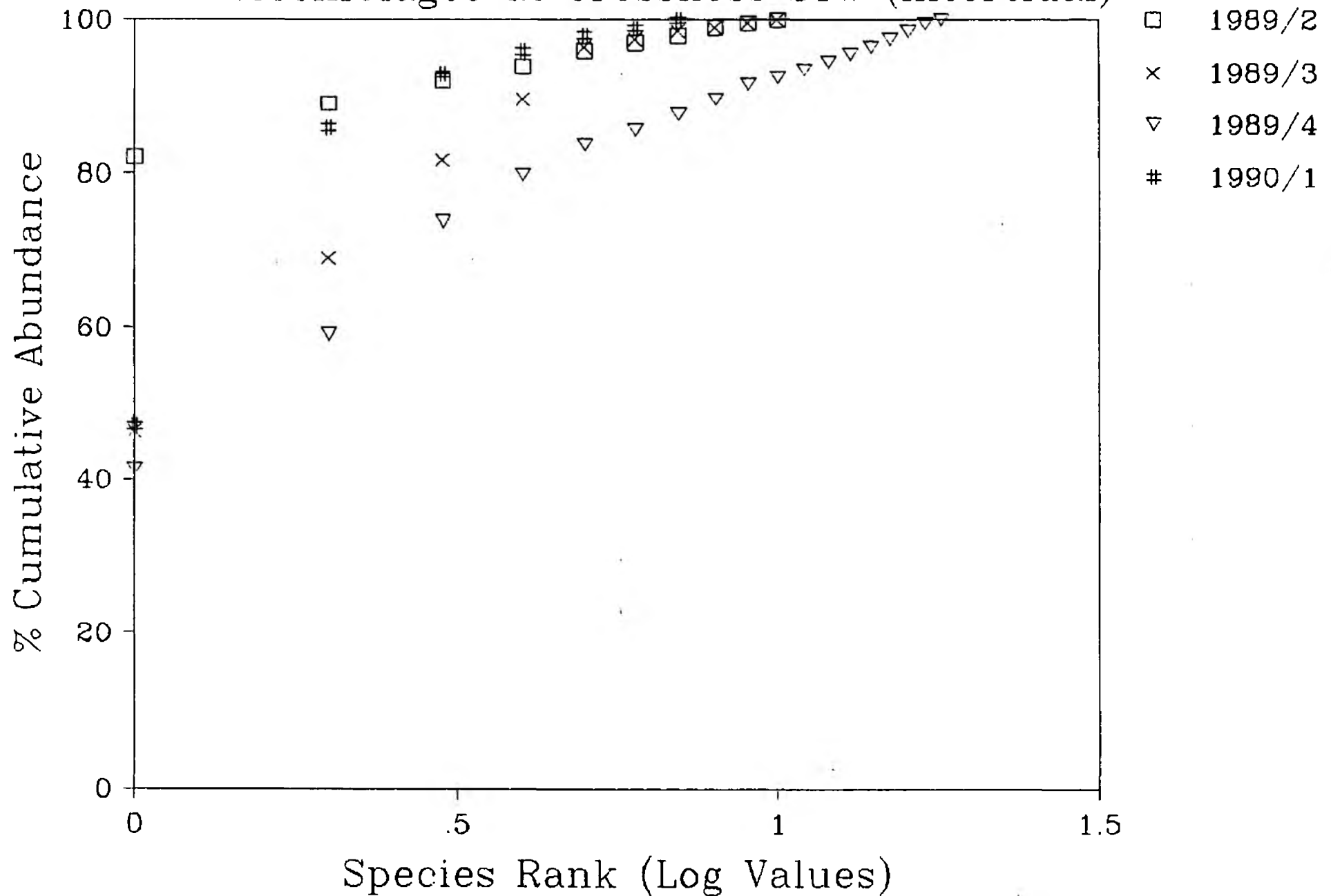


Figure 10. The dominance:diversity characteristics of the nematode assemblages observed at the Crossness intertidal sampling station between April 1989 and March 1990.

k-Dominance Curves for Nematode Assemblages at Crossness STW (Intertidal)





meiofauna at stations nearer the mouth of the estuary (see Stations 19 and 22). As noted elsewhere, the total number of species observed at the subtidal site was lower than at the intertidal (18 species as compared to 25 species).

Total meiofaunal densities were also higher in the intertidal sediments reaching nearly 62,000 individuals litre⁻¹ sediment between July and September. The latter was due principally to the high densities of 3 nematode species, namely *Dichromadora geophila*, *Anoplostoma viviparum* and *Daptonema setosa*. *D. geophila* is a comparatively large type 2A chromadorid species that is common in low salinity habitats in Britain and parts of Europe where it may feed on diatoms. *A. viviparum* is also common in low salinity waters but is suggested to feed on Protozoa (Von Thun in Bouwman, 1983). *D. setosa*, in the Thames at least, is probably diatomivorous. The latter species predominated in the intertidal sample, representing approximately 83% of the nematode population in the April - June survey (Figure 10). Dominance was lower in all other intertidal samples examined.

The harpacticoid population in the subtidal samples was restricted to a relatively few unidentified copepodites that were observed in the first survey. The intertidal site offered a slightly higher diversity including copepodites and the diosaccids *Stenhelia giesbrechti* and *Paramphiascella* species. None was observed in the last survey. The harpacticoid complement and their densities were lower than might usually have been found at this level in other estuarine systems such as the Tees or Humber. Acari were observed in the intertidal samples only and were restricted to the oribatid species and *Copidognathus rhodostigma* found in the first 2 surveys.

Other faunal groups included low densities of newly settled bivalve molluscs in the subtidal sample (<25 litre⁻¹; January - March). The identity of these could not be determined and their ability to survive in these sediments was not known. In the intertidal samples from the 1st. survey, free amphipod embryos were noted (probably *Corophium* species). These would normally be retained by the adult in the marsupium until they were fully developed and capable of a free-existence. Whilst they might have become dislodged during sample preparation, there is some suggestion that *Corophium volutator* may release its embryos into the mud that forms its burrows which it then ventilates (Feil, pers. obsvn.; Hughes, pers. comm.). Ostracod species at densities of up to 75 litre⁻¹ were also observed in the 1st. survey of the intertidal site.

Station 11, Purfleet

This station was characterised by variable densities of estuarine meiofauna. Species richness was again comparatively low with a cumulative total of 29 principal meiofauna species in the intertidal and 28 in the subtidal samples. In the subtidal samples, the nematodes *Daptonema setosa* and *Sabatieria punctata* were the only species observed in each of the surveys. Both species exhibited peak densities in the January - March survey (707 and 7069 individuals litre⁻¹, respectively). Maximum total nematode densities, however, were observed in the preceding survey (9,770 litre⁻¹, October - December) which correlated with the highest species richness (17 species). In the intertidal samples, nematode densities were much lower and ranged from 33 litre⁻¹ (October - December), when 2 species only



were observed, to 3,000 litre⁻¹ (July - September). The peak density again correlated with the highest species richness (15 species).

The trophic groups represented in the nematode assemblages were strongly biased towards the non-selective detritivores (type 1B species). However, in the Thames, some of these species may well specialise, ingesting whole benthic diatoms for example (*D. setosa* and *Desmolaimus zeelandicus*).

The numbers of harpacticoid species were greatest at the subtidal site with a total of 6 taxa being recorded during the survey period. Most of these were present in the January-March survey (4 species) and were all epibenthic estuarine species. As with the nematodes, the variability was high and none was observed in the subtidal samples in the 2nd. survey. Three harpacticoid species observed in the intertidal samples - *Tachidius discipes* and 2 *Stenhelia* species. The latter are common estuarine species that can tolerate a broad range of salinities (0 to 30 ppt) and temperatures from below zero to 30 °C. As such, it is ideally suited to the extreme conditions that can occur in the littoral habitat. It was also the most abundant species in the sublittoral samples at this station.

The Acari were sparsely distributed in the samples examined with *Copidognathus rhodostigma* occurring in the intertidal and sublittoral samples in April - June only and a record of single sublittoral oribatid in the January - March survey. Other invertebrate groups were also sparse; immature oligochaetes and crustacean nauplii accounted for the majority of other organisms observed in the sediments at this station. Amongst the algae, diatoms and their frustules were widespread and abundant.

Station 12, West Thurrock

The intertidal nematode population at West Thurrock was indicative of a comparatively healthy estuarine mud ecosystem. The feeding type ratios at this site were lower than might be expected for a moderate salinity mud but probably related to the high densities of type 2A species that exploit the surface blooms of benthic diatoms that occur during low tides. The intertidal population appeared to be more stable than those described further upstream with a higher degree of correspondence between the species complements. Six species were present in all 4 surveys and species richness varied little between the samples (16 to 18 species observed per survey). The dominance-diversity curves (Figure 11) exhibit a high degree of congruence with dominance varying between approximately 19 and 39% only. Further, the minimum faunal similarity (presence:absence) between any two sequential samples was approximately 48% (October - December and January - March surveys); the highest similarity of species complements was between the July - September survey and that in October - December assemblages (65%).

Nematode densities in the intertidal samples did not fall below 1,700 litre⁻¹ sediment and were highest in the July - September samples (17,210 litre⁻¹). This peak was principally due to the densities of *S. punctata*. However, the populations of 11 other species also

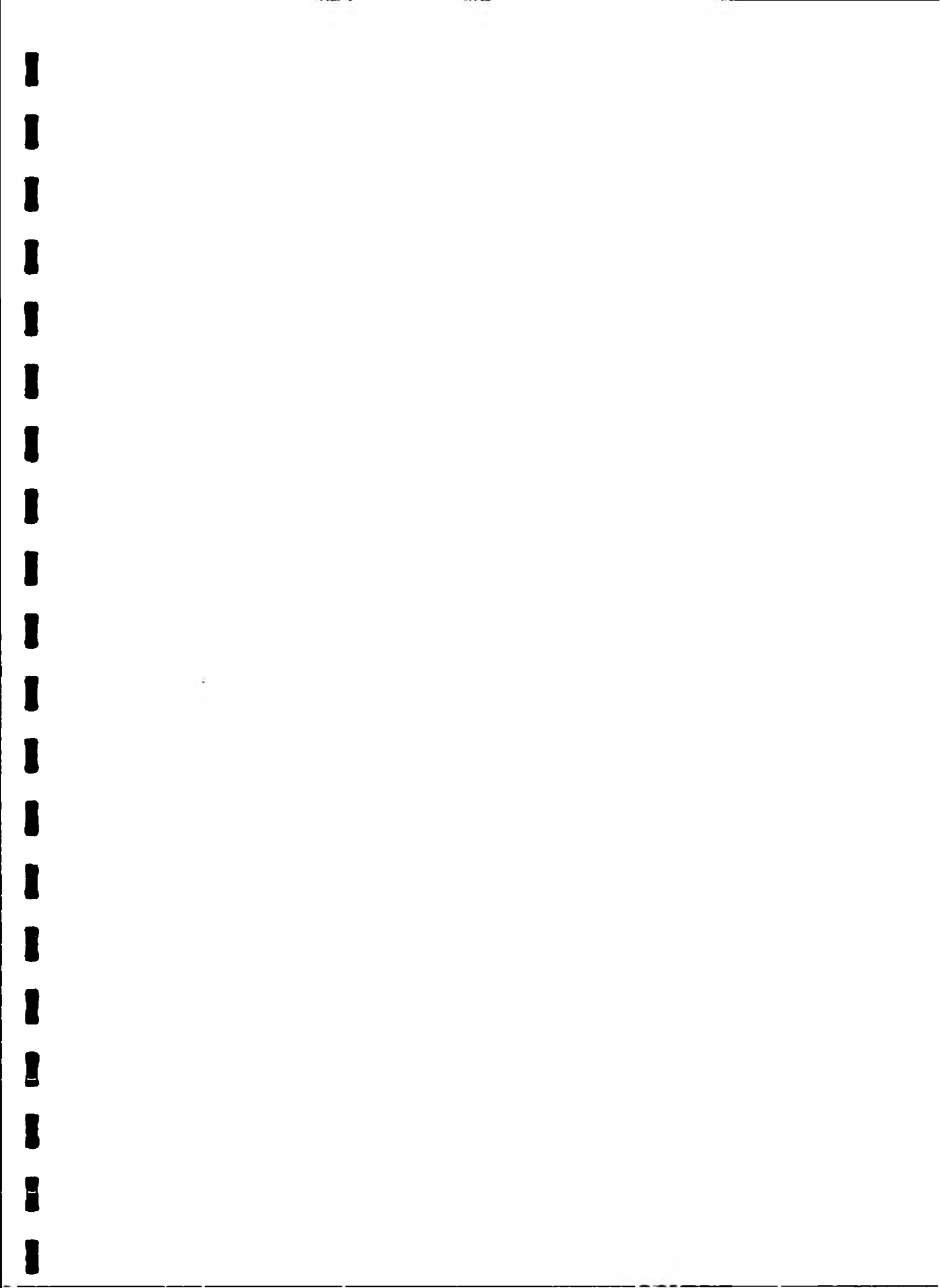
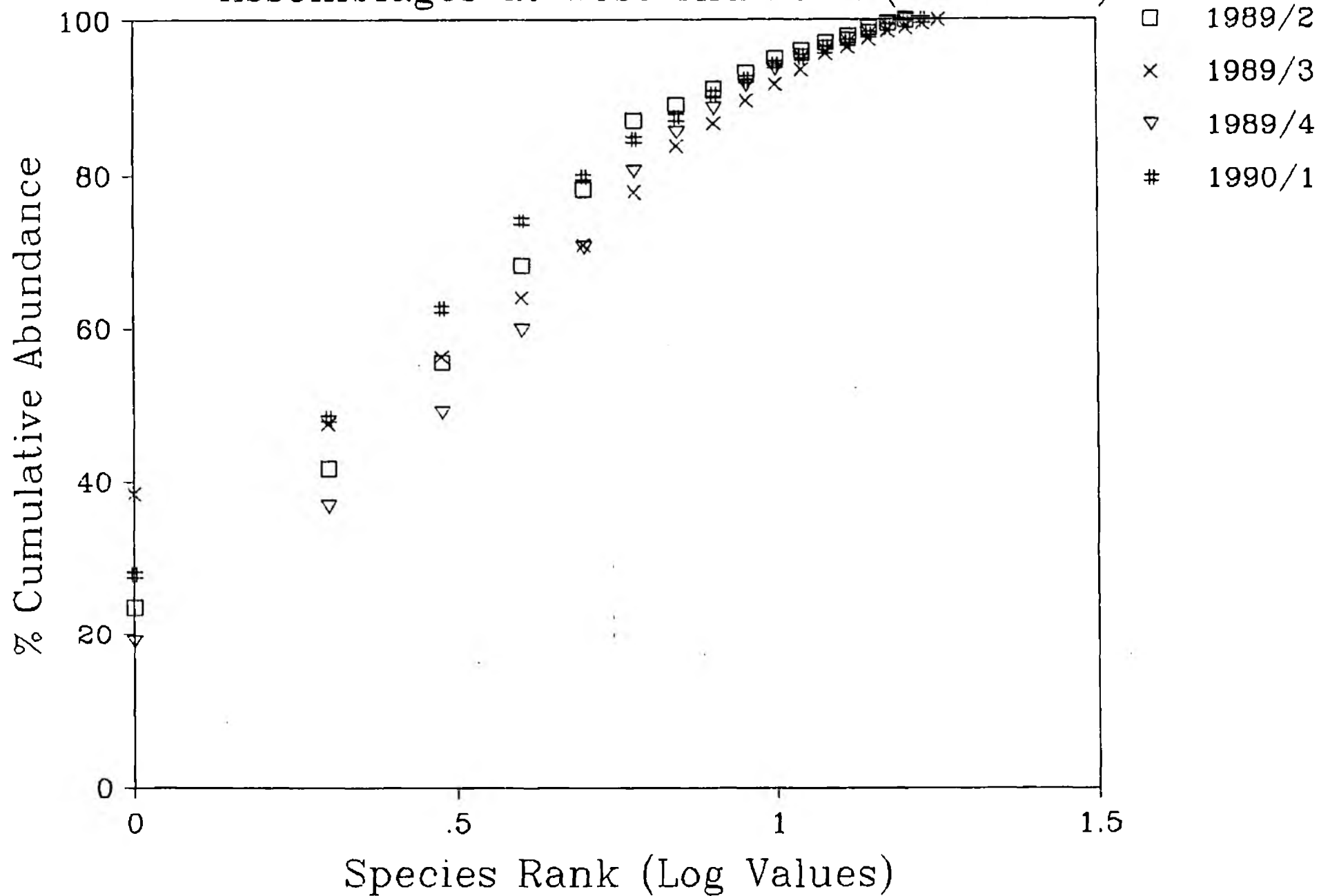


Figure 11. The dominance:diversity characteristics of the nematode assemblages observed at the West Thurrock intertidal sampling station between April 1989 and March 1990.

k-Dominance Curves for Nematode Assemblages at West Thurrock (Intertidal)





attained their highest observed densities in this sample. In the subtidal samples, the nematode populations were more variable in composition. Of a cumulative total of 20 species (as compared to 36 in the intertidal samples), 1 species only was present in all 4 samples (*S. punctata*) and 4 species only were present in 2 or more samples. The sample from the 2nd. survey appears to be anomalous as it comprised 38 *S. punctata* litre⁻¹ only. The reason for this is unknown but is believed to be reflected in the macrofaunal assemblage as well (Attrill, pers. comm.).

A similar dichotomy existed between the harpacticoid populations of the inter- and subtidal sites. In total, 8 taxa were recorded in the intertidal samples at densities of up to 270 harpacticoids litre⁻¹ (July - September). Apart from diosaccid copepodites present in the 1st. survey, a single species only (*Amphiascus* species 2) was recorded in the subtidal samples in the final survey, and then at low density.

The other invertebrate phyla assemblages were characterised by the occasional occurrence of amphipod embryos (probably *Corophium* species) and newly settled gastropod larvae. A single tardigrade, *Batillipes* species (probably *B. mirus*) was observed at the subtidal site between October and December. This marks the furthest upstream site at which this marine species was observed.

Station 13, Gravesend

The marked increase in meiofaunal species richness at the subtidal and intertidal sampling sites at Gravesend as compared to upstream stations indicates, amongst other things, the increased influence of full strength seawater. Fifty-one nematode taxa were recorded at the intertidal site and included many true marine species. The presence of some coarse grained material observed amongst the fine, oxidised mud undoubtedly promoted this diversity, enabling selective epigrowth feeders to colonise the sediments. This said, the trophic balance was always in favour of non-selective detritivores (see 1B:2A ratios, Appendix 1; Section 5). Highest nematode densities occurred in October - December (14,286 litre⁻¹) and declined to their lowest observed value (2,199 litre⁻¹) in the final survey.

The subtidal nematode assemblage was less diverse and comprised a total of 29 species. With the exception of 4 species, all those present at this site were observed in the intertidal zone. *S. punctata* and *D. setosa* were the dominant species. Maximum densities of these species occurred in the 1st. survey between April and June and coincided with the maximum total nematode density (3,141 litre⁻¹). In each of the surveys non-selective detritivores predominated although all the other feeding types were well represented.

Harpacticoid copepods were abundant and diverse in the intertidal samples with densities ranging from 54 litre⁻¹ to 2,144 litre⁻¹. The 8 taxa observed were all epibenthic and, in the July - September survey, included the freshwater *Bryocampius* species found in the upstream samples. A similar species complement was found in the subtidal samples (6 epibenthic species) but these were largely restricted to the final survey. None was observed



in the July - September survey which correlated with the lowest densities of harpacticoids observed in the littoral samples. Acari were poorly represented at Gravesend and a single specimen of *Copidognathus rhodostigma* was the only record (intertidal site; April - June).

Marine ostracods were a common component of the Gravesend fauna at densities of up to 120 litre⁻¹ occurring in 5 of the 8 samples from this station. In common with other sites such as West Thurrock, the 1st. survey also yielded several free amphipod embryos which might indicate the close proximity of *Corophium* beds. Newly settled bivalves were present in 3 of the 4 intertidal samples and 1 of the subtidal samples.

Station 14, Mucking

The reduced meiofaunal species complement at Mucking may relate to the proximity of the Creek and a localised zone of reduced salinity. However, with the exception of the plant-parasite *Criconemoides* species that was observed in the 1st. survey, there is little faunal evidence for the influence of fresh or low salinity water. Thirty-two nematode species were observed in total. Linhomoeid species 1 and *S. punctata* were the only species to occur in all the samples and were co-dominant (alternately dominant and sub-dominant). The family Linhomoeidae includes numerous estuarine and higher salinity species that are common in muddier substrata. The large, unarmed buccal cavities of many linhomoeid species (including Linhomoeid species 1) has lead them to be classified as non-selective detritivores (type 1B species). However, our own observations suggest that some may be highly specialised diatomivorous species capable of ingesting whole pennate and centric diatoms.

The high densities of Linhomoeid species 1 and *S. punctata* are reflected directly in the high total nematode densities and the elevated feeding type ratios (q.v.). Total nematode densities were uniformly high throughout the year and showed little seasonal variation, although a slight peak was observed in the 2nd. survey. This is unusual and might indicate a change over from herbivorous species to those capable of exploiting allochthonous organic material or the bacteria that it might support.

Six taxa of harpacticoid copepods observed in the Mucking samples. Of these *Halectinosoma curticorne* generally occurs in high salinity sites although it can tolerate reduced salinities. It is interesting to note that it was found in the 3rd. survey only at low density. This might relate to an earlier period of reduced freshwater flow from the upstream Thames catchment and the Creek.

Acari did not form a significant component of the meiofauna at Mucking and were represented by low densities of an oribatid species in the 2nd. and 3rd. surveys. Other invertebrates present in the samples confirmed the influence of marine communities, especially in the 3rd. survey when juvenile nemertean were also observed.

Station 15, Blythe Sands



In comparison with upstream stations, increased densities of representatives of the nematode family Desmodoridae were apparent in the sandy muds present at Blythe Sands. This family contains predominantly marine species that specialise in feeding on marine diatoms and the epigrowth that develops on sand particles. The 47 nematode species observed during the 4 surveys are all characteristic of coarser sediments in outer estuaries of European river systems. *S. punctata* was the only species common to all surveys at this site. A faunal similarity analysis (Jaccard presence:absence index) shows the species complements present in the 1st. and 4th. surveys to be most similar (41.9%) and those described in the 1st. and 2nd. surveys to be the least similar (15.7%). Similarities between the 2nd. and 3rd. surveys and the 3rd. and 4th. are 21% and 24%, respectively. This implies a cyclical loss and recruitment of species during the year.

Highest nematode densities were observed in the April - June survey and the lowest in the July - September samples. This appears unlikely to have been a natural phenomenon and correlated with the appearance of abnormally high densities of detritus in the July - September Blythe Sands sample. The origin of this material was not known but it supported several microbivorous (type 1A) species many of which were unique to this quarter's sample. These included Leptolaimid species 1, *Leptolaimus* species 1 and *Monhystera filicaudata*. These species were more characteristic of stations upstream of Blythe Sands.

As with the nematodes, the densities of harpacticoid copepods were highest in the 1st. survey (3,320 litre⁻¹) and lowest in the 2nd. survey when none was observed. The numbers of individuals then increased to 108 litre⁻¹ in the 4th. survey. The ectinosomatid species present dominated the assemblages and, in the case of *Ectinosoma melaniceps* in the 1st. survey, accounted for 99.5% of harpacticoids present. Species of the genus *Phyllothalestris* are generally associated with marine habitats and the occurrence of a species in the Blythe sample in the January - March survey may represent the inland extreme of its distribution.

Acarine assemblages comprised oribatid species present in the 2nd. and 3rd. surveys and *C. dentatus* in the 3rd. survey. Other groups included several spionid polychaete larvae (neochaetes) in the April - June and July - September samples. Marine and outer estuarine ostracods were observed in all the samples examined.

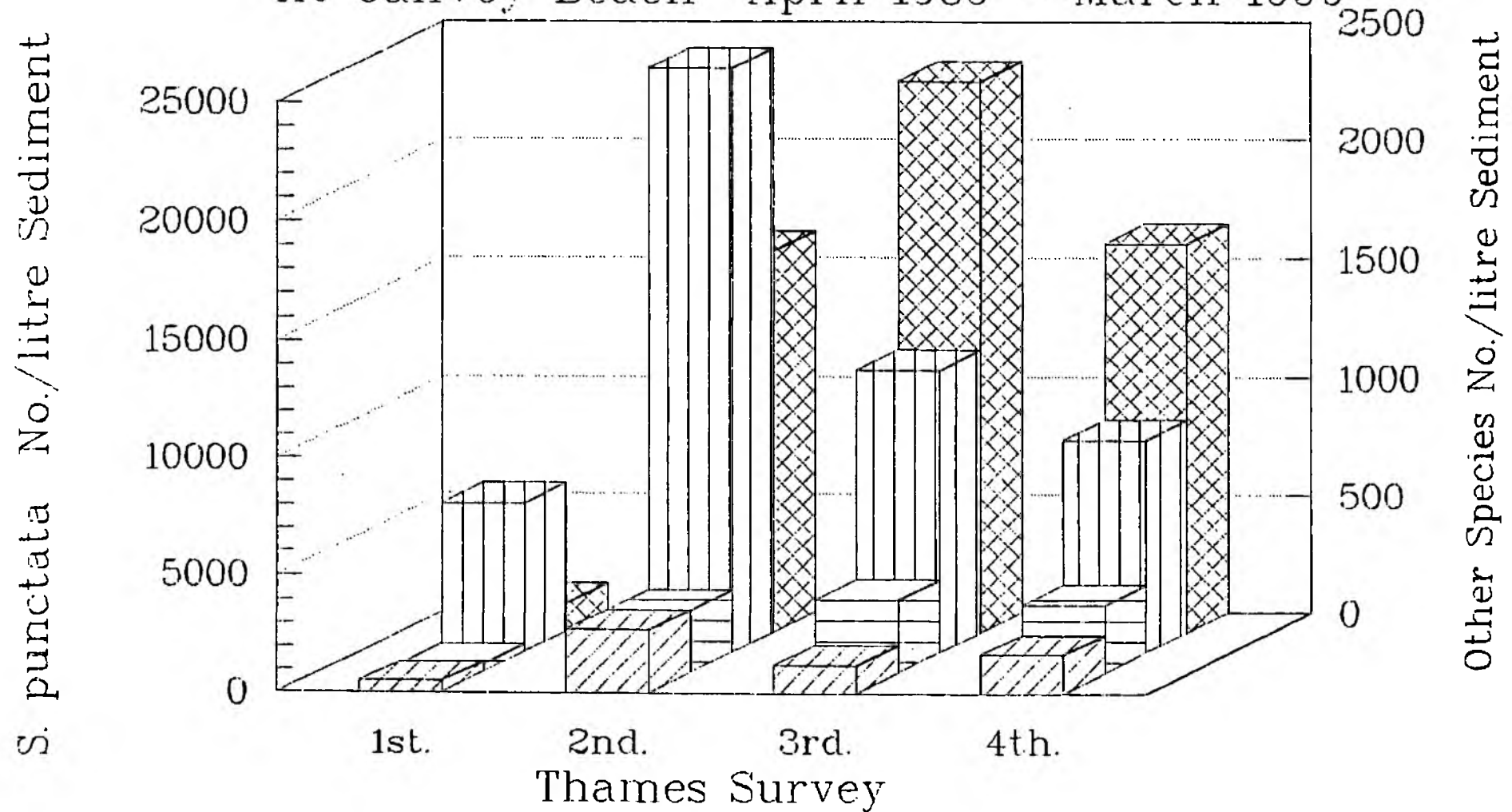
Station 16, Canvey Beach





A typical, species-rich, high salinity meiofaunal assemblage was present in the muds at Canvey Beach. The 4 assemblages of nematodes described contained a total of 64 species at densities of between 13,483 litre⁻¹ (1st. survey) and 55,433 litre⁻¹ (2nd. survey). Species discovery curves suggest that over 100 species might be present at this site. Seven species were present in all the samples examined and included *S. punctata* which was dominant in each case at between 22% and 53% (4th. and 1st. surveys, respectively). This suggests that the sediment may be detritus-rich. Although the majority of species were marine or estuarine, some freshwater species were observed. These included the plant-parasite, *Macroposthonia* species, more commonly found in agricultural soils and drainage ditches.

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Figure 12. Histograms showing the seasonal abundance of *Terschellingia longicaudata*, *Sabatieria punctata*, *Daptonema setosa* and *Ptycholaimellus ponticus* at the Canvey Beach sampling station, Thames Estuary, April 1989 - March 1990. Note different density scale for populations of *S. punctata*.

Abundance of Selected Nematode Species At Canvey Beach April 1989 – March 1990



- Key to Nematode Species:
-  *Terschellingia longicaudata*
 -  *Ptycholaimellus ponticus*
 -  *Sabatieria punctata*
 -  *Daptonema setosa*



However, the relatively high densities of oxystominids, desmoscolecids and monoposthiids, amongst others, confirm the marine nature of this site.

The data for this sampling station illustrate the ecological strategies of several different species. *S. punctata* and *Terschellingia longicaudata* have maximal densities in the samples collected between July and September (Figure 12). As a non-selective detritivore, *S. punctata* is able to exploit the detritus and associated bacteria directly and its populations are rarely substrate-limited in the early part of the year. Consequently, densities of *S. punctata* increase steadily from spring onwards in response to the higher water temperatures. The autumn decline may reflect a reduction in suitable organic material and bacteria and/or competitive stress exerted by populations of other non-selective deposit feeders. *T. longicaudata* is a type 1A species and feeds selectively on deposits, thriving on the rich bacterial flora present in muddier sediments. Its numbers also increase in the early summer in response to elevated water temperatures. In contrast, *Prycholaimellus ponticus* and *D. setosa* are dependent to a greater or lesser extent on diatoms as food and their populations peak in the final quarter of the year (October - December) after a steady period of increase over the summer months. Growth of their populations relies on spring and summer blooms of benthic and, possibly, planktonic diatoms and is probably held in check until these have occurred. Of the 4 species compared, *S. punctata* and *D. setosa* both exhibit the highest intrinsic rates of natural increase once suitable conditions arise. This opportunistic behaviour combined with their euryhaline physiology makes them characteristic elements of the meiofaunal assemblages of many of the stations examined in the Thames Estuary.

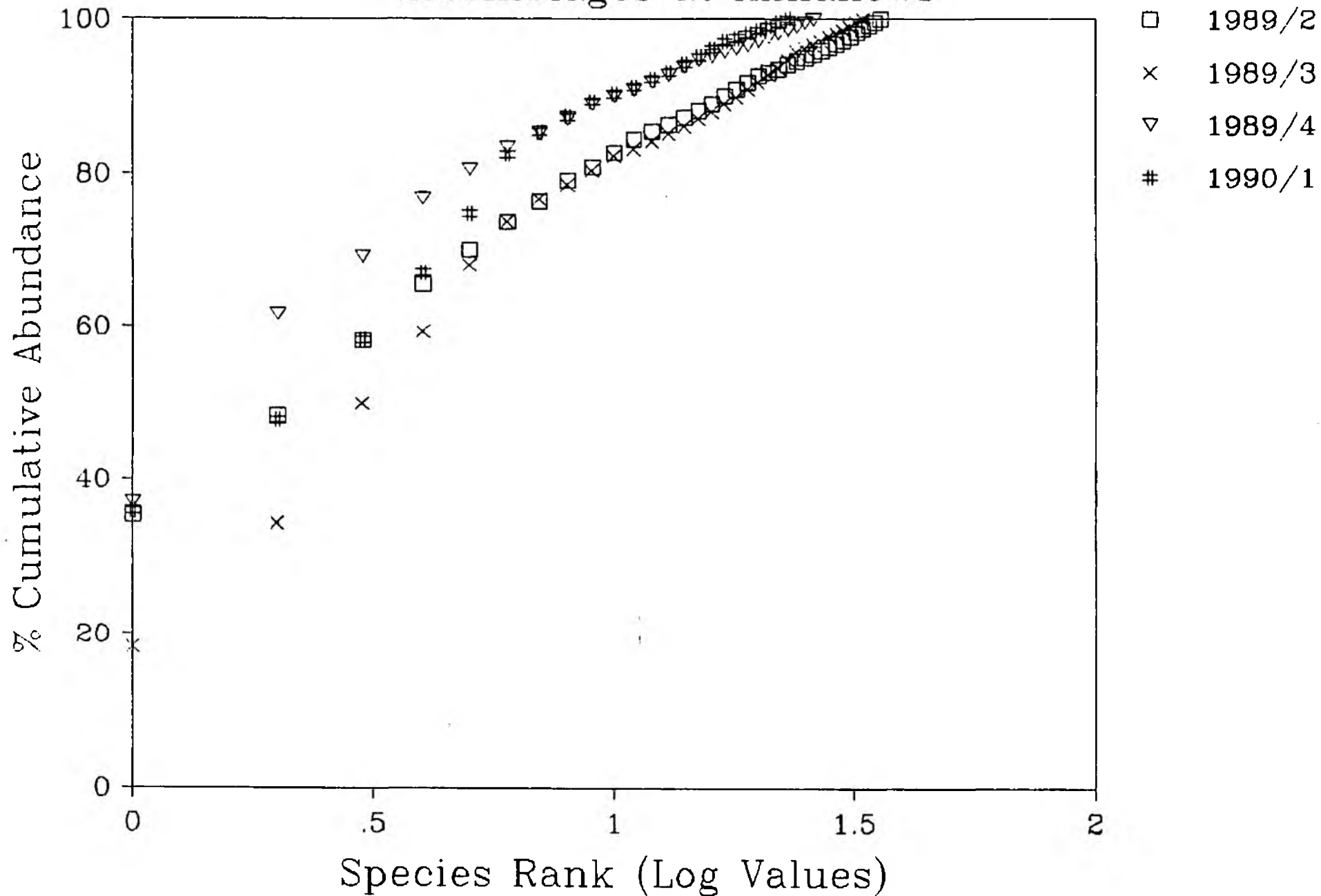
The assemblage of harpacticoid copepods observed at Canvey Beach was amongst the most diverse found in the Thames survey area. Twenty-one species were recorded during the course of the survey. All were of estuarine or marine origin and were predominantly epibenthic species feeding in the upper or surface layers of sediment on detrital material or, possibly, diatoms. Maximal total densities coincided with those of the nematodes in the 2nd. survey. However, 2 species only were observed in the July - September samples. These were *Amphiascus angusticeps* and *Cletodes limicola*. The latter species accounted for 2,687 of the total 2,731 individuals litre⁻¹ observed. This contrasted with the January - March sample, in which the harpacticoid population comprised 11 species at a total density of 1,388 litre⁻¹. Intermediate numbers of species were present at lower densities in the 1st. and 3rd. surveys. The reasons for these marked changes in the harpacticoid assemblage are unknown.

Acari observed at Canvey Beach included *C. rhodostigma*, *C. dentatus* and an oribatid species. All were present at low densities and none was observed in the 3rd. and 4th. surveys. The low numbers and species of Acari in the Thames Estuary in general remains unexplained. Polychaetes, oligochaetes, ostracods, molluscs, turbellarians and crustacean nauplii were all well represented in the Canvey Beach samples. Comparatively high densities of Foraminifera again emphasised the high prevailing salinity at this site.



Figure 13. The dominance:diversity characteristics of the nematode assemblages observed at the Allhailows sampling station between April 1989 and March 1990.

k-Dominance Curves for Nematode Assemblages at Allhallows





Station 17, Allhallows

As at Canvey Beach, the nematodes present in the muddy sediments at Allhallows were almost exclusively marine and estuarine species. The assemblages observed in each survey were similar in composition suggesting that the habitat and prevailing conditions were stable during the period sampled. Of the 60 nematode species recorded, 7 were present in all 4 surveys and 13 species in 3 or more surveys. The 1B:2A feeding type ratios were all less than 1.00, emphasising the relative importance of diatomivorous species at this site. These were mostly chromadorid species although several cyatholaimids were also observed. Dominance-diversity characteristics of the Allhallows nematode assemblages are illustrated in Figure 13. The high degree of congruence of the k-dominance curves again indicates stability of environmental conditions. Dominance alternated between *Metachromadora scotlandica*, a large, epigrowth feeding (?) desmodorid (1st. and 3rd. surveys) and the diatomivore, *Prycholaimellus ponticus* (2nd. and 4th. surveys). Peak nematode densities occurred in the July - September period, as at Canvey Beach, although almost equally high densities were recorded in the April- June and January - March surveys.

Several species of marine and estuarine harpacticoid copepods were observed with a total of 13 taxa recorded in the 4 surveys. These included a low density of *Tisbe* species (100 litre⁻¹) recorded in the 2nd. survey. Members of this genus are usually found in marine habitats although they may occur sporadically in estuaries. It is possible that some species of *Tisbe* feed on decaying macroalgae amongst which they are often found. The densities of harpacticoids correlated directly with those of the Nematoda throughout the year with a maximum density in the 2nd. survey and a minimum in the 3rd. Amongst the aquatic mites, the halacarid *Copidognathus rhodostigma* was the only species to be observed at Allhallows; this was present at a density of 40 litre⁻¹ in the 1st. survey (April - June).

A range of other invertebrate taxa were present in the Allhallows sediments, all essentially of marine origin. Kinorhyncha are a comparatively minor phylum and are generally sensitive to reduced to reduced salinities. As such they are largely restricted to coastal waters. A single specimen of a *Pycnophyes* species was observed at Allhallows during the first survey and several specimens during the 3rd. survey along with individuals belonging to the genus *Echinoderes*. This was the furthest that kinorhynchs were observed to penetrate into the Thames Estuary.

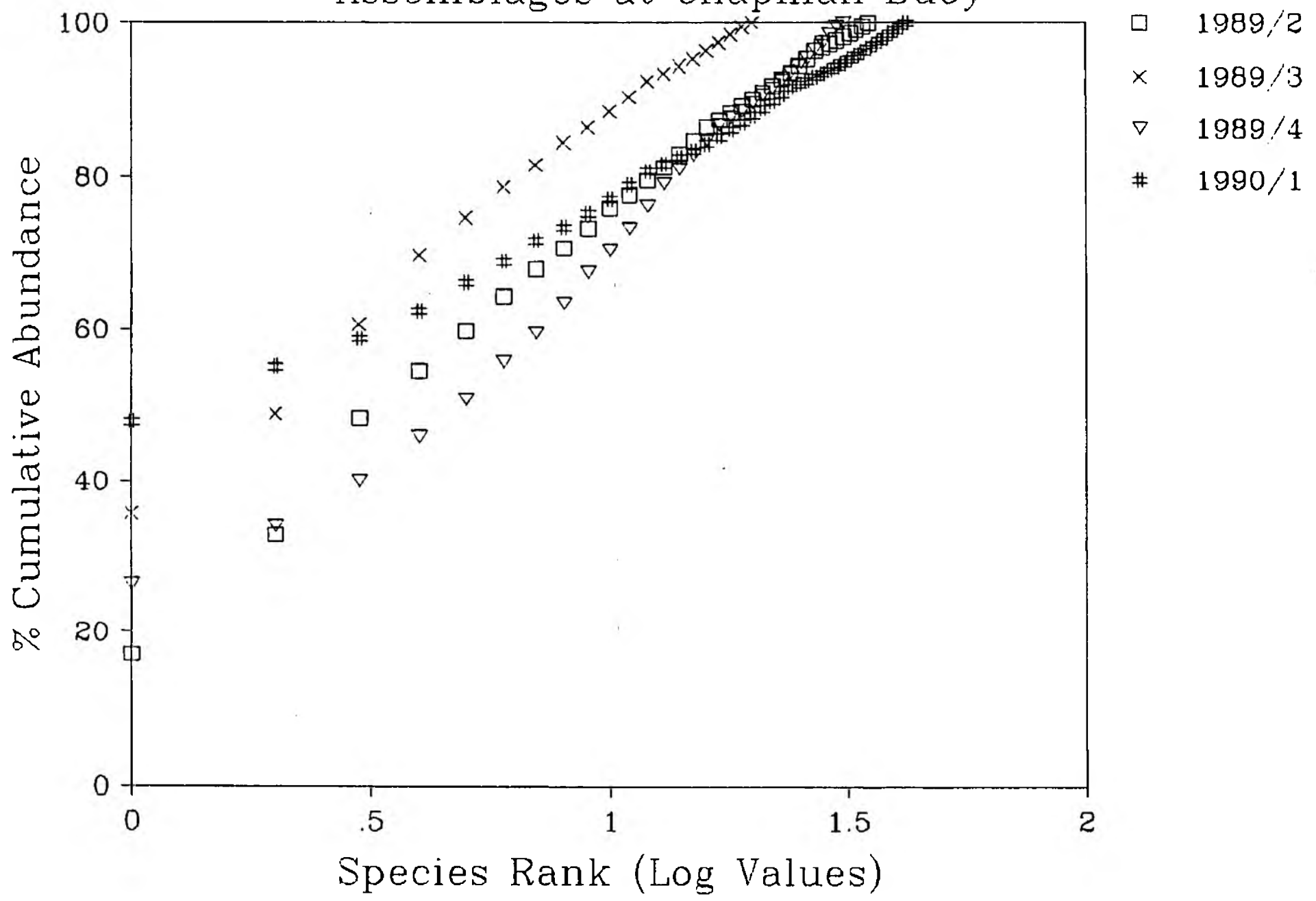
Station 18, Chapman Buoy

The total of 98 principal meiofaunal taxa (nematodes, harpacticoids and mites) observed at this mid-channel station was the highest observed during the year at any Thames sampling station. Examination of the granulometry reveals the heterogeneity of the Chapman Buoy sediments (Appendix II) which may well have accounted for this. Nematode and harpacticoid densities were highest between April and June and lowest between July and September rising again during the last 2 surveys. The lowest numbers of species of both these groups were observed during the 2nd. and 3rd. surveys. The reason for the decline in densities and species richness over the period July to December 1989 is not known.



Figure 14. The dominance:diversity characteristics of the nematode assemblages observed at the Chapman Buoy sampling station between April 1989 and March 1990.

k-Dominance Curves for Nematode Assemblages at Chapman Buoy





Amongst the nematodes, dominance in the 2nd. and 3rd. survey assemblages is intermediate between those of the 1st. and final surveys (Figure 14). This might indicate that several factors are operating as a single tolerant/opportunist species or small group of species does not flourish at these times. Dominance is greatest in the final survey when *Daptonema normandica* accounts for nearly 50% of the individuals present in the population.

Feeding type ratios reflected the muddy nature of the sediment (obscured in the granulometric studies by the high proportion of coarse material (principally wood and shell fragments)). In all cases the 1B:2A ratio was greater than 1.00 as non-selective detritivores predominated. This was least pronounced in the 2nd. survey.

The harpacticoid assemblages were unique to each survey with the majority of the 18 taxa observed occurring in the January - March sample (13 taxa). This high diversity of harpacticoid species coincided with the highest number of species recorded in the nematode samples (42 species). Acari were not seen in the October - December survey. However, the 2 species of *Copidognathus* and an oribatid species were represented in the other samples.

High densities of many other marine groups were noted at Chapman Buoy. The presence of ctenostome bryozoans and detached thecate and athecate hydroids indicated the proximity of semi-permanent or permanent substrata. Small epibenthic and interstitial marine amphipods were especially common in the January - March samples at densities of up to 300 litre⁻¹ sediment.

Station 19, Southend

Exceptionally high densities of nematodes were recorded in the intertidal samples from Southend. The sandy muds at this site supported between 85,700 and 153,000 nematodes litre⁻¹ and exhibited a steady increase in densities from the beginning to the end of the survey period. In contrast, the sandier sediments at the Southend subtidal sampling site yielded between 2,911 and 10,471 nematodes litre⁻¹. Dominance ranges also varied between the 2 sites. In the intertidal nematode assemblages dominance was comparatively low (<31%; Figure 15) and varied little between the surveys (minimum 19%). Dominance was higher in the subtidal samples (Figure 16) ranging from approximately 26% to 44%.

The subtidal assemblages were characterised throughout the year by *Richtersia inaequalis* which dominated in each survey. This selachinematid species is often abundant in sediments that contain elevated levels of organic material where the ambient salinity is sufficiently high. High densities of *R. inaequalis* have been recorded by Trett *et al.* (1990) in the Irish Sea in a study of the Fylde Coast sewage outfalls, including that at Blackpool. In the present study, the 1B:2A ratios were all greater than 1.00 except in the final survey where large numbers of juvenile cyatholaimids were observed (type 2A species grouped under the taxon "Cyatholaimid species 2"). Sandy sediments are more usually characterised by low 1B:2A ratios as high densities of non-selective detritivores rarely occur and

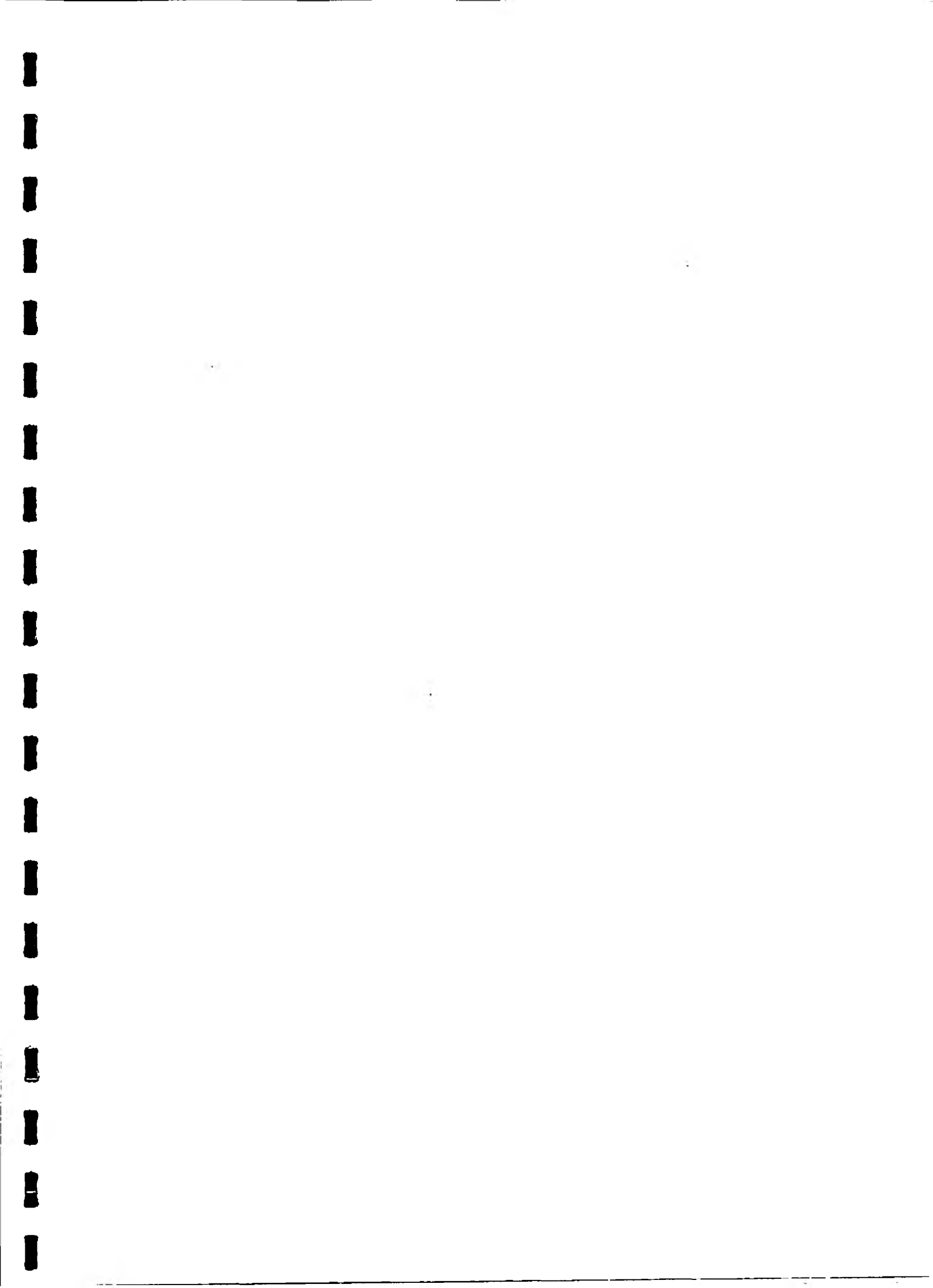
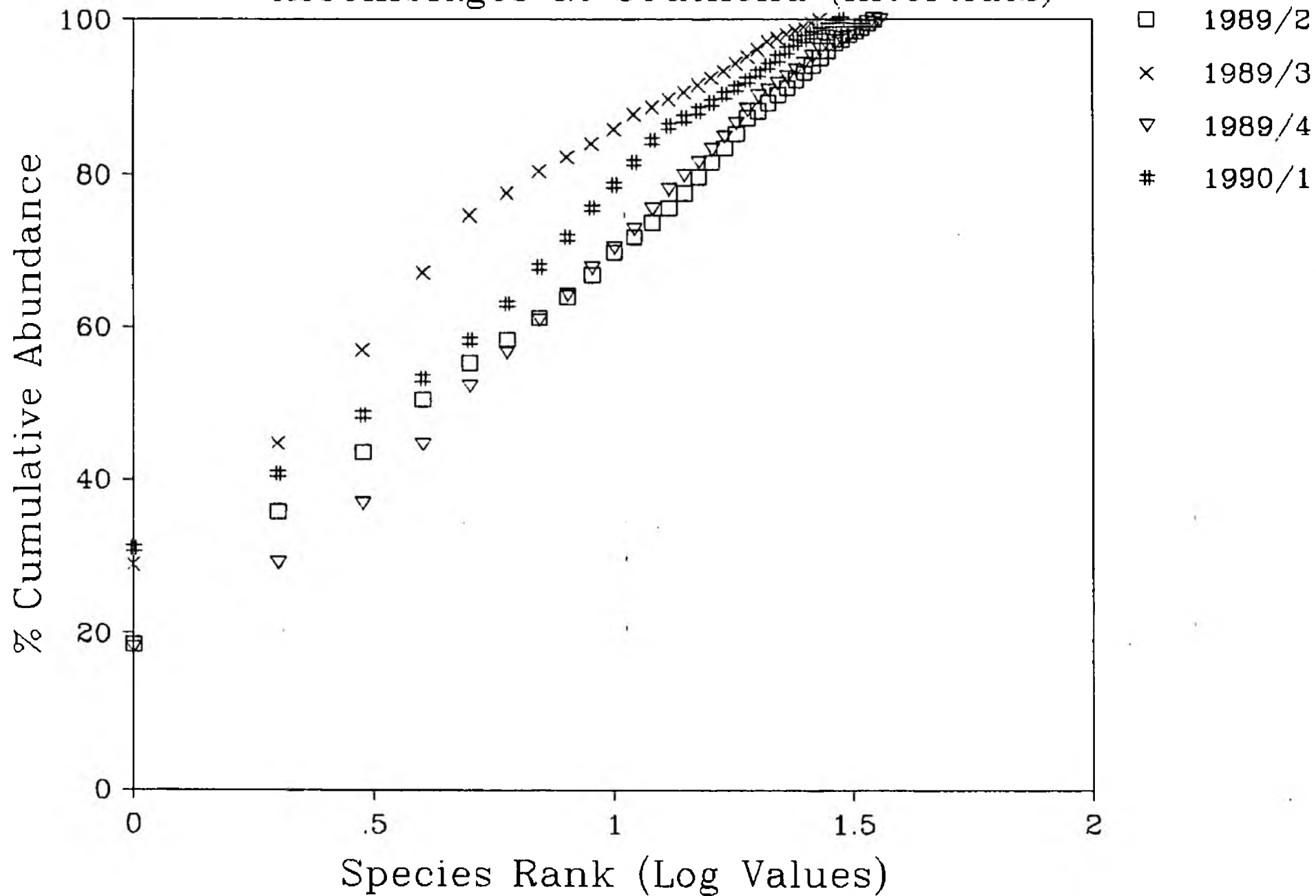


Figure 15. The dominance:diversity characteristics of the nematode assemblages observed at the Southend intertidal sampling station between April 1989 and March 1990.

k-Dominance Curves for Nematode Assemblages at Southend (intertidal)



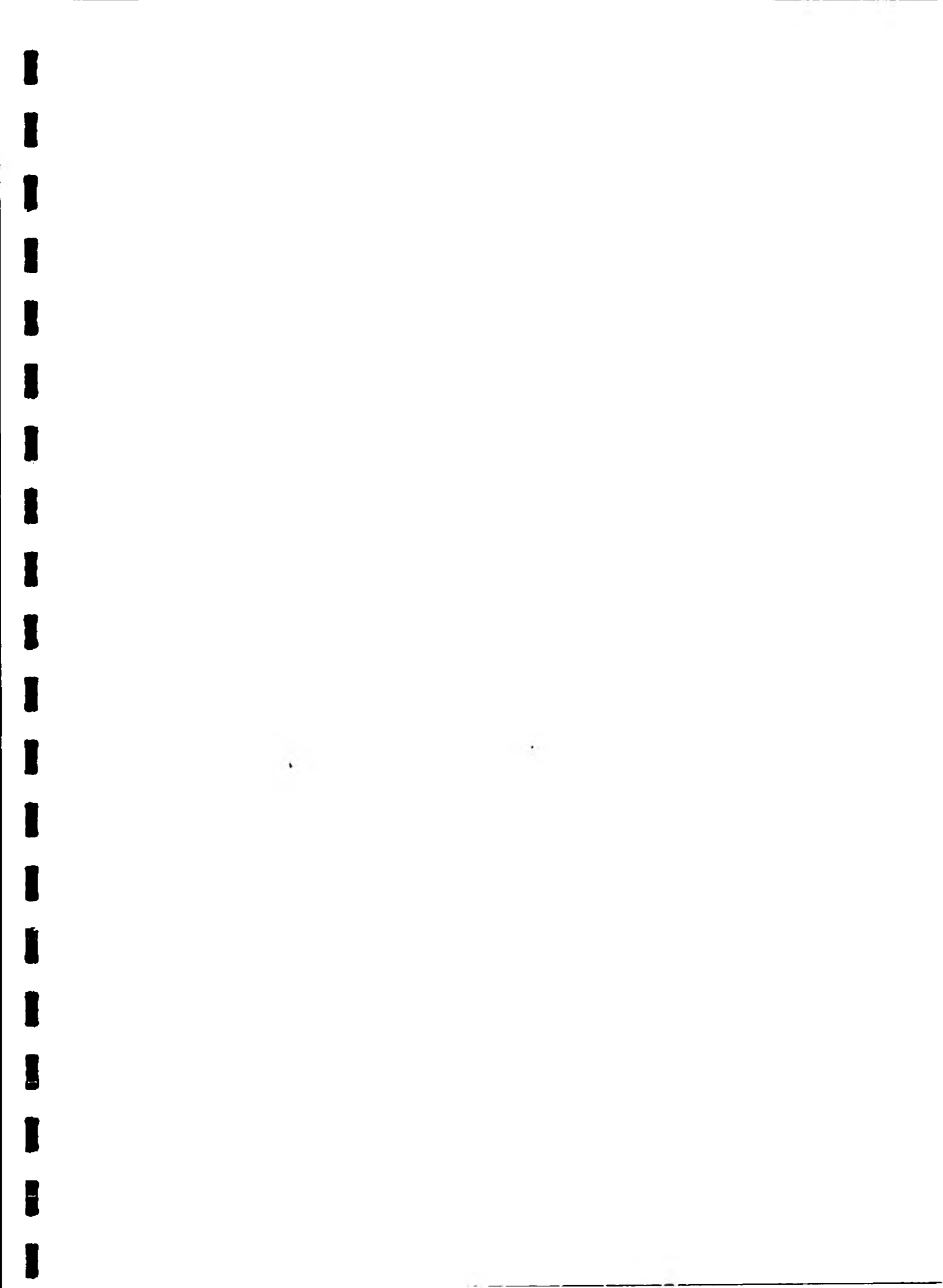
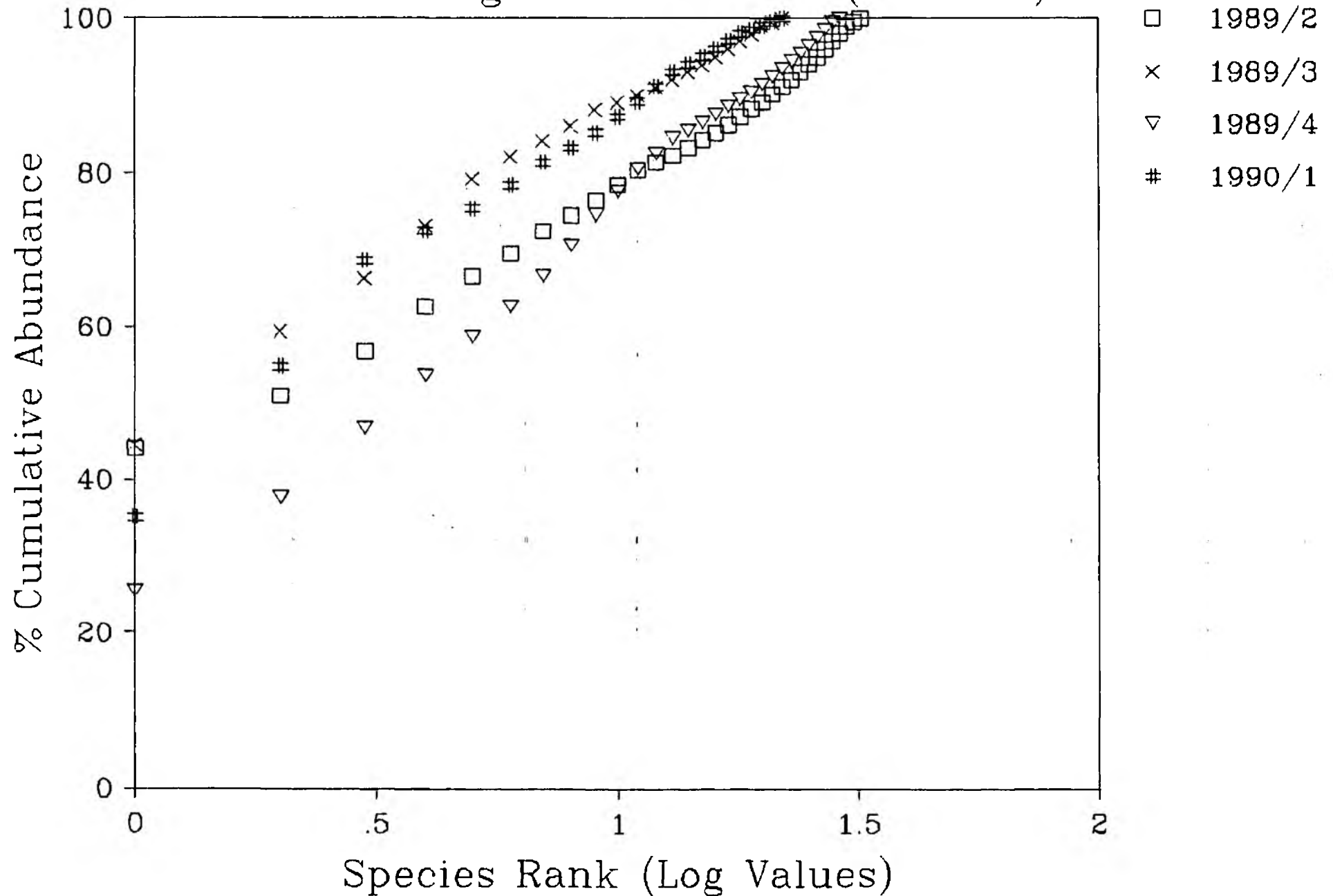


Figure 16. The dominance:diversity characteristics of the nematode assemblages observed at the Southend subtidal sampling station between April 1989 and March 1990.

k-Dominance Curves for Nematode Assemblages at Southend (Subtidal)





epipsammic and diatomivorous species are favoured.

Although similar to one another in species composition (minimum faunal similarity between consecutive surveys 42%), each of the intertidal assemblages was dominated by a different species. All were type 2A species and contributed strongly to the low 1B:2A ratios. In this context, the family Desmodoridae was well represented, numerically, in each of the intertidal samples. Eight species occurred in all 4 intertidal surveys including *Sabatieria punctata*, although this 1B species did not account for more than 10% of any of the nematode populations. As in the subtidal samples, *R. inaequalis* was also present in each survey but at this site represented less than 1% of the nematodes present.

Thirteen harpacticoid taxa were identified in the 4 surveys of the intertidal site at Southend. These were predominantly epibenthic species, many belonging to the family Diosaccidae. The high densities recorded in the April - June survey declined throughout the year from 3,154 to 258 litre⁻¹ in the 4th. survey. Fewer species were found in the subtidal samples (total 6 taxa) and their densities were lower, ranging from 18 litre⁻¹ (January - March) to 281 litre⁻¹ (October - December).

Low densities of oribatid species were noted in 3 of the 4 intertidal samples which might have related to mite populations present in decaying intertidal material. The only acarine observed in the subtidal samples was a single individual of *Copidognathus dentatus* in the final survey.

In general the intertidal assemblages of other invertebrate groups were more diverse than those observed in the subtidal samples. The marine tardigrade *Batillipes mirus* was recorded at both the inter- and subtidal sites at low densities (8 - 16 litre⁻¹) although it was not present in all samples. Turbellaria, however, were ubiquitous. The distribution of ostracods is noteworthy in that they occurred at moderate densities (50 - 125 litre⁻¹) in all the intertidal samples that were examined but were not seen in any of the subtidal samples.

Station 20, Grain Flats

The meiofaunal assemblages at the Grain Flats sampling site are almost indistinguishable from those of sites around the southern coast of England and the southern shores of the North Sea coast of Europe. However, the lower total number of nematode species observed (55 species over the 4 surveys) at Grain Flats was lower than at more inland stations under the influence of full strength seawater. This might have reflected a change in sediment type. In the 1st. survey the sediment conformed to a fine sand that contained some silty material (mostly frustules of centric marine diatoms). During the 2nd. and 3rd. survey the sediment type changed progressively through a muddy sand to a mud with a low sand content. In the final survey the sediment returned to a muddy fine sand. As expected, the highest numbers of nematode species were associated with the coarser grained sediments in the 1st. and 4th. surveys (22 and 29 species, respectively) whilst the muddier sediments of the 2nd. and 3rd. surveys were less species rich (13 and 17 species,

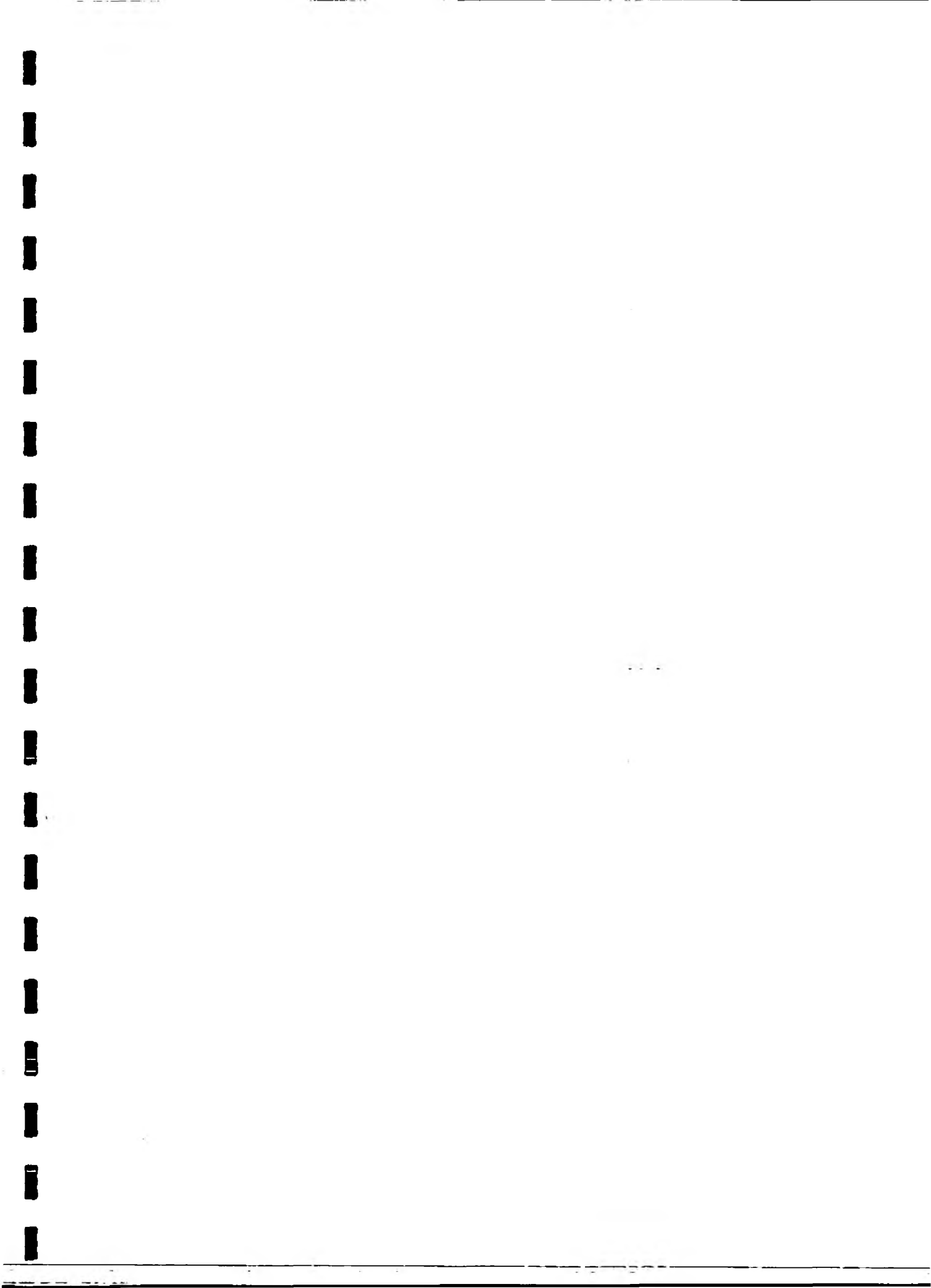
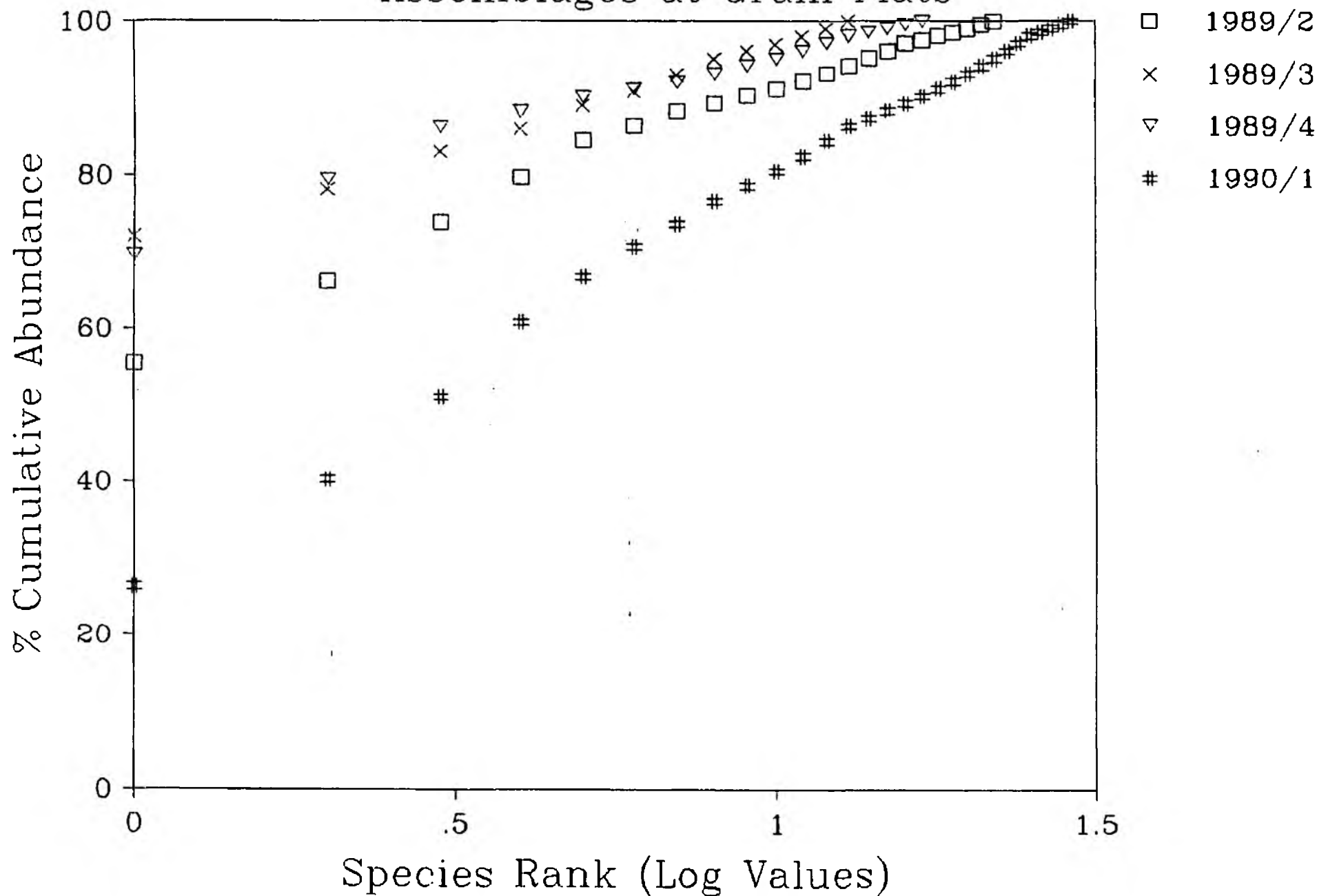


Figure 17. The dominance:diversity characteristics of the nematode assemblages observed at the Grain Flats sampling station between April 1989 and March 1990.

k-Dominance Curves for Nematode Assemblages at Grain Flats





respectively). The same pattern was also noted amongst the harpacticoid copepods although fewer species were observed (see below). The muddier conditions appear to correlate with periods of reduced flow of the Thames and Medway and may form part of an annual cycle.

The changes in species numbers may not be fully explained by the altered granulometry. Although muddy soft substrates support lower numbers of meiofaunal species than coarser grained sediments, they usually support higher total densities of individuals, especially in the marine environment. In the present study, the higher silt or silt/clay fraction sediments yielded 1,278 nematodes litre⁻¹ (2nd. survey) and 5,246 nematodes litre⁻¹ (3rd. survey). The sands of the 1st. and final surveys maintained 15,409 and 14,179 individuals litre⁻¹, respectively. This cannot be explained on existing data, especially as the densities of the nematode populations would be expected to increase over the summer months. The change in sediment type also correlated with a marked increase in dominance (Figure 17) with *S. punctata* accounting for over 70% of the nematode population during the 2nd. and 3rd. surveys.

Given its near marine location, the harpacticoid copepod assemblage of Grain Flats was depauperate with only 3 species being identified over the year (cf. 18 taxa at Stations 18 and 21). Acari were not observed in any of the samples examined. Densities and diversity of other invertebrate taxa (mostly marine species) were also lowest in the July - December surveys. The 1st. and 4th. surveys produced comparatively high densities of meiofaunal kinorhynchs, ostracods, turbellarians and tardigrades and several other, transitional meiofaunal groups such as polychaetes and bryozoans.

Station 21, Shoeburyness East

This site was second only to Chapman Buoy (Station 18) in the species richness of the principal meiofaunal groups that it supported. A total of 90 taxa (nematodes, harpacticoids and mites) was noted. The species composition of the meiofaunal assemblages was comparatively stable throughout the year, with percentage faunal similarities (presence:absence) of 38%, 44% and 51% between consecutive surveys and 47% between the 1st. and 4th. surveys. This stability is also reflected in the dominance-diversity characteristics of the Shoeburyness East nematode assemblages (Figure 18) where dominance varied between 27% and 39% only. The dominant species were all members of the Microlaimidae - type 2A species characteristic of coarser grained sediments. The feeding type ratios were accordingly lower than 1.00 throughout the year although several non-selective detritivore species, such as *S. punctata* and species of *Daptonema*, *Theristus* and *Ascolaimus*, were present at low densities. Total nematode densities were high for a fine sand and ranged from 37,108 litre⁻¹ (October - December) to 125,012 litre⁻¹ (January - March).

With the exception of *Bryocamptus* species, which is a freshwater species, the harpacticoids were all of marine or outer estuarine origin and typical of British waters. The *Arenostella* species noted in the 3rd. survey (October - December) is a small, vermiform species with reduced limbs which adopts a true interstitial existence. Species of this and

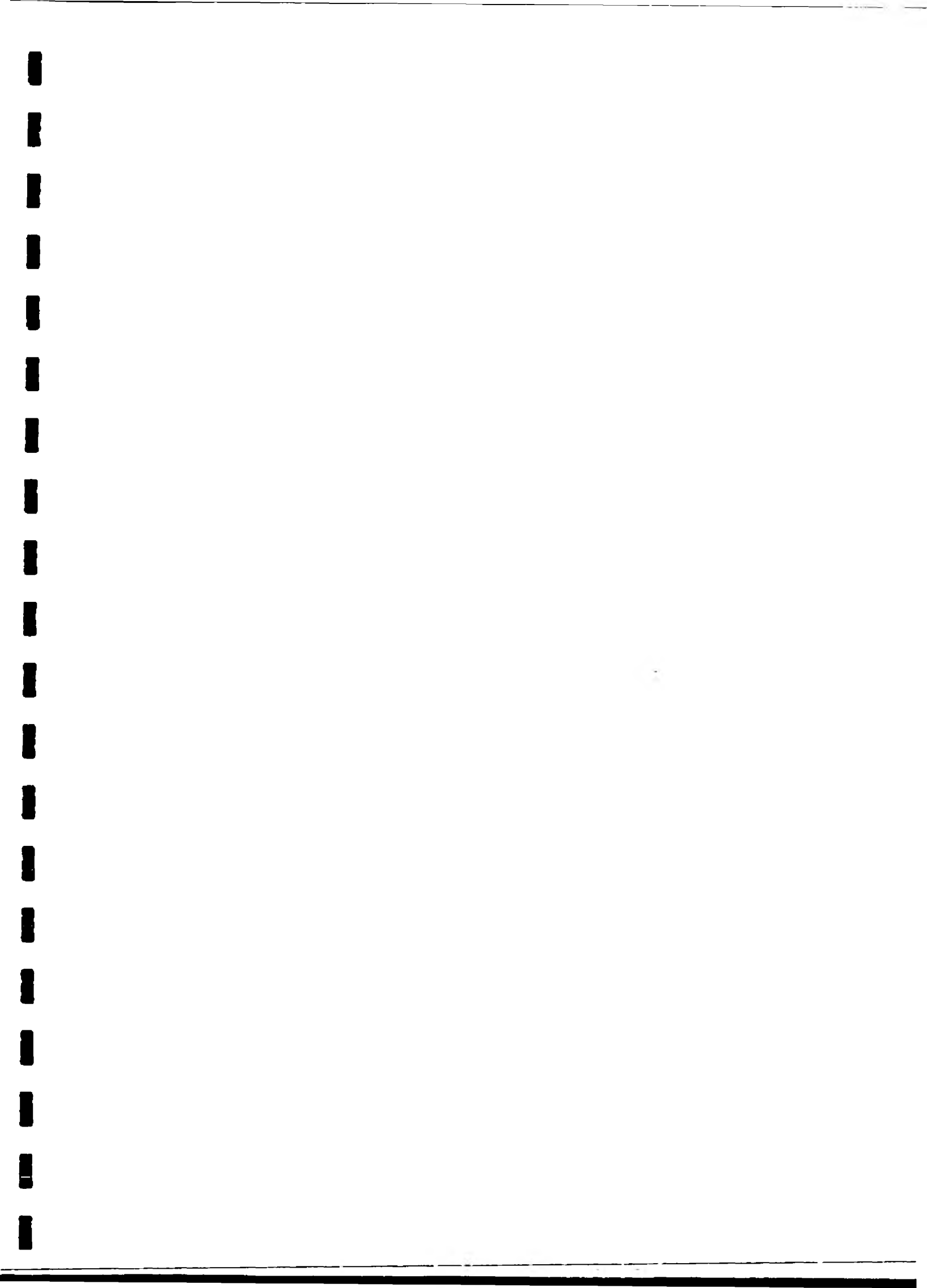
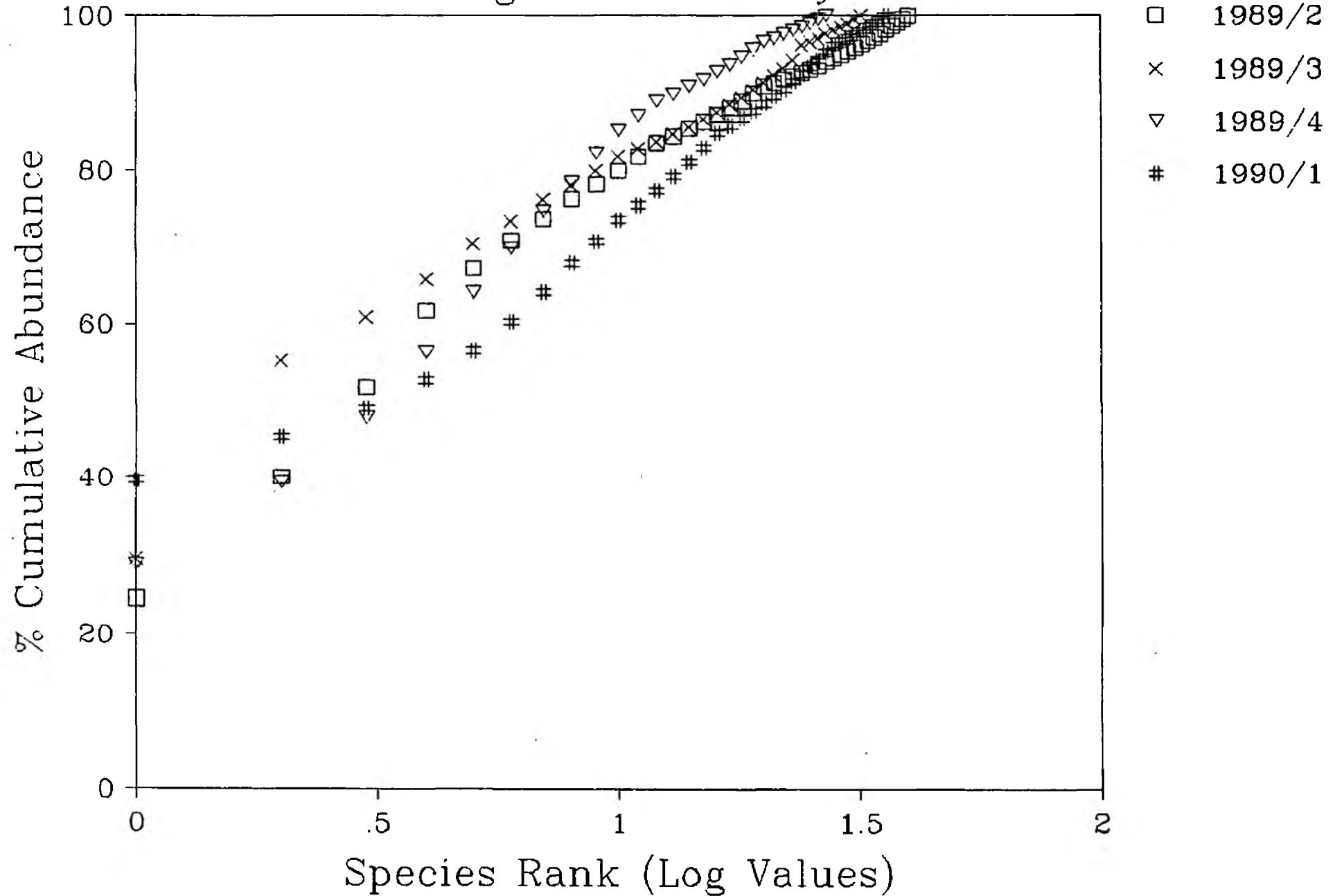


Figure 18. The dominance:diversity characteristics of the nematode assemblages observed at the Shoeburyness sampling station between April 1989 and March 1990.

k-Dominance Curves for Nematode Assemblages at Shoeburyness East





related genera are common in clean, well oxygenated sands and exceptionally high densities ($> 1,000 \text{ litre}^{-1}$) have been recorded in pollution surveys off the North Sea coast of France (Feil, pers. obsvn.).

Acarine species were not well represented at the Shoeburyness East site and a single unidentified halacarid mite was noted in the 2nd. survey only. In temperate waters, densities of halacarid mites are not often high in coarser sediments unless macro- or filamentous algae are also present. This occasionally occurs where fish farm effluent is discharged to coastal waters (Trett and Forster, pers. obsvn.). Other invertebrate phyla were abundant and included interstitial marine ciliates, larvae of littorinid gastropod molluscs and one of the few records of Gnathostomulida in this part of England. All groups were in keeping with a clean, marine sand habitat.

Station 22, Sea Reach No. 2 Buoy.

Potentially the most marine of the Tideway habitats surveyed, Sea Reach No. 2 Buoy yielded a variable assemblage of meiofaunal species. As at the Grain Flats site (Station 20) this may again have related to a partial change that was noted in sediment type; fine sands present in the 1st. survey were replaced by a muddier sand in subsequent surveys. Fifty-seven nematode taxa were recorded, 36 in the 1st. survey and between 15 and 17 in the following surveys. That this decline in species richness was related in part to substrate type may be indicated by the nematode feeding type ratio. This became greater than 1.00 in the samples examined from July 1989 onwards, indicating a greater relative importance of the non-specialist deposit feeding species as opposed to the highly specialist epipsammic and diatomivorous species. Unlike the Grain Flats nematode assemblages, the densities recorded were uniformly high throughout the year. Dominance also increased after the 1st. survey and remained high during the rest of the survey period (Figure 19). As at the Southend subtidal site, the dominant species in each of the surveys was *Richtersia inaequalis* which accounted for between 33 and 65% of the nematodes observed. As stated earlier (see Station 19), this probably indicates elevated levels of organic material in the sediments at this station.

Given its location, comparatively few harpacticoid copepod species were observed at Sea Reach No. 2 Buoy (total 8 taxa). Those present included the larger epibenthic, detritivorous species. Acari were observed at low densities in all the samples except that collected in July - September. Other invertebrate phyla were represented by several species at low densities in the 1st. survey and fewer species in subsequent surveys. Gastrotrichs were notable records at this site although very few were seen.

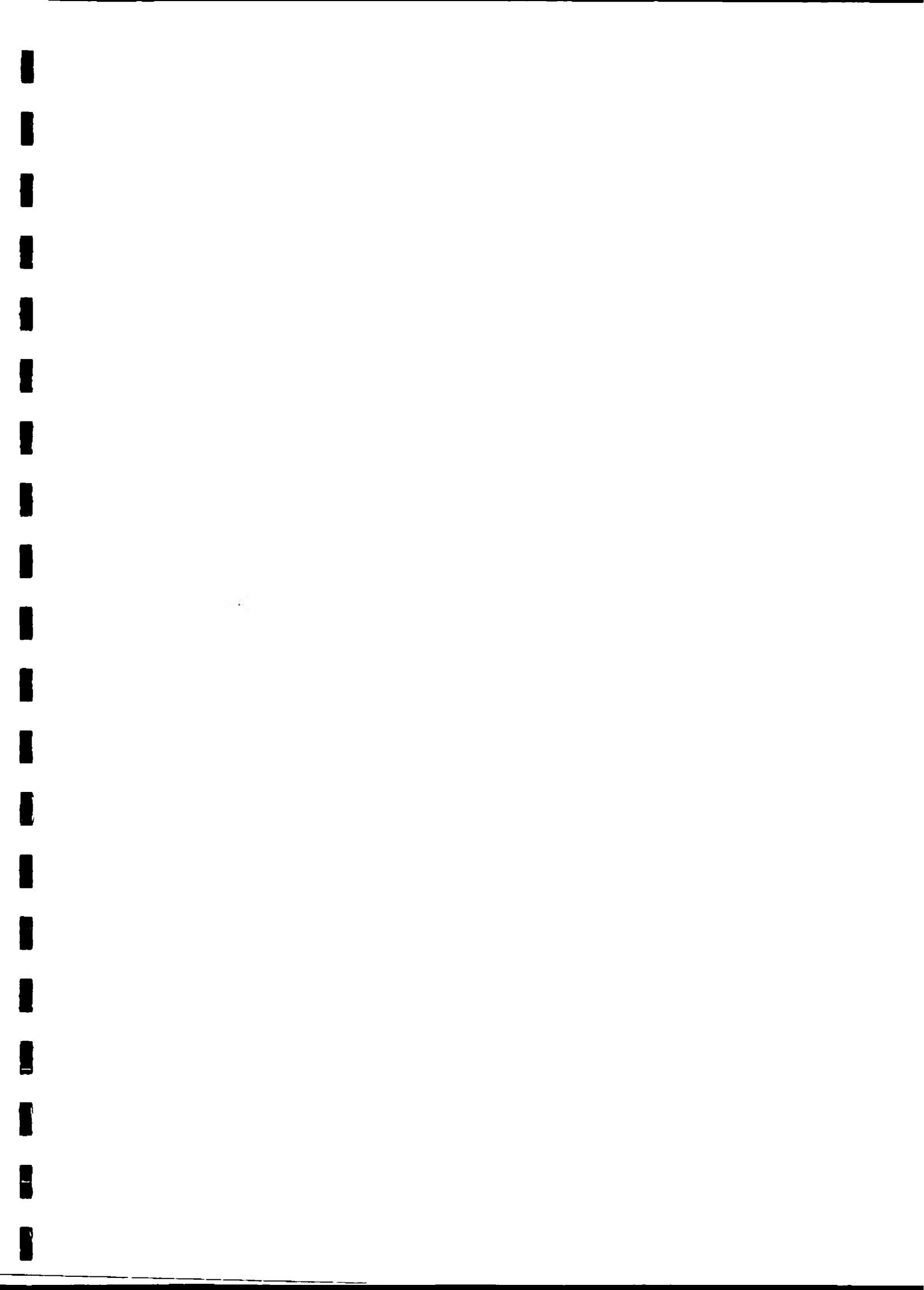
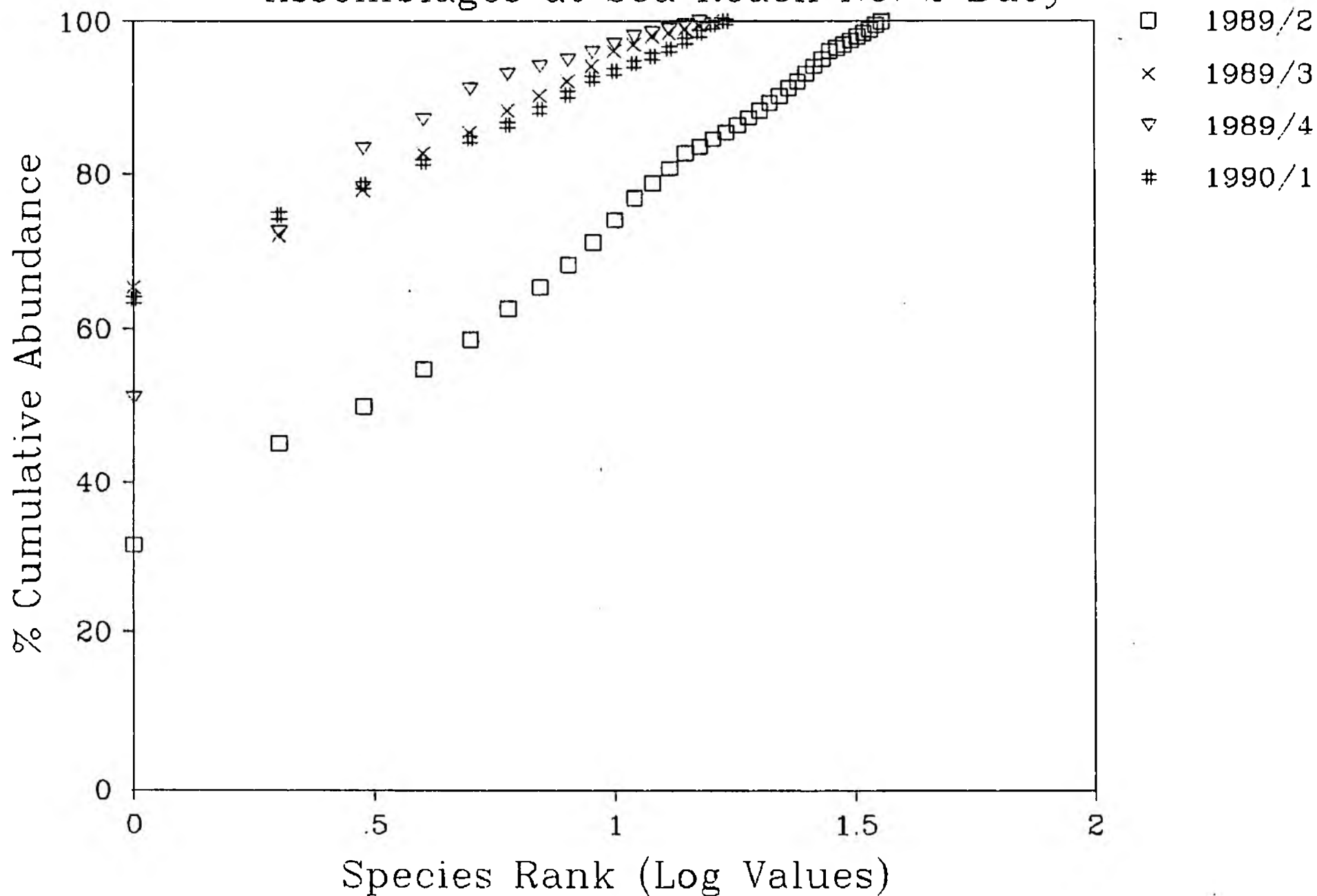


Figure 19. The dominance:diversity characteristics of the nematode assemblages observed at the Sea Reach No. 2 Buoy sampling station between April 1989 and March 1990.

k-Dominance Curves for Nematode

Assemblages at Sea Reach No. 2 Buoy



5. Acknowledgements

We would like to thank colleagues at *Physalia* and staff at the Centre for Research in Aquatic Biology (CRAB) for their assistance in the preparation of this study. Special thanks are due to Professor James Green, Dr. Judith Hutchinson, Dr. Philip Rainbow and Dr. Rob Hughes (CRAB) and Dr. Derek Tinsley and Dr. Martin Attrill (NRA, Thames Region) for their comments and observations on the Thames macrofauna. We are also grateful to Simon Forster (*Physalia*) who separated the meiofauna and prepared the material for the taxonomists; he also assisted in the enumeration of nematodes at more densely populated stations. Finally, we would like to thank Rebecca Marshall (*Physalia*) who processed the final manuscript.



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7. APPENDIX

Thames Estuary Survey

Sections 1 to 5

Meiofaunal Species Lists and Faunal Results Tables



Section 1

CUMULATIVE MEIOFAUNAL SPECIES LIST

THAMES ESTUARY

April 1989 - March 1990

COPEPODA

Copepoda: Harpacticoida

A. Ameiridae:

Leptomesochra macintoshi

B. Canthocamptidae:

Attheyella species

Bryocamptus (*Limnocamptus*) species (?*echinatus*)

Canthocamptus species

Elaphoidella gracilis

Epactophanes richardi

Moraria species

C. Canuellidae:

Canuella perplexa

D. Cletodidae:

Cletodes longicaudatus

Cletodid species

Enhydrosoma propinquum

E. Diosaccidae:

Amphiascella species

Amphiascoides species

Amphiascus angusticeps

Amphiascus species 1

Amphiascus species 2

Amphiascus species 3

Amphiascus varians

Bulbamphiascus species

Diosaccid copepodites

Paramphiascella species

Pseudomesochra species (?*latifurea*)

Schizopera clandestina

Stenhelia aemula

Stenhelia giesbrechti

Stenhelia palustris

Stenhelia species

Typhlamphiascus species

F. Ectinosomatidae:

Arenostella species
Ectinosoma melaniceps
Ectinosoma species
Halectinosoma curticorne
Halectinosoma herdmani
Halophytophilus species
Pseudobradyia brevicornis

G. Harpacticidae:

Harpacticella species

H. Laophontidae:

Laophonte species (?*denticornis*)
Onychocamptus species (?*bengalensis*)

I. Longipediidae:

Longipedia coronata
Longipedia species

J. Tachidiidae:

Microarthridion species
Tachidius discipes
Tachidius species

K. Thalestridae:

Idomene forficata
Phyllothalestris species

L. Tisbidae:

Tisbe species

M. Unascribed specimens:

Unidentified copepodites

NEMATODA

i. Adenophorea: Araeolaimida

A. Plectidae:

Plectid species (?Paraplectonema)
Plectus granulatus

ii. Adenophorea: Enoplida

A. Alaimidae:

Alaimus species

B. Anoplostomatidae:

Anoplostoma viviparum
Chaetonema riemanni

C. Anticomminidae:

Anticoma acuminata

D. Cryptonchidae:

Cryptonchus species

E. Enchelidiidae:

Belbolla teisseiri
Calyptonema maxweberi
Chaetonema riemanni
Eurystomina species

F. Enoplidae:

Enoplus brevis
Enoplus communis

G. Ironidae:

Ironus ignavus

H. Leptosomatidae:

Pseudocella coecum

I. Oncholaimidae:

Adoncholaimus fuscus
Adoncholaimus thalassophygas
Oncholaimellus calvadosicus
Oncholaimid species 1
Oncholaimid species 2 (?Viscosia)
Oncholaimus brachycercus
Oncholaimus campylocercoides
Oncholaimus oxyuris
Oncholaimus skawensis
Viscosia cobbi
Viscosia elegans
Viscosia glabra
Viscosia species
Viscosia viscosa

J. Oxystominidae:

Halalaimus capitulatus
Halalaimus isaitshikovi
Halalaimus longicaudatus
Nemanema cylindratICAUDATUM
Oxystomina asetosa
Oxystomina elongata
Thalassoalaimus tardus

K. Pristomatolaimidae:

Pristomatolaimus species (?verrucosus)
Pristomatolaimus stenurus

L. Thoracostomopsidae:

Enoploides brunettii
Enoplolaimus vulgaris
Mesacanthion diplochma

M. Tripyloididae:

Bathylaimus capacosus
Tobrilus gracilis
Tobrilus species
Tripyla affinis
Tripyloides marinus
Tripyloides species

iii. Adenophorea: Chromadorida

A. Aegialoalaimidae:

Aegialoalaimid species (?*Aegialoalaimus*)

B. Ceramonematidae:

Dasynemella species
Ceramonematid species

C. Chromadoridae:

Chromadora macrolaima
Chromadorella species
Chromadorid species
Chromadorina viridis
Chromadorita leuckarti
Chromadorita species
Chromadorita tentabunda
Dichromadora cephalata
Dichromadora geophila
Dichromadora species (?*cucullata*)
Euchromadora vulgaris
Hypodontolaimus balticus
Hypodontolaimus inaequalis
Innocuonema species
Neochromadora poecilosoma
Neochromadora tricophora
Prochromadora species (?*orleji*)
Prochromadorella attenuata
Prochromadorella ditlevseni
Ptycholaimellus ponticus
Punctodora species
Spilophorella candida
Spilophorella paradoxa

D. Comesomatidae:

Comesomatid species
Sabatieria breviseta
Sabatieria celtica
Sabatieria longisetosa
Sabatieria punctata
Setosabatieria species (?*hilarula*)

E. Cyatholaimidae:

Cyatholaimid species 1 (?*Cyatholaimus*)
Cyatholaimid species 2
Cyatholaimus species (?*gracilis*)
Marylynnia species
Paracanthonchus caecus
Paracanthonchus heterodontus
Paracanthonchus longus
Paracanthonchus species
Paracyatholaimus intermedius
Paralongicyatholaimus species
Pomponema species
Praeacanthonchus opheliae
Praeacanthonchus species

F. Desmodoridae:

Desmodora communis
Desmodora species
Desmodorid species 1 (?*Chromaspirina*)
Desmodorid species 2
Metachromadora remanei
Metachromadora scotlandica
Metachromadora species
Metachromadora suecica
Molgolaimus cuanensis
Onyx perfectus
Pseudonchus species
Sigmophoranema rufum
Spirinia parasitifera

G. Desmoscolecidae:

Desmoscolex falcatus
Pareudesmoscolex species
Quadricoma species

H. Leptolaimidae:

Antomicron elegans
Camacolaimus barbatus
Camacolaimus tardus
Chronogaster species
Deontolaimus species
Leptolaimid species
Leptolaimoides species
Leptolaimus papilliger
Leptolaimus species 1 (?*ampullaceus*)
Leptolaimus species 2 (?*limicolus*)

Leptolaimus species 3
Onchium conicaudatus
Stephanolaimus jayassrei
Stephanolaimus species (?*spartinae*)

I. Microlaimidae:

Calomicrolaimus honestus
Microlaimus conothesis
Microlaimus globiceps
Microlaimus marinus
Microlaimus robustidens

J. Monoposthiidae:

Monoposthia costata
Monoposthia mirabilis

K. Selachinematidae:

Gammanema rapax
Halichoanolaimus robustus
Richtersia inaequalis

iv. Adenophorea: Monhysterida

A. Axonolaimidae:

Ascolaimus elongatus
Axonolaimus paraspinosus
Axonolaimus species
Odontophora setosa
Odontophora villoti

B. Coninckidae:

Coninckia species

C. Diplopeltidae:

Campylaimus species (?*inaequalis*)
Diplopeltid species (?*Diplopeltula*)

D. Linhomoeidae:

Desmolaimus zeelandicus
Eleutherolaimus species
Linhomoeid species 1 (?*Terschellingia*)
Linhomoeid species 2

Linhomoeid species 3
Linhomoeus species 1
Linhomoeus species 2
Terschellingia longicaudatus

E. Monhysteridae:

Diplolaimella ocellata
Monhystera filicaudata
Monhystera disjuncta
Monhystera species
Monhystera stagnalis
Monhystera vulgaris
Monhysterid species 1
Monhysterid species 2

F. Sphaerolaimidae:

Parasphaerolaimus species (??*paradoxa*)
Sphaerolaimus balticus
Sphaerolaimus gracilis

G. Xyalidae:

Daptonema furcata
Daptonema normandica
Daptonema setosa
Daptonema tenuispiculum
Daptonema species 1
Daptonema species 2
Linhystera species
Paramonhystera species
Theristus acer
Theristus species 1
Theristus species 2
Theristus species 3
Xyalid species 1
Xyalid species 2

v. Adenophorea: Trefusida

A. Trefusiidae:

Halanonchus species
Rhabdocoma riemanni
Trefusia longicaudata
Trefusia zostericola
Trefusiid species

vi. Adenophorea: Dorylaimida

A. Dorylaimidae:

Dorylaimid species 1 (?*Dorylaimus*)
Dorylaimid species 2
Dorylaimid species 3
Labronema species

B. Mononchidae:

Iotonchus species
Mononchus aquaticus

vii. Secernentea: Rhabditida

A. Cephalobidae:

Acrobeles species
Cephalobus species

B. Diplogasteridae:

Butlerius butleri
Diplogaster species
Diplogasterid species
Paroigolaimella bernensis

C. Panagrolaimidae:

Panagrolaimus species

D. Rhabditidae:

Diploscapter coronata
Mononchoides species
Mononchoides striatus
Panagrellus species
Rhabditid species 1
Rhabditid species 2
Rhabditis species

viii. Secernentea: Tylenchida

A. Criconematidae:

Criconema species
Criconemoides species
Macroposthonia species

B. Heteroderidae:

Heteroderid species (?*Globodera*)

C. Hoplolaimidae:

Hirschmanniella species

D. Tylenchidae:

Tylenchid species 1
Tylenchid species 2
Tylenchid species 3

ix. Unascribed Species

Miscellaneous unidentified specimens

ACARINA

i. Prostigmata

A. Bdellidae:

Bdellid species

B. Oribatidae:

Oribatid species

ii. Cryptostigmata

A. Hydracarina:

Hydracarine species

B. Halacaridae:

Copidognathus dentatus
Copidognathus rhodostigma
Unidentified species



Section 2

QUARTERLY MEIOFAUNAL RESULTS LISTS

April 1989 - March 1990

Station TW1

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Diplogaster</i> sp.	376			
<i>Tobrilus gracilis</i>	1953	7850	2300	85
<i>Monhystera stagnalis</i>	901	1926	1054	36
<i>Labronema</i> sp.	676			
<i>Dichromadora geophila</i>	1803			
<i>Chromadorella</i> sp.	300			
<i>Chromadorina viridis</i>	676		144	24
<i>Hirschmanniella</i> sp.	150			24
<i>Tripyla affinis</i>	75	741		
Dorylaimid sp. 1	+	148	48	24
<i>Monhystera vulgaris</i>		1185		
<i>Paroigrolaimellus bernensis</i>		592	383	
<i>Mononchoides</i> sp.		444		
<i>Tobrilus</i> sp.		592		12
<i>Mononchus aquaticus</i>		296		85
Tylenchid sp. 1		148		24
Dorylaimid sp. 2		148		24
<i>Ironus ignavus</i>		296		12
<i>Paracanthonchus</i> sp.		148	48	
<i>Paracyatholaimus intermedius</i>				278
Tylenchid sp. 2				24
<i>Daptonema normandica</i>				109
Chromadorid sp. 1				12
<i>Plectus granulatus</i>				85
<i>Criconema</i> sp.				24
<i>Chromadorita leuckarti</i>				121
<i>Monhystera filicaudata</i>				109
Heteroderid sp.				12
Rhabditid sp. 1				12
<i>Daptonema setosa</i>				12
<i>Cephalobus</i> sp.				73
<i>Iotonchus</i> sp.				24
Dorylaimid sp. 3				12
<i>Panagrolaimus</i> sp.				12
Indet.	1502	889	383	206
1B:2A	1.03	57.00	12.00	0.69
N	8402	15847	5127	1560
S	12	14	7	26

continued ...

Station TW1 continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Copepoda				
<i>Pseudomesochra</i> sp.	242			
Diosaccid copepodites	88			
<i>Idomene forficata</i>	66			
<i>Tachidius</i> sp.	44			
<i>Laophonte</i> sp.	22			
<i>Canthocamptus</i> sp.	159			
<i>Moraria</i> sp.	22			
<i>Elaphoidella gracilis</i>	67			
<i>Bryocamptus</i> sp.			8	
<i>Onychocamptus</i> sp.			92	
<i>Attheyella</i> sp.				30
N	573	0	100	30
S	8	0	2	1
Acari				
Bdellid sp.				10
Oribatid sp.				10
N	0	0	0	20
S	0	0	0	2

Station TW2

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Tobrilus gracilis</i>	1249	2448	284	601
<i>Diplogaster</i> sp.	187			
<i>Paracanthonus</i> sp.	156			
Monhysterid sp. 1	344		417	
<i>Tripyla affinis</i>	187		19	+
<i>Mononchoides striatus</i>	94			63
<i>Monhystera stagnalis</i>	250	306	19	158
<i>Ironus ignavus</i>	94	31	19	32
Xyalid sp. 1	625			
Dorylaimid sp. 1	31			63
<i>Plectus granulatus</i>	31	31		63
<i>Chromadorina viridis</i>	31	+		
<i>Dichromadora geophila</i>	+	61		32
<i>Mononchus aquaticus</i>	+	31		
<i>Viscosia</i> sp.	+			
Rhabditid sp.	+		19	63
<i>Chromadorella</i> sp.	+			
<i>Tobrilus</i> sp.		61		32
Heteroderid sp.		31		32
<i>Punctodora</i> sp.		31		
<i>Mononchoides</i> sp.		31		
<i>Leptolaimus</i> sp. 1			57	
<i>Diplolaimella ocellata</i>			38	
<i>Paroigrolaimellus bernensis</i>			95	221
<i>Theristus</i> sp. 1			38	
<i>Butlerius butleri</i>			95	475
Diplogasterid sp.			76	
<i>Theristus</i> sp. 2			19	
<i>Daptonema normandica</i>			474	601
<i>Microlaimus globiceps</i>			38	
<i>Diploscapter coronata</i>			19	
<i>Daptonema setosa</i>				538
<i>Hirschmanniella</i> sp.				32
Monhysterid sp. 2				127
<i>Monhystera filicaudata</i>				32
Indet.	248	153	303	791
1B:2A	11.50	20.25	6.14	21.67
N	3527	3215	2200	3956
S	17	11	16	18

continued ...

Station TW2 continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<hr/>				
Copepoda				
<i>Schizopera clandestina</i>	8			
<i>Idomene forficata</i>	8			
Copepodites		17		
<i>Bryocamptus</i> sp.				73
N	16	17	0	73
S	2	1	0	1
Acari				
Bdellid sp.				27
Oribatid sp.				9
N	0	0	0	36
S	0	0	0	2

Station TW3

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Daptonema</i> sp. 2	36			
Xyalid sp. 2	36			
Monhysterid sp. 1	9			
<i>Daptonema</i> sp. 1	18	12		
<i>Ascolaimus elongatus</i>	18		10	
<i>Paracanthochus</i> sp.	18	12	10	
<i>Microlaimus globiceps</i>	9		20	
<i>Monhystera stagnalis</i>	45		100	
<i>Chromadorina viridis</i>	18			
<i>Rhabditis</i> sp. 1	9			
<i>Desmodora communis</i>	18			
Dorylaimid sp. 1	9		10	
<i>Metachromadora</i> sp.	9			
<i>Chromadorella</i> sp.	9			
Trefusiid sp.	63			
<i>Onchium conicaudatus</i>		12		
<i>Monhystera vulgaris</i>		12		
<i>Oncholimus campyloceroides</i>		12		
Diplogasterid sp.			10	
<i>Daptonema setosa</i>			350	173
<i>Axonolaimus paraspinosus</i>			10	
<i>Odontophora villosi</i>			10	
<i>Butlerius butleri</i>			50	
<i>Leptolaimus</i> sp. 1			20	
<i>Daptonema furcata</i>			10	
<i>Anoplostoma viviparum</i>			10	
<i>Mononchus aquaticus</i>			10	
Rhabditid sp. 1			10	
<i>Theristus</i> sp 1			10	
<i>Dichromadora geophila</i>			20	9
<i>Mononchoides striatus</i>			40	9
<i>Chromadorita leukarti</i>				9
<i>Plectus granulatus</i>				9
<i>Sabatieria punctata</i>				9
<i>Tobrilus</i> sp.				9
<i>Hirschmanniella</i> sp.				9

continued ...

Station TW3 continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<i>Monhystera filicaudata</i>				18
<i>Tobrilus gracilis</i>				9
Chromadorid sp. 1				9
Monhysterid sp. 2				9
<i>Viscosia</i> sp.				9
<i>Criconemoides</i> sp.				9
<i>Paracyatholaimus intermedius</i>				9
<i>Daptonema normandica</i>				18
Indet.	63	0	40	55
1B:2A	1.44	1.00	3.08	5.00
N	333	72	720	381
S	15	6	18	16
Copepoda				
<i>Attheyella</i> sp.				9
N	0	0	0	9
S	0	0	0	1
Acari				
Hydracarine sp.	9	12		
Oribatid sp.			10	
N	9	12	10	0
S	1	1	1	0

Station TW4

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Monhystera stagnalis</i>	475		53	178
<i>Monhystera filicaudata</i>	850	150		200
<i>Mononchoides striatus</i>	25			44
<i>Monhystera vulgaris</i>	125	365		
<i>Mononchus aquaticus</i>	25	43		22
Dorylaimid sp. 1	125	21		
<i>Tobrilus gracilis</i>	125			445
<i>Diploscapter coronata</i>	25			
<i>Plectus granulatus</i>	25			
<i>Paracanthonchus</i> sp.	75		53	
<i>Daptonema furcata</i>	25	1397	214	
Rhabditid sp. 1	25	21		
<i>Daptonema normandica</i>	100	21		89
<i>Tripyla affinis</i>	25			
<i>Anoplostoma viviparum</i>		21	80	
<i>Diplolaimella ocellata</i>		21	187	22
<i>Paroigrolaimellus bernensis</i>		21		89
<i>Chromadorina viridis</i>		21		22
<i>Butlerius butleri</i>		+		
<i>Daptonema setosa</i>			1148	467
Diplogasterid sp.			160	
Monhysterid sp. 1			160	
<i>Innocuonema</i> sp			27	
<i>Tripyloides gracilis</i>			53	44
<i>Theristus</i> sp. 1			53	133
<i>Chromadora macrolaima</i>			80	
<i>Leptolaimus</i> sp. 1			107	
<i>Axonolaimus paraspinosus</i>			27	
<i>Adoncholaimus thalassophygus</i>			53	22
<i>Leptolaimus papilliger</i>			27	
<i>Ascolaimus elongatus</i>			133	
<i>Dichromadora geophila</i>			27	89
<i>Desmolaimus zeelandicus</i>			27	
<i>Sphaerolaimus gracilis</i>			+	
<i>Tobrilus</i> sp.				44
<i>Rhabditis</i> sp. 1				89

continued ...

Station TW4 continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Tylenchid sp. 1				44
Monhysterid sp. 2				44
<i>Paracyatholaimus intermedius</i>				44
Dorylaimid sp. 3				22
Plectid sp.				22
<i>Halalaimus isaitshikovi</i>				22
<i>Microaimus globiceps</i>				22
Indet.	50	21	27	422
1B:2A	3.00	34.50	3.59	6.00
N	2050	2166	2696	2641
S	14	13	20	23
Copepoda				
<i>Microarthridion</i> sp.				46
N	0	0	0	46
S	0	0	0	1
Acari	None observed			

Station TW5	Q U A R T E R			
	Species	1989/2	1989/3	1989/4
Nematoda				
Xyalid sp. 1	44	13		
<i>Monhystera stagnalis</i>	22			9
<i>Daptonema</i> sp. 1	11			
<i>Paracanthonus</i> sp.	33		10	
<i>Plectus granulatus</i>	22			
<i>Mononchoides striatus</i>	11			
<i>Theristus</i> sp. 2	22			
<i>Butlerius butleri</i>	11			
<i>Daptonema furcata</i>		50	10	
<i>Anticoma acuminata</i>		13		
<i>Tripyloides gracilis</i>		13		
<i>Axonolaimus paraspinosus</i>			31	
<i>Sabatieria punctata</i>			31	9
<i>Daptonema setosa</i>			94	291
<i>Innocuonema</i> sp.			10	
<i>Diplolaimella ocellata</i>			10	
<i>Ptycholaimellus ponticus</i>			10	
<i>Chromadora macrolaima</i>			10	
<i>Chromadorita</i> sp.			52	
<i>Dichromadora geophila</i>			42	
<i>Odontophora setosa</i>			10	
<i>Adoncholaimus thalassophygas</i>			10	
Diplogasterid sp.			10	
<i>Viscosia</i> sp.				9
<i>Chronogaster</i> sp.				9
Indet.	11	0	10	27
1B:2A	3.00	x2A	1.07	x2A
N	187	102	350	354
S	8	5	14	5
Copepoda None observed				
Acari				
Oribatid sp.	11			
<i>Copidognathus dentatus</i>				10
N	11	0	0	10
S	1	0	0	1

Station TW6

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Microlaimus robustidens</i>	9			
<i>Monhystera filicaudata</i>	43	1136		364
<i>Prochromadora</i> sp.	52			
Xyalid sp. 1	17			
Cyatholaimid sp. 1	26			
<i>Plectus granulatus</i>	9			
<i>Daptonema setosa</i>	17		154	1405
<i>Diplolaimella ocellata</i>	26	120	19	52
<i>Monhystera disjuncta</i>	9			
<i>Theristus</i> sp. 2	26			
<i>Tobrilus gracilis</i>	9			
<i>Daptonema furcata</i>		538		
<i>Dichromadora geophila</i>		359	346	260
<i>Daptonema normandica</i>		3827	788	104
<i>Deontolaimus</i> sp.		60		
Dorylaimid sp. 2		60		
<i>Paracanthonus heterodontus</i>		120	58	
<i>Axonolaimus paraspinosus</i>		60		
<i>Monhystera vulgaris</i>		60		
Trefusiid sp.		60		
<i>Chromadorina viridis</i>		60		
<i>Halalaimus isaithshikovi</i>		60		260
<i>Mononchus aquaticus</i>		+		
<i>Tobrilus gracilis</i>		+		
<i>Leptolaimus papilliger</i>		+	19	52
<i>Oxystomina asetosa</i>		+		
<i>Tripyloides</i> sp.		+		156
<i>Anoplostoma viviparum</i>			77	
<i>Adoncholaimus thalassophygass</i>			38	52
<i>Sabatieria punctata</i>				58
Diplogasterid sp.			269	
<i>Leptolaimus</i> sp. 1			58	
Monhysterid sp. 1			19	
<i>Sphaerolaimus gracilis</i>			19	
<i>Ascolaimus elongatus</i>			19	
<i>Rhabditis</i> sp. 1				52

continued ...

Station TW6 continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<i>Paracyatholaimus intermedius</i>				1509
<i>Microaimus globiceps</i>				364
<i>Chromadorita leukarti</i>				104
<i>Mononchoides striatus</i>				104
<i>Monhystera stagnalis</i>				104
<i>Criconema</i> sp.				52
<i>Panagrolaimus</i> sp.				52
Plectid sp.				52
Tylenchid sp. 3				+
Indet.	43	120	115	468
1B:2A	0.88	8.22	1.66	0.78
N	286	6640	2056	5670
S	11	19	14	20
Copepoda				
<i>Bryocamptus</i> sp.	42			
N	42	0	0	0
S	1	0	0	0
Acari None observed				

Station TW7s	Q U A R T E R				
	Species	1989/2	1989/3	1989/4	1990/1
Nematoda					
<i>Daptonema furcata</i>		486			10
<i>Rhabditis</i> sp.		29			
<i>Monhystera filicaudata</i>		29			
<i>Adoncholaimus thalassophygas</i>		14		33	40
<i>Theristus acer</i>		14			
<i>Paracanthonchus caecus</i>		14			
<i>Daptonema setosa</i>		14		11	80
<i>Mononchoides striatus</i>		14			
<i>Microlaimus robustidens</i>		14			
<i>Daptonema</i> sp. 1				11	
<i>Diplolaimella ocellata</i>				22	140
<i>Sabatieria punctata</i>				11	
<i>Ascolaimus elongatus</i>				22	
<i>Chromadorita</i> sp.				67	
<i>Leptolaimus papilliger</i>				33	20
<i>Dichromadora geophila</i>				11	10
<i>Axonolaimus paraspinosus</i>				11	
<i>Paracanthonchus heterodontus</i>				11	10
<i>Anoplostoma viviparum</i>				11	10
<i>Ptycholaimellus ponticus</i>					10
<i>Paracyatholaimus intermedius</i>					130
<i>Paroigrolaimellus bernensis</i>					60
<i>Halalaimus isaitshikovi</i>					30
<i>Hypodontolaimus balticus</i>					10
<i>Microlaimus globiceps</i>					20
<i>Monhystera disjuncta</i>					40
<i>Plectid</i> sp.					10
<i>Daptonema normandica</i>					10
Indet.	-	14		0	100
1B:2A	-	12.00		0.88	0.95
N	-	656		254	750
S	-	10		12	18

continued ...

Station TW7s continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<hr/>				
Copepoda				
<i>Microarthridion</i> sp.			11	
N	0	0	11	0
S	0	0	1	0

Acari None observed

Station TW7i

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Diploscapter coronata</i>	17			
<i>Butlerius butleri</i>	8			
<i>Monhystera disjuncta</i>	17			8
<i>Oncholaimellus calvadosicus</i>	8		13	
<i>Rhabditis</i> sp.	17			8
<i>Plectus granulatus</i>	25			
Tylenchid sp. 1	8			
Diplogasterid sp.	8		40	8
Heteroderid sp.	8			
<i>Dichromadora geophila</i>	8		13	42
<i>Axonolaimus paraspinosus</i>	8			
Tylenchid sp.	8			
<i>Metachromadora suecica</i>	8			
Xyalid sp. 1	8			
<i>Monhystera filicaudata</i>	8	216		17
<i>Cryptonchus</i> sp.	8			
<i>Diplolaimella ocellata</i>		130	13	42
<i>Paracanthonchus heterodontus</i>		86	93	
<i>Daptonema setosa</i>		3285	652	108
<i>Theristus acer</i>		43		
<i>Sabatieria punctata</i>		86	106	8
<i>Anoplostoma viviparum</i>		86	133	33
<i>Leptolaimus papilliger</i>		130	93	25
<i>Microlaimus globiceps</i>		130		8
<i>Viscosia elegans</i>		43		
<i>Tripyloides</i> sp.		43		8
<i>Dichromadora cephalata</i>		43		
<i>Prochromadorella attenuata</i>		+		
<i>Adoncholaimus thalassophygus</i>		+	133	8
<i>Sphaerolaimus gracilis</i>			13	
<i>Monhystera</i> sp. 2			40	
<i>Antomicron elegans</i>			13	
<i>Microlaimus marinus</i>			27	
<i>Halalaimus isaitshikovi</i>			+	
Leptolaimid sp. 1				8
<i>Tripyla affinis</i>				8

continued ...

Station TW7i continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<i>Daptonema normandica</i>				25
<i>Paracyatholaimus intermedius</i>				17
Tylenchid sp. 3				8
<i>Tobrilus gracilis</i>				25
<i>Mononchoides striatus</i>				8
<i>Viscosia</i> sp.				25
<i>Neochromadora poecilosoma</i>				8
Monhysterid sp. 2				25
<i>Panagrellus</i> sp.				8
<i>Chromadorita leukarti</i>				8
Indet.	42	0	13	42
1B:2A	4.00	13.50	5.23	2.25
N	214	4321	1409	538
S	16	14	15	25
Copepoda				
<i>Stenhelia giesbrechti</i>	44			
<i>Microarthridion</i> sp.			3068	
<i>Harpacticella</i> sp.			741	
<i>Stenhelia palustris</i>			423	
Copepodites			317	
N	44	0	4549	0
S	1	0	4	0
Acari				
Oribatid sp.	16			8
N	16	0	0	0
S	1	0	0	0

Station TW8s

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Sabatieria punctata</i>	25	14		191
<i>Dichromadora geophila</i>		800	29	
<i>Adoncholaimus thalassophygas</i>		43	43	9
<i>Daptonema furcata</i>		143		
<i>Theristus acer</i>		14		
<i>Leptolaimus papilliger</i>		14	29	
<i>Monhystera disjuncta</i>		14		
<i>Daptonema normandica</i>		14	14	
<i>Anoplostoma viviparum</i>		29		9
<i>Daptonema setosa</i>			29	18
<i>Sabatieria breviseta</i>			14	
<i>Oncholaimuellus calvadosicus</i>			29	
Monhysterid sp. 1			14	
<i>Diplolaimella ocellata</i>			43	
<i>Acrobeles</i> sp.			14	
<i>Monhystera filicaudata</i>			29	
<i>Hypodontolaimus inaequalis</i>				9
Indet.	0	0	0	0
1B:2A	x2A	0.27	2.00	24.00
N	25	1085	287	236
S	1	9	11	5
Copepoda				
Copepodites				
			29	
N	0	0	29	0
S	0	0	1	0
Acari				
Oribatid sp.				
		22		
N	0	22	0	0
S	0	1	0	0

Station TW8i

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Dichromadora geophila</i>	152	128	212	
<i>Diplolaimella ocellata</i>	32			
<i>Monhystera filicaudata</i>	72			
Tylenchid sp. 3	8			
Monhysterid sp. 2	32			
<i>Sabatieria punctata</i>	8			
<i>Monoposthia mirabilis</i>	8			
<i>Microlaimus robustidens</i>	8			
<i>Mononchoides</i> sp.	24			
<i>Daptonema setosa</i>	312	3978	6574	8
<i>Chromadorella</i> sp.	8			
Oncholaimid sp. 2	16			
Cyatholaimid sp. 1	16			
<i>Chromadorina viridis</i>	16			
<i>Desmoscolex falcatus</i>	8			
<i>Microlaimus globiceps</i>	8			
<i>Chromadorita</i> sp.	16			
<i>Criconema</i> sp.	8			
<i>Anoplostoma viviparum</i>	8	26	+	
<i>Daptonema</i> sp. 1	8			
<i>Plectus granulatus</i>	8			
<i>Theristus</i> sp. 2	8			
<i>Axonolaimus paraspinosus</i>	8			
<i>Paracanthonchus heterodontus</i>		43	71	
<i>Theristus acer</i>		43		
<i>Adoncholaimus thalassophygass</i>		+	+	
<i>Prochromadorella attenuata</i>		+		
<i>Leptolaimus papilliger</i>			141	
Diplogasterid sp.			71	
<i>Daptonema furcata</i>				17
<i>Richtersia inaequalis</i>				8
<i>Daptonema normandica</i>				8
Indet.	32	0	0	8
1B:2A	1.55	24.00	18.60	x2A
N	824	4278	7069	49
S	23	7	7	4

continued ...

Station TW8i continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Copepoda				
<i>Pseudomesochra latifurea</i>	8			
<i>Stenhelia palustris</i>	24		29	
<i>Stenhelia giesbrechti</i>		32		
<i>Microarthridion</i> sp.			86	17
Copepodites			14	
N	32	32	129	17
S	2	1	3	1
Acari				
Oribatid sp.	16	22		
N	16	22	0	0
S	1	1	0	0

Station TW9

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Rhabditis</i> sp. 1	8		25	
<i>Daptonema furcata</i>		20		
Monhysterid sp. 1		10		
<i>Mononchoides striatus</i>		10		
<i>Butlerius butleri</i>			63	
<i>Prycholaimellus ponticus</i>			50	
<i>Anoplostoma viviparum</i>			13	55
<i>Leptolaimus papilliger</i>			13	
<i>Leptolaimoides</i> sp.			13	
<i>Monhystera</i> sp. 1			13	
<i>Enoplolaimus vulgaris</i>			13	
<i>Daptonema setosa</i>				82
<i>Sabatieria punctata</i>				27
<i>Paroigrolaimus</i> sp.				+
Indet.	0	0	50	18
1B:2A	x1B/x2A	2.00	0.25	x2A
N	8	40	253	191
S	1	3	8	4
Copepoda				
Cletodid sp.				9
N	0	0	0	9
S	0	0	0	i
Acari				
Oribatid sp.		10		
N	0	10	0	0
S	0	1	0	0

Station TW10s

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Daptonema setosa</i>	400			11
<i>Sabatieria punctata</i>	112	63	10	267
<i>Daptonema furcata</i>		100		
<i>Dichromadora geophila</i>		38		11
<i>Leptolaimus papilliger</i>		13		
<i>Oncholaimid sp. 2</i>		13		
<i>Paracanthonchus heterodontus</i>		13		
<i>Monhystera filicaudata</i>		13		
<i>Daptonema sp. 1</i>		13		
<i>Adoncholaimus thalassophygas</i>			19	11
<i>Monhystera disjuncta</i>				11
<i>Richtersia inaequalis</i>				11
<i>Anoplostoma viviparum</i>				78
<i>Camacolaimus barbatus</i>				11
<i>Diplolaimella ocellata</i>				22
<i>Hypodontolaimus balticus</i>				11
<i>Ironus ignavus</i>				11
Diplogasterid sp.				11
Indet.	8	13	0	11
1B:2A	x2A	3.50	x2A	8.25
N	520	279	29	477
S	2	8	2	12
Copepoda				
Copepodites				
	16			
N	16	0	0	0
S	1	0	0	0
Acari None observed				

Station TW10i

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Diploscapter coronata</i>	48			
Xyalid sp. 1	48			
<i>Daptonema setosa</i>	3992	7947	749	1650
<i>Sabatieria punctata</i>	144	4279	321	296
<i>Adoncholaimus thalassphygas</i>	96	4891	36	42
<i>Dichromadora geophila</i>	96	28732	71	42
<i>Anoplostoma viviparum</i>	337	14061	267	1988
<i>Microlaimus robustidens</i>	48			
<i>Mononchoides</i> sp.	+	611		
Oncholaimid sp. 2	+			
<i>Daptonema normandica</i>		4891	18	
<i>Adoncholaimus fuscus</i>		+		
<i>Sabatieria breviseta</i>		+	18	
<i>Hypodontolaimus balticus</i>		+		127
Diplogasterid sp.			36	
<i>Hypodontolaimus inaequalis</i>			107	
<i>Sphaerolaimus balticus</i>			18	
<i>Leptolaimus papilliger</i>			36	
<i>Viscosia cobbi</i>			36	
<i>Mononchoides striatus</i>			18	85
<i>Butlerius butleri</i>			18	
<i>Theristus</i> sp. 1			18	
<i>Praeacanthonchus</i> sp.			18	
<i>Ptycholaimellus ponticus</i>			18	
<i>Prochromadorella attenuata</i>			+	
Indet.	0	611	53	42
1B:2A	31.67	0.92	5.33	15.50
N	4809	61743	1856	4272
S	10	10	18	7
Copepoda				
Copepodites	10			
<i>Stenhelia giesbrechti</i>		12		
<i>Paramphiascella</i> sp.			9	
N	10	12	9	0
S	1	1	1	0

Station TW10i continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Acari				
Oribatid sp.	10	44		
<i>Copidognathus rhodostigma</i>	10			
N	20	44	0	0
S	2	1	0	0

Station TW11s

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Calytronema maxweberi</i>	8		376	
<i>Daptonema setosa</i>	40	71	658	707
<i>Sabatieria punctata</i>	32	900	5542	7069
<i>Sphaerolaimus gracilis</i>	8		94	
<i>Prycholaimellus ponticus</i>	8		+	
<i>Microlaimus conothelis</i>		14		
<i>Richtersia inaequalis</i>		14		
<i>Sabatieria breviseta</i>			94	
<i>Daptonema furcata</i>			1784	
<i>Leptolaimus papilliger</i>			188	
<i>Monhystera disjuncta</i>			94	
<i>Monhystera filicaudata</i>			282	
<i>Dichromadora cephalata</i>			94	
Monhysterid sp. 1			188	
<i>Sphaerolaimus balticus</i>			94	+
<i>Antomicron elegans</i>			94	
<i>Daptonema</i> sp. 1			+	
<i>Camacolaimus tardus</i>			+	
<i>Oncholaimus brachycercus</i>			+	
<i>Terchellingia longicaudata</i>				79
Indet.	0	14	188	0
1B:2A	10.00	69.00	88.00	x2A
N	96	1013	9770	7855
S	5	4	17	4
Copepoda				
Copepodites	8			
<i>Paramphiascella</i> sp.			42	
<i>Amphiascus</i> sp. 2				45
<i>Harpacticella</i> sp.				164
<i>Microarthridion</i> sp.				64
<i>Tachidius discipes</i>				409
N	8	0	42	682
S	1	0	1	4

Station TW11s continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Acari				
<i>Copidognathus rhodostigma</i>	8			
Oribatid sp.				9
N	8	0	0	9
S	1	0	0	1

Station TW11i

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Daptonema setosa</i>	1063	1830	11	
<i>Sabatieria punctata</i>	48	330		533
<i>Diplolaimella ocellata</i>	12			
<i>Daptonema</i> sp. 1	12			
<i>Metachromadora suecica</i>	12	30		
<i>Leptolaimus papilliger</i>	12	30		
<i>Dichromadora geophila</i>	12	150		
<i>Anoplostoma viviparum</i>	12	30		
<i>Monhystera filicaudata</i>	12			25
<i>Daptonema normandica</i>	+	300		
<i>Daptonema furcata</i>		30		
Monhysterid sp.		30		
<i>Spilophorella paradoxa</i>		90		8
<i>Antomicron elegans</i>		30		
<i>Hypodontolaimus balticus</i>		30		
<i>Spirinia parasitifera</i>		30		
<i>Calomicrolaimus honestus</i>		30		
<i>Spilophorella candida</i>			22	
<i>Paraoigrolaimellus bernensis</i>				8
<i>Desmolaimus zeelandicus</i>				8
<i>Mononchoides</i> sp.				42
<i>Prycholaimellus ponticus</i>				17
<i>Richtersia inaequalis</i>				8
<i>Viscosia cobbi</i>				8
Indet.	0	0	0	25
1B:2A	47.00	7.73	0.50	9.50
N	1207	3000	33	757
S	10	15	2	10
Copepoda				
<i>Stenhelia palustris</i>	24	11		
<i>Stenhelia giesbrechti</i>		11		
<i>Tachidius discipes</i>				17
N	24	22	0	17
S	1	2	0	1

Station TW11i continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Acari				
<i>Copidognathus rhodostigma</i>	8			
N	8	0	0	0
S	1	0	0	0

Station TW12s

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Theristus acer</i>	12			
<i>Sabatieria punctata</i>	447	38	200	1938
<i>Sphaerolaimus gracilis</i>	162			
<i>Sabatieria breviseta</i>	25			
<i>Oncholaimid</i> sp. 1	25			
<i>Tripyloides</i> sp.	87			
<i>Anticoma acuminata</i>	12			
<i>Leptolaimus papilliger</i>	236			42
<i>Axonolaimus paraspinosus</i>	12			
<i>Dichromadora cephalata</i>	112			
<i>Paracanthonchus heterodontus</i>	50			
<i>Monhystera</i> sp.	12			
<i>Daptonema setosa</i>	25		13	42
<i>Diplolaimella ocellata</i>	12			
<i>Monhystera filicaudata</i>	12		13	
<i>Enoplus brevis</i>	+			
<i>Daptonema</i> sp. 1			100	
<i>Theristus</i> sp. 3			13	
<i>Leptolaimus</i> sp. 2			13	21
<i>Prycholaimellus ponticus</i>				21
<i>Desmolaimus zeelandicus</i>				21
Indet.	37	0	0	0
1B:2A	4.31	x2A	x2A	96.00
N	1278	38	352	2085
S	16	1	6	1
Copepoda				
Copepodites	8			
<i>Amphiascus</i> sp. 2				8
N	8	0	0	8
S	1	0	0	1
Acari None observed				

Station TW12i

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Daptonema tenuispiculum</i>	1120			
<i>Anoplostoma viviparum</i>	124			
<i>Leptolaimus papilliger</i>	560	1475	1165	192
<i>Cyatholaimus</i> sp.	809			
<i>Ptycholaimellus ponticus</i>	1493	983	647	80
<i>Dichromadora geophila</i>	62	1475	194	
<i>Spilophorella paradoxa</i>	871	1311		465
<i>Axonolaimus paraspinosus</i>	622	1147	324	32
<i>Spaerolaimus gracilis</i>	62	164		
<i>Sabatieria punctata</i>	124	6392	1229	337
<i>Desmolaimus zeelandicus</i>	124	492	712	48
<i>Sabatieria celtica</i>	62			
<i>Diplolaimella ocellata</i>	62			
<i>Sphaerolaimus balticus</i>	124		65	
<i>Calyptronema maxweberi</i>	+	+	129	48
<i>Automicron elegans</i>	+		+	
<i>Praeacanthonchus</i> sp.		492	194	
<i>Dichromadora cephalata</i>		328	776	16
<i>Metachromadora suecica</i>		164		
<i>Sabatieria breviseta</i>		328	65	
<i>Daptonema</i> sp. 2		1147		
<i>Molgolaimus cuanensis</i>		328		
<i>Hypodontolaimus balticus</i>		164		
<i>Parasphaerolaimus paradoxa</i>		+	712	
<i>Chromadora macrolaima</i>		+		
<i>Spilophorella candida</i>			65	
<i>Daptonema</i> sp. 1			129	
<i>Halalaimus isaitshikovi</i>			65	16
<i>Daptonema setosa</i>				242
<i>Praeacanthonchus opheliae</i>				32
<i>Metachromadora remanei</i>				16
<i>Neochromadora poecilosoma</i>				96
<i>Monhystera filicaudata</i>				16
<i>Diploscapter coronata</i>				16
<i>Richtersia inaequalis</i>				+

continued ...

Station TW12i continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<i>Adoncholaimus thalassophygas</i>				+
Indet.	435	820	129	48
1B:2A	0.71	2.14	1.14	0.91
N	6654	17210	6600	1700
S	16	18	16	17
Copepoda				
<i>Pseudomesochra latifurea</i>	40			
<i>Stenhelia palustris</i>	64	10		
Copepodites	40			
<i>Bryocamptus</i> sp.		10		
<i>Stenhelia giesbrechti</i>		180		
<i>Amphiascus angustipes</i>		70		
<i>Tachidius discipes</i>			25	
<i>Amphiascus</i> sp. 2				8
N	144	270	25	8
S	3	4	1	1
Acari				
Oribatid sp.		30		
N	0	30	0	0
S	0	1	0	0

Station TW13s

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Sabatieria punctata</i>	799	482	278	25
<i>Daptonema setosa</i>	1598	18		125
<i>Leptolaimus papilliger</i>	148	27	41	8
<i>Halalaimus isaitshikovi</i>	30			8
<i>Oncholaimid</i> sp. 1	30			
<i>Richtersia inaequalis</i>	30			17
<i>Anoplostoma viviparum</i>	30			
<i>Terschellingia longicaudatus</i>	30	18		
<i>Monhystera filicaudata</i>	89			
<i>Sabatieria celtica</i>	30			
<i>Desmolaimus zeelandicus</i>	30		10	
<i>Linhomoeid</i> sp. 1	59		31	
<i>Antomicron elegans</i>	30	9		
<i>Dichromadora cephalata</i>	30	9		
<i>Camacolaimus barbatus</i>	+			8
<i>Sphaerolaimus balticus</i>	+		10	
<i>Adoncholaimus thalassophygas</i>	+			
<i>Calyptronema maxweberi</i>	+	45		
<i>Ptycholaimellus ponticus</i>		18		17
<i>Sphaerolaimus gracilis</i>		36	10	
<i>Leptolaimus</i> sp. 1		9		
<i>Sabatieria breviseta</i>		9		
<i>Paracanthonchus heterodontus</i>		9		
<i>Parasphaerolaimus paradoxa</i>		9		
<i>Spirinia parasitifera</i>			10	
<i>Desmodora communis</i>			10	8
<i>Leptolaimus</i> sp. 2				8
<i>Monhystera disjuncta</i>				8
<i>Metachromadora scotlandica</i>				17
Indet.	178	9	10	33
1B:2A	85.00	15.25	30.00	3.33
N	3141	725	410	282
S	18	14	8	11

continued ...

Station TW13s continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<hr/>				
Copepoda				
<i>Schizopera clandestina</i>	48			
<i>Tachidius discipes</i>			10	25
<i>Amphiascus</i> sp. 2				25
<i>Harpacticella</i> sp.				83
<i>Halectinosoma curticorne</i>				8
<i>Stenhelia palustris</i>				33
N	48	0	10	174
S	1	0	1	5

Acari None observed

Station TW13i

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Daptonema tenuispiculum</i>	1477			
<i>Sabatieria punctata</i>	2769	1271	544	209
<i>Leptolaimus papilliger</i>	369	741	272	63
<i>Desmolaimus zeelandicus</i>	738	635	272	42
<i>Sphaerolaimus gracilis</i>	646	106	136	42
<i>Daptonema setosa</i>	831		188	
<i>Molgolaimus cuanensis</i>	92			
<i>Axonolaimus paraspinosus</i>	462	1800		167
<i>Dichromadora cephalata</i>	369	212	3946	63
<i>Monhystera</i> sp. 1	185			
<i>Calomicrolaimus parhonestus</i>	92			
<i>Oncholaimellus calvadosicus</i>	92			
<i>Linhomoeus</i> sp. 1	185			
<i>Anoplostoma viviparum</i>	92			
<i>Monhystera filicaudata</i>	92			
<i>Tripyloides</i> sp.	92			
<i>Sphaerolaimus balticus</i>	277			
<i>Calyptronema maxweberi</i>	92	212	+	63
<i>Sabatieria breviseta</i>	92			
<i>Prochromadora</i> sp.	92			
<i>Metachromadora suecica</i>	92	+		
<i>Antomicron elegans</i>	+			
<i>Cyatholaimus gracilis</i>	+			
<i>Ascolaimus elongatus</i>	+			
<i>Halalaimus isaitshikovi</i>	+			
<i>Paracanthonchus heterodontus</i>	+			42
<i>Parasphaerolaimus paradoxa</i>	+	424		
<i>Spirinia parasitifera</i>	+			
<i>Richtersia inaequalis</i>	+			+
<i>Dichromadora geophila</i>		529		
<i>Daptonema</i> sp. 1		4024		774
<i>Adoncholaimus thalassophygas</i>		106		
<i>Chromadora macrolaima</i>		212		
<i>Praeacanthonchus</i> sp.		318	1225	
<i>Viscosia elegans</i>		+		
<i>Terschellingia longicaudata</i>		+		
<i>Daptonema</i> sp. 2			5443	

continued ...

Station TW13i continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<i>Linhomoeid</i> sp. 1			680	
<i>Viscosia viscosa</i>			680	
<i>Daptonema furcata</i>			136	
<i>Chromadorita tentabunda</i>			136	
<i>Microlaimus marinus</i>			136	
<i>Prycholaimellus ponticus</i>			136	
<i>Leptolaimus</i> sp. 2			+	
<i>Desmodora communis</i>			+	
<i>Daptonema normandica</i>				109
<i>Diplolaimella ocellata</i>				63
<i>Praeacanthonchus opheliae</i>				126
<i>Viscosia cobbi</i>				21
<i>Metachromadora scotlandica</i>				+
Indet.	554	635	544	126
1B:2A	10.00	8.89	1.78	4.87
N	9782	11225	14286	2199
S	29	16	13	19
Copepoda				
<i>Pseudomesochra latifurea</i>	496			
<i>Schizopera clandestina</i>	328			
<i>Stenhelia palustris</i>	992	18		118
Copepodites	328		56	
<i>Bryocamptus</i> sp.		36		
<i>Tachidius discipes</i>			78	300
<i>Harpacticella</i> sp.				109
<i>Amphiascoides</i> sp.				73
N	2144	54	134	600
S	4	2	2	4
Acari				
<i>Copidognathus rhodostigma</i>	8			
N	8	0	0	0
S	1	0	0	0

Station TW14

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Sphaerolaimus gracilis</i>	107			
<i>Metachromadora suecica</i>	213	+		103
Linhomoeid sp. 1	5016	3460	6636	3608
<i>Sabatieria punctata</i>	4375	8073	3792	5155
<i>Daptonema normandica</i>	427			
<i>Sabatieria celtica</i>	107			
<i>Chromadora macrolaima</i>	107			
Cyatholaimid sp. 1	107			
<i>Desmolaimus zeelandicus</i>	107			
<i>Ptycholaimellus ponticus</i>	107			
<i>Microlaimus conothesis</i>	+			
<i>Richtersia inaequalis</i>	+			103
<i>Monhystera</i> sp.	+			
<i>Ascolaimus elongatus</i>	+			+
<i>Eleutherolaimus</i> sp.	+			
<i>Criconemoides</i> sp.	+			
<i>Molgolaimus cuanensis</i>		897		
<i>Parasphaerolaimus paradoxa</i>		128		
<i>Daptonema setosa</i>		128		515
<i>Leptolaimus</i> sp. 1		128		
<i>Daptonema furcata</i>		+		
<i>Daptonema</i> sp. 2		+		
<i>Paracanthonchus heterodontus</i>		+		+
<i>Viscosia viscosa</i>			105	
<i>Spirinia parasitifera</i>			+	
<i>Sabatieria breviseta</i>			+	
<i>Leptolaimus</i> sp. 3			+	
<i>Daptonema</i> sp. 1			+	103
<i>Viscosia cobbi</i>				515
<i>Sphaerolaimus balticus</i>				103
<i>Terschellingia longicaudatus</i>				103
Indet.	213	256	0	0
1B:2A	19.00	9.29	x2A	58.00
N	10886	13070	10533	10308
S	16	10	71	11

continued ...

Station TW14 continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<hr/>				
Copepoda				
<i>Stenhelia giesbrechti</i>		20		
<i>Schizopera</i> sp.		10		
<i>Stenhelia palustris</i>		60		
<i>Halectinosoma curticorne</i>			17	
Cletodid sp.				18
<i>Tachidius</i> sp.				18
N	0	90	17	36
S	0	3	1	2
Acari				
Oribatid sp.		30	33	
N	0	30	33	0
S	0	1	1	0

Station TW15

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Daptonema setosa</i>	114		25	15
<i>Eleutherolaimus</i> sp.	3430			
Desmodorid sp. 1	1372			59
<i>Richtersia inaequalis</i>	1829		13	117
<i>Microalaimus robustus</i>	343			
Oncholaimid sp. 2	915			
<i>Daptonema furcata</i>	572			
<i>Daptonema normandica</i>	800			396
<i>Sphaerolaimus gracilis</i>	114			
<i>Enoploides brunettii</i>	114			15
<i>Oncholaimellus calvadosicus</i>	572			
<i>Odontophora villoti</i>	114			
<i>Sigmophoranema rufum</i>	114			
<i>Rhabdocoma riemani</i>	114			
<i>Ascolaimus elongatus</i>	343			147
<i>Sabatieria punctata</i>	114	33	239	235
<i>Chromadorita tentabunda</i>	457			15
<i>Metachromadora suecica</i>	+	11		
<i>Monoposthia mirabilis</i>	+		13	44
<i>Camacolaimus barbatus</i>	+	33		
<i>Ptycholaimellus ponticus</i>		11		
<i>Odontophora setosa</i>		22		
<i>Daptonema</i> sp. 1		11	880	
<i>Leptolaimus</i> sp. 1		11		
<i>Monhystera filicaudata</i>		11		
<i>Spirinia parasitifera</i>		11		
<i>Calyptronema maxweberi</i>		22		15
Monhysterid sp. 2		44		15
<i>Adoncholaimus thalassophygas</i>		11		
<i>Microalimus conothesis</i>		11		15
<i>Leptolaimus papilliger</i>		11		15
<i>Onchium conicaudatum</i>		11	13	
<i>Daptonema</i> sp. 2		11		
<i>Viscosia glabra</i>		11		
Linhomoeid sp. 1			13	29
<i>Desmodora communis</i>			38	15
<i>Viscosia viscosa</i>			13	

continued ...

Station TW15 continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<i>Linhomoeus</i> sp. 1			13	
<i>Eleutherolaimus</i> sp.				15
<i>Viscosia cobbi</i>				191
<i>Calaomicrolaimus honestus</i>				29
<i>Halalaimus isaitshikovi</i>				15
<i>Monhystera</i> sp.				15
<i>Setosabatieria hilarula</i>				44
Diplopelid sp. 1				15
<i>Sphaerolaimus balticus</i>				+
Indet.	1143	22	13	73
1B:2A	7.60	0.71	23.50	0.11
N	12574	319	1273	1544
S	20	18	10	23
Copepoda				
<i>Ectinosoma melaniceps</i>	3305			
Copepodites	15			
<i>Halectinosoma curticorne</i>			9	50
<i>Phyllothalestris</i> sp.				25
<i>Pseudobradia brevicornis</i>				33
N	3320	0	9	108
S	2	0	1	3
Acari				
Oribatid sp.		11	9	
<i>Copidognathus dentatus</i>			26	
N	0	11	35	0
S	0	1	2	0

Station TW16

Q U A R T E R

Species	1989/2	1982/3	1989/4	1990/1
Nematoda				
<i>Sabatieria celtica</i>	239	+	+	1645
<i>Leptolaimus papilliger</i>	119		+	411
<i>Daptonema normandica</i>	1910			1234
<i>Sabatieria punctata</i>	6325	24757	12039	9050
<i>Microlaimus robustidens</i>	477			
<i>Tripyloides</i> sp.	239			
<i>Paracanthonchus heterodontus</i>	716	+		
Oncholaimid sp. 2	119			
<i>Daptonema furcata</i>	358		388	3702
<i>Leptolaimus</i> sp. 1	239	538		
<i>Chromadora macrolaima</i>	239	8611		6170
<i>Ptycholaimellus ponticus</i>	358	2691	3107	2880
<i>Richtersia inaequalis</i>	119	538	+	+
<i>Halalaimus isaitshikovi</i>	119			+
<i>Eleutherolaimus</i> sp.	119	1076	+	+
Xyalid sp. 2	119			
<i>Metachromadora suecica</i>	119	+	+	
<i>Terschellingia longicaudatus</i>	477	2691	1165	1645
<i>Eurystomina</i> sp.	239			
<i>Daptonema setosa</i>	119	1615	2330	1645
Linhomoeid sp. 1	119		1165	1234
<i>Spirinia parasitifera</i>	119			411
<i>Macroposthonia</i> sp.	+			
Linhomoeid sp. 2	+			411
<i>Setosabatieria hilarula</i>	+		388	411
<i>Spilophorella paradoxa</i>	+			
<i>Oncholaimus oxyuris</i>	+			
<i>Oncholaimellus calvadosicus</i>	+	+		
<i>Monoposthia costata</i>	+		775	+
<i>Monoposthia mirabilis</i>	+		+	
<i>Theristus</i> sp. 3	+			
<i>Dichromadora cephalata</i>		2691	8932	1234
<i>Dichromadora</i> sp.		1076		
<i>Daptonema</i> sp. 2		2153		
<i>Microlaimus marinus</i>		538	+	
<i>Molgolaimus cuanensis</i>		1615		
<i>Chromadorina viridis</i>		538		

continued ...

Station TW16 continued

Q U A R T E R

Species	1989/2	1982/3	1989/4	1990/1
<i>Sabatieria breviseta</i>		538		
<i>Pareudesmoscolex</i> sp.		1076		411
<i>Paracanthochus</i> sp.		538		
<i>Microloaimus conothelis</i>		538		
<i>Sigmophoranema rufum</i>		+		
<i>Trefusia longicaudata</i>		+		
<i>Metachromadora</i> sp.		+		
<i>Calyptronema maxweberi</i>		+		
<i>Ascolaimus elongatus</i>		+	+	411
<i>Chromadorita</i> sp.		+		
<i>Oxystomina asetosa</i>		+	388	
<i>Sphaerolaimus gracilis</i>		+		
<i>Daptonema</i> sp. 1			3107	2468
Cyatholaimid sp. 1			775	
Aegialoalaimid sp. 1			775	411
<i>Paracanthochus caecus</i>			1942	2057
<i>Tripyloides</i> sp.			775	411
<i>Desmodora communis</i>			388	
<i>Chromadorita tentabunda</i>			388	
<i>Sphaerolaimus balticus</i>			+	
<i>Leptolaimus</i> sp. 2				411
<i>Sabatieria longisetosa</i>				823
<i>Metachromadora remanei</i>				411
<i>Calomicrolaimus honestus</i>				823
<i>Desmolaimus zeelandicus</i>				411
<i>Viscosia viscosa</i>				+
Indet.	477	1615	1165	1645
1B:2A	4.59	1.63	1.12	1.64
N	13483	55433	39992	42776
S	31	30	27	31
Copepoda				
<i>Epactophanes richardi</i>	25			
<i>Pseudomesochra latifurea</i>	8			
<i>Amphiascus angusticeps</i>	292	44	453	
<i>Schizopera clandestina</i>	33			
<i>Ectinosoma melaniceps</i>	8			27

continued ...

Station TW16 continued

Q U A R T E R

Species	1989/2	1982/3	1989/4	1990/1
Copepodites	158			
<i>Cletodes limicola</i>		2687		
<i>Stenhelia palustris</i>			133	9
<i>Cletodes longicaudatus</i>			53	
<i>Longipedia coronata</i>			120	
<i>Stenhelia</i> sp.			67	
<i>Paramphiascella</i> sp.			40	
<i>Canuella perplexa</i>				27
<i>Pseudobradia brevicornis</i>				18
<i>Amphiascus varians</i>				18
<i>Microarthridion</i> sp.				18
<i>Enhydrosoma propinquum</i>				167
<i>Stenhelia aemula</i>				27
<i>Tachidius discipes</i>				289
<i>Amphiascus</i> sp. 2				773
<i>Amphiascus</i> sp. 3				55
N	524	2731	866	1388
S	6	2	6	11
Acari				
Oribatid sp.	8	22		
<i>Copidognathus rhodostigma</i>	8			
<i>Copidognathus dentatus</i>		22		
N	16	44	0	0
S	2	2	0	0

Station TW17

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Dichromadora cephalata</i>	5313		596	
<i>Oncholaimellus calvadosicus</i>	483	1074		
<i>Daptonema tenuispiculum</i>	1932	537		525
<i>Metachromadora scotlandica</i>	18838	9133	11615	4725
<i>Monoposthia costata</i>	966			
<i>Ptycholaimellus ponticus</i>	6762	10207	7743	19425
<i>Richtersia inaequalis</i>	183			525
<i>Daptonema furcata</i>	2415	5372	2383	4200
<i>Ascolaimus elongatus</i>	1449			
<i>Chromadorita tentabunda</i>	1449	537		+
<i>Sabatieria celtica</i>	966	537		
<i>Antomicron elegans</i>	483	537		
<i>Axonolaimus paraspinosus</i>	483	1074	1191	525
<i>Sabatieria punctata</i>	3864	8595	893	1575
<i>Odontophora setosa</i>	966	3223	298	525
<i>Paracanthonchus heterodontus</i>	483			52
<i>Camacolaimus tardus</i>	483	537		
<i>Cyatholaimus gracilis</i>	483			
<i>Desmolaimus zeelandicus</i>	483			
<i>Linhomoeus</i> sp. 1	483		298	
<i>Linhomoeus</i> sp. 2	+			
<i>Quadricoma</i> sp. +				
<i>Hypodontolaimus balticus</i>	+			1050
<i>Rhabdocoma riemani</i>	+	+		
<i>Eleutherlaimus</i> sp.	+	+		
<i>Neochromadora poecilosoma</i>	+			+
<i>Oncholaimus brachycercus</i>	+	+		
<i>Calyptronema maxweberi</i>	+	537	+	
<i>Anoplostoma viviparum</i>	+	+	+	
<i>Sphaerolaimus balticus</i>	+		+	+
<i>Tripyloides marinus</i>	+		+	
<i>Oxystomina asetosa</i>	+			
<i>Viscosia glabra</i>	+			
<i>Halalaimus isaitshikovi</i>	+	537	596	525
<i>Odontophora villoti</i>	+	+	+	
<i>Spirinia parasitifera</i>	+			

continued ...

Station TW17 continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<i>Linhomoeid</i> sp. 1		537		
<i>Sigmophoranema rufum</i>		4835		6300
<i>Chromadora macrolaima</i>		1612		5775
<i>Pareudesmoscolex</i> sp.		537		
<i>Sphaerolaimus gracilis</i>		537	596	
<i>Metachromadora suecica</i>		537		
<i>Daptonema setosa</i>		1074	298	4200
<i>Parasphaerolaimus paradoxa</i>		537		
<i>Microlaimus conothesis</i>		537		
<i>Monhystera disjuncta</i>		537	298	
<i>Adoncholaimus thalassophygas</i>		+		
<i>Terschellingia longicaudata</i>		+	+	
<i>Stephanolaimus elegans</i>		+		
<i>Praeacanthonchus</i> sp.		+	298	+
<i>Spilophorella candida</i>			2383	
<i>Desmodora communis</i>			298	
<i>Viscosia viscosa</i>			+	1050
<i>Leptolaimus</i> sp. 3			+	
<i>Viscosia elegans</i>			+	
<i>Linhomoeid</i> sp. 2			+	
Aegialoalaimid sp. 1				525
Cyatholaimid sp. 1				525
<i>Belbolla teisseiri</i>				+
<i>Enoploides brunettii</i>				+
Indet.	483	4835	0	1050
1B:2A	0.34	0.63	0.26	0.30
N	49750	58552	29782	53550
S	36	33	26	23
Copepoda				
<i>Canuella perplexa</i>	24	322		
<i>Amphiascus angusticeps</i>	120	11	108	
<i>Schizopera clandestina</i>	408			
<i>Ectinosoma</i> sp.	24			
Copepodites	120			
<i>Stenhelium giesbrechti</i>		522		
<i>Tisbe</i> sp.		100		
<i>Stenhelium palustris</i>			64	

continued ...

Station TW17 continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<i>Stenhelia</i> sp.			36	25
<i>Halophytophilus</i> sp.				83
<i>Amphiascella</i> sp.				700
<i>Ectinosoma melaniceps</i>				33
<i>Enhydrosoma propinquum</i>				50
N	696	960	708	891
S	5	4	3	5
Acari				
<i>Copidognathus rhodostigma</i>	40			
N	40	0	0	0
S	1	0	0	0

Station TW18

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Theristus</i> sp. 3	146			
<i>Microloaimus marinus</i>	878	138	67	
Xyalid sp. 3	2488			
<i>Leptolaimus papilliger</i>	146	104	270	
<i>Sabatieria punctata</i>	2635	450	911	157
<i>Monhystera filicaudata</i>	293			
Aegialoalaimid sp. 1	732			78
<i>Setosabatieria hilarula</i>	439		101	78
<i>Spirinia parasitifera</i>	146	1245	34	314
<i>Daptonema</i> sp. 1	585			
Linhomoeid sp. 2	293			
<i>Monoposthia costata</i>	293			+
<i>Sabatieria longiseta</i>	2781		67	
<i>Linhomoeus</i> sp. 1	293	35	101	78
Leptolaimid sp. 1	146	69		
<i>Leptolaimus</i> sp. 2	439			157
Oncholaimid sp. 2	1025			
<i>Terschellingia longicaudata</i>	146		34	
<i>Spilophorella paradoxa</i>	146			+
<i>Oxystomina asetosa</i>	146			
<i>Halalaimus isaitshikovi</i>	293			+
<i>Daptonema normandica</i>	439		67	4076
<i>Sabatieria celtica</i>	146		101	157
<i>Anticomma acuminata</i>	146			
Cyatholaimid sp. 2	146			
<i>Campylaimus inaequalis</i>	293			
Linhomoeid sp. 2	146	173	135	
<i>Odontophora setosa</i>	+	35	34	157
<i>Paralongicyatholaimus</i> sp.	+			
<i>Quadricoma</i> sp.	+			
<i>Metachromadora remanei</i>	+		34	314
<i>Sphaerolaimus gracilis</i>	+		135	
<i>Thalassoalaimus tardus</i>	+		+	+
<i>Viscosia elegans</i>	+		101	
<i>Bathylaimus capocosus</i>	+			
<i>Calaomicrolaimus honestus</i>		311		
<i>Stephanolaimus elegans</i>		69		

continued ...

Station TW18 continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<i>Camacolaimus barbatus</i>		69		627
<i>Monhystera disjuncta</i>		104		
<i>Cricolaimus</i> sp.		415		
<i>Viscosia cobbi</i>		69		78
<i>Sabatieria breviseta</i>		35		78
<i>Sabatieria longiseta</i>		35		
<i>Molgolaimus cuanensis</i>		35		
<i>Desmolaimus zeelandicus</i>		35		78
<i>Ascolaimus elongatus</i>		+	67	235
<i>Linhomoeid</i> sp. 3		202		
<i>Eleutherolaimus</i> sp.			34	+
<i>Leptolaimus</i> sp. 3			202	
<i>Trefusia longicaudata</i>			135	157
<i>Richterisa inaequalis</i>			34	314
<i>Xyalid</i> sp. 1			169	+
<i>Linhomoeus</i> sp. 2			169	
<i>Camacolaimus tardus</i>			34	
<i>Chromadorita</i> sp.			34	
<i>Oncholaimus oxyuris</i>			34	
<i>Monhystera disjuncta</i>			34	
<i>Theristus</i> sp. 2			34	
<i>Theristus acer</i>			34	
<i>Daptonema setosa</i>				78
<i>Calomicrolaimus honestus</i>				235
<i>Chromadora macrolaima</i>				78
<i>Bathylaimus capacosus</i>				78
<i>Daptonema furcata</i>				78
<i>Oncholaimus</i> sp.				78
<i>Axonolaimus paraspinosus</i>				78
<i>Oxystomina asetosa</i>				+
<i>Halanonchus</i> sp.				+
<i>Oncholaimus campylocercoides</i>				+
<i>Monoposthia mirabilis</i>				+
<i>Sphaerolaimus balticus</i>				+
<i>Pseudocella coecum</i>				+
<i>Prochromadorella ditlevseni</i>				+
<i>Linhystera</i> sp.				+
<i>Calyptronema maxweberi</i>				+

continued ...

Station TW18 continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<i>Mescacanthion diplochma</i>				+
<i>Anoplostoma viviparum</i>				+
Indet.	1171	243	641	392
1B:2A	6.18	1.47	11.67	3.82
N	16976	3669	4083	8228
S	35	20	31	42
Copepoda				
<i>Tachidius</i> sp.	183			
<i>Schizopera clandestina</i>	117			
<i>Laophonte</i> sp.	17			
<i>Schizopera</i> sp.		8		
<i>Amphiascus angusticeps</i>			42	
<i>Stenhelia palustris</i>				80
Copepodites				63
<i>Stenhelia aemula</i>				3
<i>Bulbamphiascus</i> sp.				3
<i>Halectinosoma curticorne</i>				53
<i>Ectinosoma melaniceps</i>				5
<i>Enhydrosoma propinquum</i>				3
<i>Pseudobradyia brevicornis</i>				5
<i>Paramphiascella</i> sp.				3
<i>Amphiascus</i> sp. 1				33
<i>Canuella perplexa</i>				15
<i>Amphiascus</i> sp. 2				18
<i>Amphiascus</i> sp. 3				20
N	317	8	42	304
S	3	1	1	13
Acari				
Oribatid sp.	8			3
<i>Copidognathus dentatus</i>	17	17		5
<i>Copidognathus rhodostigma</i>		8		
N	25	25	0	8
S	2	2	0	2

Station TW19s

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Metachromadora suecica</i>	81		81	+
<i>Metachromadora</i> sp.	81			
<i>Chaetonema riemanni</i>	27			
<i>Richtersia inaequalis</i>	27		27	199
<i>Halalaimus longicaudatus</i>	27			
<i>Leptolaimus</i> sp. 2	54			
<i>Sabatieria punctata</i>	27	49	108	
<i>Dichromadora</i> sp.	163			
Desmodorid sp. 1	163			
Aegialolaimid sp. 1	54	343		199
<i>Linhomoeus</i> sp. 2	27			
<i>Theristus</i> sp. 3	54			
<i>Desmodora</i> sp. 1	190			
<i>Calamicrolaimus honestus</i>	27	98	54	
<i>Prochromadorella ditlevseni</i>	27	49		
Diplopeltid sp. 1	27	49		
<i>Prycholaimellus ponticus</i>	27			
<i>Halalaimus isaitshikovi</i>	27	49	27	299
<i>Cyatholaimus gracilis</i>	27			
<i>Spilophorella candida</i>	27			299
<i>Daptonema normandica</i>	27			
<i>Onyx perfectus</i>	54	147	135	199
<i>Hypodontolaimus balticus</i>	27			
<i>Paracanthonchus</i> sp.	27			
<i>Camacolaimus tardus</i>	109			
<i>Leptolaimus papilliger</i>	27	49		
<i>Spirinia parasitifera</i>	27		27	+
<i>Pseudonchus</i> sp.	27			
<i>Oncholaimus brachycercus</i>	27			
<i>Ascolaimus elongatus</i>	+	49		
<i>Mesacanthion diplochma</i>	+	98		199
<i>Spilophorella paradoxa</i>		294	27	100
Cyatholaimid sp. 1		736		
<i>Odontophora setosa</i>		343	27	
<i>Oncholaimellus calvadosus</i>		49	+	
Xyalid sp. 2		98		
<i>Trefusia longicaudata</i>		49		
Ceramonematid sp. 1		49		

continued ...

Station TW19s continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<i>Microlaimus marinus</i>		49	27	
<i>Cricolaimus</i> sp.		+		
<i>Daptonema setosa</i>		+		
<i>Dichromadora cephalata</i>			242	1396
<i>Sigmophoranema rufum</i>			81	
Xyalid sp. 1			323	
<i>Viscosia viscosa</i>			108	
<i>Daptonema</i> sp. 2			27	
Rhabditid sp. 2			27	
<i>Paralongicyatholaimus</i> sp.			189	
Cyatholaimid sp. 2			108	1995
<i>Microlaimus conothesis</i>			27	
<i>Desmoscolex falcatus</i>			27	+
<i>Theristus</i> sp. 3			27	
<i>Enoplolaimus vulgaris</i>			54	
<i>Chromadorita tentabunda</i>			27	
<i>Odontophora villoti</i>			27	299
<i>Sabatieria longiseta</i>			27	
<i>Viscosia glabra</i>				299
<i>Oncholaimus skawensis</i>				100
<i>Paracanthoichus caecus</i>				199
<i>Eleutherolaimus</i> sp.				100
<i>Neochromadora trichophora</i>				100
<i>Leptolaimus</i> sp. 3				100
<i>Halichoanolaimus robustus</i>				+
Indet.	326	343	215	499
1B:2A	1.49	1.36	1.44	0.69
N	3063	5247	2911	10471
S	32	22	29	22
Copepoda				
<i>Ectinosoma melaniceps</i>	80			
<i>Halecrinosoma</i> sp.	16		245	
Copepodites	16		9	
<i>Stenhelia giesbrechti</i>		8		
<i>Canuella perplexa</i>		25	9	18
<i>Cletodes longicaudatus</i>			18	
N	112	33	281	18
S	3	2	3	1

continued ...

Station TW19s

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Acari				
<i>Copidognathus dentatus</i>				9
N	0	0	0	9
S	0	0	0	1

Station TW19i

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Odontophora villosi</i>	1587			
<i>Chromadora macrolaima</i>	5555			15039
<i>Prycholaimellus ponticus</i>	15078	10468	7821	7520
<i>Tripylloides</i> sp.	1587	13689	869	1504
<i>Metachromadora riemani</i>	794		1738	48125
<i>Ascolaimus elongatus</i>	2381	805		7520
<i>Desmolaimus zeelandicus</i>	6349			
<i>Theristus</i> sp. 3	794		869	
<i>Microlaimus marinus</i>	3968	24962		
<i>Metachromadora</i> sp.	14284	1610		
<i>Monoposthia costata</i>	2381	2416	2607	1504
Linhomoeid sp. 1	2381		2607	
<i>Anoplostoma viviparum</i>	1587	1610	7821	7520
<i>Paracanthonchus heterodontus</i>	2381	805		
<i>Terschellingia longicaudatus</i>	1587	805	869	1504
<i>Calomicrolaimus honestus</i>	794	805	4345	
<i>Richtersia inaequalis</i>	794	+	+	+
<i>Axonolaimus paraspinosus</i>	794	805		
<i>Praeacanthonchus</i> sp.	1587	1610	869	
<i>Calyptronema maxweberi</i>	794			+
Linhomoeus sp. 2	794			1504
<i>Eleutherolaimus</i> sp.	794	+	869	
<i>Daptonema normandica</i>	1587	6442	7821	
<i>Daptonema tenuispiculum</i>	1587	805	1738	
<i>Sabatieria punctata</i>	1587	8858	4345	4512
<i>Oncholaimellus calvadosicus</i>	1587			
<i>Viscosia</i> sp. 2	2381			
<i>Spirinia parasitifera</i>	794	1610	1738	6016
<i>Daptonema</i> sp. 2	794			+
<i>Hypodontolaimus balticus</i>	+		+	3008
<i>Sphaerolaimus balticus</i>	+		+	1504
<i>Sabatieria longicaudatus</i>	+			
<i>Nemanema cylindricaudatum</i>	+		+	
<i>Quadricoma</i> sp.	+			
<i>Desmoscolex falcatus</i>	+			
<i>Chromadorita tentabunda</i>		2416	1738	1504
<i>Viscosia glabra</i>		805		
Diplopeltid sp. 1		805		

continued ...

Station TW19i continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<i>Oncholaimus skawensis</i>		805	1738	1504
<i>Daptonema setosa</i>		805	1738	
Comesomatid sp. 1		+		
<i>Euchromadora vulgaris</i>		+		
<i>Parasphaerolaimus paradoxa</i>		+		
<i>Dichromadora cephalata</i>			11296	
<i>Leptolaimus</i> sp. 3			2607	
Aegialoalaimid sp. 1			3476	1504
<i>Theristus acer</i>			869	
Xyalid sp. 1			1738	
<i>Microlaimus conothesis</i>			18247	
<i>Monoposthia mirabilis</i>			869	
<i>Anticoma acuminata</i>			+	
<i>Mesacanthion diplochma</i>			+	
<i>Halalaimus isaitshikovi</i>			+	
<i>Viscosia elegans</i>			+	
<i>Setosabatieria hilarula</i>			+	
<i>Sigmphoranema rufum</i>				6016
<i>Paracanthonchus caecus</i>				7520
<i>Odontophora setosa</i>				4512
<i>Microlaimus robustidens</i>				12031
<i>Oxystomina asetosa</i>				1504
<i>Viscosia cobbi</i>				1504
<i>Tripyloides marinus</i>				4512
<i>Praeacanthonchus opheliae</i>				1502
<i>Oxystomina elongata</i>				+
<i>Thalassoalaimus tardus</i>				+
Indet.	6349	4026	4345	3008
1B:2A	0.40	0.34	0.54	0.26
N	85711	87767	101670	153403
S	35	27	36	30
Copepoda				
<i>Bryocamptus</i> sp.	709			
<i>Pseudomesochra latifurea</i>	205			
<i>Schizopera clandestina</i>	103			
<i>Laophonte</i> sp.	1829			
Copepodites	308	10	53	
<i>Ectinosoma</i> sp.		80		

continued ...

Station TW19i continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<i>Stenhelia geisbrechti</i>		760		
<i>Paramphiascella</i> sp.		10		
<i>Longipedia</i> sp.		70		
<i>Longipedia coronata</i>			441	
Cletodid sp.				83
<i>Stenhelia palustris</i>				100
<i>Amphiascus</i> sp. 1				75
N	3154	930	494	258
S	5	5	2	3
Acari				
Oribatid sp.	17		18	8
N	17	0	18	8
S	1	0	1	1

Station TW20

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Desmodora communis</i>	288			
<i>Daptonema tenuispiculum</i>	8209	25		+
<i>Oncholaimellus calvadosicus</i>	1584			394
<i>Desmolaimus zeelandicus</i>	288			
<i>Richtersia inaequalis</i>	1152		353	1444
<i>Metachromadora</i> sp.	144			
<i>Linhomoeus</i> sp. 2	144			
<i>Paracanthonchus</i> sp.	144			
<i>Metachromadora scotlandica</i>	864			
<i>Monoposthia mirabilis</i>	720		50	
<i>Ascolaimus elongatus</i>	144	62		1313
<i>Onyx perfectus</i>	144			
<i>Sabatieria punctata</i>	144	895	3584	3545
<i>Mesacanthion diplochma</i>	144			263
<i>Microlaimus robustidens</i>	144			131
<i>Chaetonema riemani</i>	144			+
<i>Eleutherolaimus</i> sp.	+			131
<i>Mononchus aquaticus</i>	+			
<i>Leptolaimus</i> sp. 2	+			
<i>Oncholaimus skawensis</i>	+			
<i>Xyalid</i> sp. 3	+			263
<i>Sphaerolaimus gracilis</i>	+			
<i>Metachromadora suecica</i>		25	101	263
<i>Cricolaimus</i> sp.		12		
<i>Microlaimus marinus</i>		25		
<i>Terschellingia longicaudatus</i>		37	505	789
<i>Chromadora macrolaima</i>		75		131
<i>Leptolaimus papilliger</i>		12	+	
<i>Theristus acer</i>		12		
<i>Daptonema normandica</i>		12	50	1838
<i>Calomicrolaimus honestus</i>		37	50	
<i>Paramonhystera</i> sp.		12		
<i>Sabatieria celtica</i>			50	
<i>Spirinia parasitifera</i>			50	
<i>Molgolaimus cuanensis</i>			101	131
<i>Trefusia longicaudatus</i>			50	131
<i>Dichromadora cephalata</i>			50	
<i>Setosabatieria hilarula</i>			50	131

continued ...

Station TW20 continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<i>Viscosia elegans</i>			+	
Xyalid sp. 1			+	
<i>Leptolaimus</i> sp. 3			+	
<i>Viscosia viscosia</i>				263
Desmodorid sp. 1				394
<i>Sabatieria breviseta</i>				263
Aegialoalaimid sp. 1				525
<i>Microlaimus conothesis</i>				131
<i>Chromadorita tentabunda</i>				131
<i>Marylynnia</i> sp.				131
<i>Ptycholaimellus ponticus</i>				131
<i>Leptolaimus papilliger</i>				131
<i>Trefusia zostericola</i>				131
<i>Sphaerolaimus balticus</i>				+
<i>Odontophora setosa</i>				+
Indet.	1008	37	202	1050
1B:2A	4.18	5.92	11.57	6.00
N	15409	1278	5246	14179
S	22	13	17	29
Copeopda				
<i>Ectinosoma melaniceps</i>	1905			
Copepodites	38	10		
<i>Halectinosoma herdmani</i>				73
Cletodid sp.				18
N	1943	10	0	73
S	2	1	0	2
Acari None observed				

Station TW21

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Daptonema normandica</i>	1523	2959		+
<i>Microlaimus robustidens</i>	20556			
<i>Desmodora communis</i>	8374		360	
Aegialoalaimid sp. 1	761	592	+	2404
<i>Ascolaimus elongatus</i>	9874	592	+	2404
<i>Viscosia cobbi</i>	4568			
<i>Dichromadora cephalata</i>	761		360	
<i>Richtersia inaequalis</i>	2284	+	+	4808
<i>Theristus</i> sp. 3	1523		2882	
<i>Metachromadora</i> sp.	1523			
Xyalid sp. 3	1294			
Cyatholaimid sp. 2	761			2404
<i>Microlaimus conothesis</i>	761	2959	2162	3606
<i>Odontophora villoti</i>	3045			4808
<i>Prochromadorella ditlevseni</i>	761	592		
<i>Calomicrolaimus honestus</i>	1523	18346	3243	50488
<i>Viscosia</i> sp.	761			
<i>Daptonema setosa</i>	2284		+	+
<i>Camacolaimus tardus</i>	761			
<i>Eleutherolaimus</i> sp.	761			+
<i>Sabatieria celtica</i>	+			
<i>Metachromadora scotlandica</i>	+			
<i>Stephanolaimus spartinae</i>	+			
<i>Nemanema cylindratucaudatum</i>	+			
<i>Oncholaimellus clavadosicus</i>	+	592	3963	4808
<i>Calyptronema maxweberi</i>	+	+		
<i>Oxystomina asetsoa</i>	+			+
<i>Enoploides brunettii</i>	+	+	+	1202
<i>Daptonema furcata</i>	+	+		4808
<i>Bathylaimus capacosus</i>	+	1184	360	2404
<i>Sphaerolaimus balticus</i>	+		+	1202
<i>Oncholaimus brachycercus</i>	+			
<i>Daptonema tenuispiculum</i>	+			
<i>Sphaerolaimus gracilis</i>	+			
<i>Coninckia</i> sp.	+			
<i>Axonolaimus paraspinosus</i>	+	592		1202
<i>Prycholaimellus ponticus</i>	+			1202

continued ...

Station TW21 continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<i>Paramonhystera</i> sp.	+	15979		4808
<i>Quadricoma</i> sp.	+			
<i>Thalassoalaimus tardus</i>	+			
<i>Microlaimus marinus</i>		3551	10808	
<i>Theristus acer</i>		1184		
<i>Trefusia longicaudata</i>		592	360	1202
Xyalid sp. 2		592		
<i>Chromadora macrolaima</i>		1775		1202
<i>Sabatieria punctata</i>		592	360	+
<i>Spilophorella paradoxa</i>		592		
Desmodorid sp. 2		1175		1202
<i>Metachromadora</i> sp.		592		
<i>Stephanolaimus jayassreei</i>		592		1202
Cyatholaimid sp. 1		592		
<i>Sigmophoranema rufum</i>		592		1202
<i>Odontophora setosa</i>		592	721	
<i>Viscosia glabra</i>		1184		
<i>Parasphaerolaimus paradoxa</i>		+		
<i>Adoncholaimus fuscus</i>		+		+
<i>Monoposthia costata</i>		+		1202
<i>Monoposthia mirabilis</i>		+	360	
<i>Chromadorita tentabunda</i>			1441	2404
<i>Metachromadora suecica</i>			1801	
<i>Chromadorita</i> sp.			360	
<i>Oncholaimus skawensis</i>			3243	2404
Cermonematid sp. 1			721	
Trefusiid sp. 1			1081	
Xyalid sp. 1			1441	3606
<i>Viscosia viscosa</i>			360	1202
<i>Spirinia parasitifera</i>			+	
<i>Pomponema</i> sp.				1202
<i>Metachromadora remanei</i>				7212
<i>Trefusia zostericola</i>				4808
<i>Mesacanthion diplochma</i>				+
<i>Terschellingia longicaudata</i>				+
Indet.	2284	7694	721	2404
1B:2A	0.72	0.74	0.23	0.32
N	78391	66875	37108	125012
S	40	32	27	36

continued ...

Station TW21 continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Copepoda				
<i>Leptomesochra macintoshi</i>	127			
<i>Bryocamptus</i> sp.	45			
<i>Amphiascus angusticeps</i>	18	8		
<i>Pseudomesochra latifurea</i>	55			
<i>Schizopera clandestina</i>	64			
<i>Ectinosoma</i> sp.	27			
<i>Ectinosoma melaniceps</i>	264			55
<i>Laophonte</i> sp.	482	25		
<i>Stenhelia giesbrechti</i>		167		
<i>Cletodes limicola</i>		342		
<i>Longipedia</i> sp.		50		
<i>Canuella perplexa</i>		8		27
<i>Cletodes longicaudatus</i>			345	
<i>Pseudobradyia brevicornis</i>			9	
<i>Arenostella</i> sp.			200	
<i>Halectinosoma curticorne</i>			18	
<i>Halectinosoma herdmani</i>				182
Cletodid sp.				327
N	1082	600	572	591
S	8	6	4	4
Acari				
Halacarid sp. indet.		8		
N	0	8	0	0
S	0	1	0	0

Station TW22

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Nematoda				
<i>Richtersia inaequalis</i>	2214	5811	3773	3738
<i>Metachromadora scotlandica</i>	939			
<i>Onyx perfectus</i>	134			
<i>Neochromadora trichophora</i>	201			
<i>Paracanthonchus longus</i>	67			
<i>Molgolaimus cuanensis</i>	67			
<i>Calomicrolaimus honestus</i>	67		290	
Desmodorid sp. 1	134	598	290	175
<i>Theristus</i> sp. 3	67			
<i>Spilophorella paradoxa</i>	201	256	73	643
<i>Cyatholaimus gracilis</i>	67			
<i>Daptonema normandica</i>	67			
<i>Desmodora communis</i>	335			
<i>Leptolaimus papilliger</i>	67	171		
<i>Chromadorita tentabunda</i>	67	171		
<i>Microlaimus conothesis</i>	67			
<i>Ascolaimus elongatus</i>	268			175
<i>Prochromadorella ditlevseni</i>	201			
Aegialoalaimid sp. 1	134	171	145	
<i>Halalaimus isaitshikovi</i>	201	513	+	234
<i>Camacolaimus tardus</i>	268			
<i>Sabatieria breviseta</i>	67			
<i>Leptolaimus</i> sp. 3	201			
<i>Rhabdocoma riemani</i>	67	+		
<i>Gammanema rapax</i>	67			
<i>Dichromador cephalata</i>	335		1596	
<i>Viscosia elegans</i>	67	+		
<i>Microlaimus robustidens</i>	67			58
<i>Thalassoalaimus tardus</i>	+			
<i>Odontophora setosa</i>	+	256	798	
Xyalid sp. 3	+			
<i>Pomponema</i> sp.	+			58
Comesomatid sp.	+			
<i>Oncholaimellus calvadosicus</i>	+			
<i>Enoplolaimus vulgaris</i>	+			
<i>Monoposthia mirabilis</i>	+			
Cyatholaimid sp. 1		427		

continued ...

Station TW22 continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
<i>Dichromadora</i> sp.		171		
<i>Monhystera disjuncta</i>		85	73	
<i>Sabatieria punctata</i>		85		
<i>Monoposthia costata</i>		+		58
Cermonematid sp. 1		+		
<i>Microlaimus marinus</i>			73	
Linhomoeid sp. 2			73	
<i>Paracyatholaimus</i> sp. 1			73	
<i>Metachromadora suecica</i>			+	
<i>Halalaimus capitulatus</i>			+	
Leptolaimid sp. 1			+	
<i>Eleutherolaimus</i> sp.				58
<i>Spilophorella candida</i>				117
<i>Mesacanthion diplochma</i>				58
<i>Odontophora villoti</i>				58
Cyatholaimid sp. 2				117
<i>Daptonema furcata</i>				117
<i>Dichromadora cucullata</i>				117
<i>Oncholaimus</i> sp. 1				58
<i>Viscosia glabra</i>				+
Indet.	201	1025	363	292
1B:2A	0.93	3.14	1.20	3.09
N	6905	9740	7620	6131
S	36	16	15	17
Copepoda				
<i>Typhlamphiascus</i> sp.	33			
<i>Ectinosoma melaniceps</i>	89			17
<i>Halectinosoma curticorne</i>	22	8		58
<i>Laophonte</i> sp.	156			
Copepodites			19	
<i>Canuella perplexa</i>				8
<i>Pseudobradia brevicornis</i>				83
<i>Halectinosoma herdmani</i>				8
N	300	8	19	174
S	4	1	1	5

continued ...

Station TW22 continued

Q U A R T E R

Species	1989/2	1989/3	1989/4	1990/1
Acari				
<i>Copidognathus rhodostigma</i>	33		19	8
<i>Copidognathus dentatus</i>			9	
Halacarid sp. indet.			9	
N	33	0	37	8
S	1	0	3	1



Section 3

**OTHER MEIOFAUNAL GROUPS
PRESENT IN THAMES SEDIMENT SAMPLES**

APRIL 1989 - MARCH 1990

Section 3: Table 1. Key to Table of Other Meiofaunal Groups observed in Thames Sediment Samples, April - June 1989

- | | |
|---|--|
| 1 | Rhizopod Amoebae, tests of <i>Centropyxis</i> species |
| 2 | Foraminifera, planispiral and multilocular species (e.g. <i>Elphidium</i> and <i>Braziliana</i> species) |
| 3 | Turbellaria |
| 4 | Gnathostomulida |
| 5 | Gastrotricha, ? <i>Chaetonotus</i> species |
| 6 | Kinorhyncha, <i>Pycnophyes</i> species |
| 7 | Tardigrada, <i>Macrobiotus dispar</i> and <i>Batillipes mirus</i> |
| 8 | Ostracoda, juveniles of several species indet. |
-

Section 3: Table 2. Other Meiofaunal Groups Observed in Thames Sediment Samples, April - June 1989

Stn.	G R O U P							
	1	2	3	4	5	6	7	8
1	+		+				+	+
2	+		+				+	+
3	+						+	
4	+		+				+	
5		+	+				+	
6								
7	+	+					+	+
8s		+						
8i			+					
9		+						
10s								
10i			+					+
11s	+							
11i	+							
12s			+					
12i			+					
13s								+
13i		+	+					+
14	+	+						
15	+							+
16		+	+				+	+
17		+				+		+
18		+	+					+
19s	+	+				+		
19i	+	+					+	
20			+			+	+	+
21		+	+	+	+		+	
22		+	+		+		+	+

Section 3: Table 3. Key to Table of Other Meiofaunal Groups observed in Thames Sediment Samples, July - September 1989

- | | |
|---|--|
| 1 | Ciliate Protozoa |
| 2 | Foraminifera, planispiral, uni- and multilocular species (e.g. <i>Elphidium</i> , <i>Braziliana</i> and <i>Lagena</i> species) |
| 3 | Turbellaria (several species) |
| 4 | Tardigrada, <i>Echiniscus</i> species and <i>Baillipes mirus</i> |
| 5 | Gastrotricha, <i>Chaetonotus</i> species |
| 6 | Ostracoda, juveniles of several species |
-

Section 3: Table 4. Other Meiofaunal Groups Observed in Thames Sediment Samples, July - September 1989

Stn.	G R O U P					
	1	2	3	4	5	6
1		+	+			+
2						+
3				+		
4			+			
5			+	+		
6			+			+
7s						
7i				+		
8s			+			
8i						
9		+				
10s						
10i			+			
11s			+			
11i		+				
12s		+				
12i			+			
13s		+				+
13i		+	+			
14		+				
15		+				+
16	+					
17	+	+				+
18			+			
19s		+	+			
19i	+	+	+			+
20		+			+	
21	+	+	+			+
22		+		+	+	
Stn.	1	2	3	4	5	6

Section 3: Table 5. Key to Table of Other Meiofaunal Groups observed in Thames Sediment Samples, October - December 1989

- | | |
|---|--|
| 1 | Ciliate Protozoa |
| 2 | Foraminifera, planispiral, uni- and multilocular species (e.g. <i>Elphidium</i> , <i>Braziliana</i> and <i>Lagena</i> species) |
| 3 | Turbellaria (several species) |
| 4 | Nemertea, juveniles indet. |
| 5 | Tardigrada, <i>Echiniscus</i> species and <i>Batillipes mirus</i> |
| 6 | Gastrotricha, <i>Chaetonotus</i> species |
| 7 | Kinorhyncha, <i>Pycnophyes</i> and <i>Echinoderes</i> species |
| 8 | Ostracoda, juveniles of several species |
-

Section 3: Table 6. Other Meiofaunal Groups Observed in Thames Sediment Samples, October - December 1989

Stn.	G R O U P							
	1	2	3	4	5	6	7	8
1			+		+	+		+
2								
3			+		+			+
4								
5			+					
6		+	+					
7s		+	+					+
7i								+
8s		+						
8i			+					
9		+						+
10s								
10i								
11s		+						+
11i		+						
12s		+			+			
12i		+	+					
13s								
13i		+						
14		+	+	+				+
15								+
16		+	+					+
17		+					+	
18			+					
19s		+	+		+			
19i		+	+					+
20		+						
21	+	+						
22	+	+						+

Section 3: Table 7. Key to Table of Other Meiofaunal Groups observed in Thames Sediment Samples, January - March 1989

- | | |
|---|---|
| 1 | Ciliate Protozoa |
| 2 | Foraminifera, planispiral, uni- and multilocular species
(e.g. <i>Elphidium</i> , <i>Braziliana</i> and <i>Lagena</i> species) |
| 3 | Turbellaria (several species) |
| 4 | Gnathostomulida |
| 5 | Gastrotricha, <i>Chaetonotus</i> species |
| 6 | Kinorhyncha, <i>Pycnophyes</i> and <i>Echinoderes</i> species |
| 7 | Tardigrada, species of <i>Echiniscus</i> and <i>Macrobiotus</i> and
<i>Batillipes mirus</i> |
| 8 | Ostracoda, juveniles of several species |
-

Section 3: Table 8. Other Meiofaunal Groups Observed in Thames Sediment Samples, January - March 1990

Stn.	G R O U P							
	1	2	3	4	5	6	7	8
1	+		+				+	+
2			+				+	+
3	+		+					+
4		+	+				+	+
5			+					
6		+	+					+
7s		+						
7i								
8s								
8i								
9								
10s			+					+
10i			+					
11s			+					+
11i								
12s								
12i								
13s		+						+
13i		+						+
14		+	+					+
15		+	+					+
16		+	+					+
17			+					+
18	+	+	+					
19s			+					
19i			+					+
20		+				+		+
21	+				+	+	+	+
22	+		+	+			+	+
Stn.	1	2	3	4	5	6	7	8



Section 4

NON-MEIOFAUNAL GROUPS
PRESENT IN THAMES SEDIMENT SAMPLES
APRIL 1989 - MARCH 1990

Section 4: Table 1. Key to Table of Non-meiofaunal Groups observed in Thames Sediment Samples, April - June 1989

- 1 Filamentous algae
 - 2 Desmids
 - 3 Diatoms, mostly centric species, some pennate
 - 4 Ciliate Protozoa
 - 5 Thecate hydroids, detached hydranths and sections of perisarc
 - 6 Nemertea, juvenile
 - 7 Rotifera, loricae of *Keratella* species (e.g. *K. quadrata* and *K. cochlearis*) and contracted remains of *Bracionus* species
 - 8 Annelida: Oligochaeta, Tubificidae and Naididae
 - 9 Annelida: Polychaeta, spionid and nereid neochaetes
 - 10 Mollusca: Gastropoda, littorinid egg capsules
 - 11 Mollusca: Bivalvia, newly settled spat, ?tellinid species
 - 12 Crustacean nauplii
 - 13 Cladocera, mostly *Bosmina* species, some *Daphnia* species
 - 14 Amphipoda, eggs/embryoes of *Corophium* species
 - 15 Copepoda: Calanoida, *Eurytemora affinis*
 - 16 Hexapoda: Diptera: Nematocera: Chironomidae, larvae
-

Section 4: Table 2. Other Meiofaunal Groups Observed in Thames Sediment Samples, April - June 1989

Stn.	G R O U P															
	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
1	+		+				+	+	+			+	+		+	+
2	+		+				+	+	+				+	+		+
3			+				+	+	+							
4			+				+	+	+							
5		+	+				+	+								
6			+				+									
7	+		+													
8s			+					+	+							
8i			+		+		+	+	+							
9	+		+				+	+	+							
10s			+					+	+							
10i	+		+				+	+	+					+		
11s			+													
11i			+									+				
12s			+						+			+		+		
12i			+					+	+			+		+		
13s	+		+					+	+					+		
13i	+		+		+			+	+		+	+		+		
14			+						+			+				
15					+				+		+	+		+		
16			+						+	+	+	+				
17			+		+				+	+	+	+				
18			+	+	+				+			+				
19s		+									+					
19i		+		+	+			+	+	+	+					
20			+	+												
21			+					+			+	+			+	
22			+	+				+			+	+				

Section 4: Table 3. Key to Table of Non-meiofaunal Groups observed in Thames Sediment Samples, July - September 1989

- 1 Diatoms, mostly centric species, some pennate
 - 2 Filamentous algae
 - 3 Desmids
 - 4 Thecate hydroids, detached hydranths and sections of perisarc
 - 5 Rotifera, loricae of *Keratella* species (e.g. *K. quadrata* and *K. cochlearis*) and contracted remains of *Bracionus* species
 - 6 Bryozoa: sessobalsts, probably of freshwater species
 - 7 Annelida: Oligochaeta, Tubificidae and Naididae
 - 8 Annelida: Polychaeta, spionid and nereid neochaetes (? *Manayunkia* species also present in upper reaches of estuary)
 - 9 Mollusca: Bivalvia, newly settled spat, ?tellinid species
 - 10 Mollusca: Gastropoda, littorinid egg capsules
 - 11 Crustacean nauplii
 - 12 Cladocera, mostly *Alona affinis* but also 2 species of *Pleuroxis*
 - 13 Copepoda: Calanoida (*Eurytemora affinis*)
 - 14 Copepoda: Cyclopoida (unidentified species)
 - 15 Hexapoda: Diptera: Nematocera: Chironomidae, larvae
-

Section 4: Table 4. Other Meiofaunal Groups Observed in Thames Sediment Samples, July - September 1989

Stn.	G R O U P														
	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15
1	+	+			+	+	+				+			+	+
2	+	+	+		+	+	+				+	+			+
3					+	+	+								+
4	+				+		+								
5															
6	+						+	+							
7s	+						+								
7i	+						+								
8s	+			+											
8i	+										+				
9						+	+								
10s	+	+									+				
10i	+						+	+							
11s	+										+				
11i	+				+		+	+							
12s	+														
12i	+							+			+				
13s	+			+							+				
13i	+						+	+							
14	+			+			+	+							
15	+			+		+	+	+							
16	+				+		+	+	+	+	+				
17	+			+		+		+	+		+			+	
18	+			+					+						
19s	+						+	+	+						
19i	+							+		+					
20	+														
21	+							+		+	+				
22	+							+	+						

Stn. 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15

Section 4: Table 5. Key to Table of Non-meiofaunal Groups observed in Thames Sediment Samples, October - December 1989

- 1 Diatoms, mostly centric species, some pennate
 - 2 Filamentous algae
 - 3 Desmids
 - 4 Thecate hydroids, detached hydranths and sections of perisarc
 - 5 Rotifera, loricae of *Keratella* species (e.g. *K. quadrata* and *K. cochlearis*) and contracted remains of *Brachionus* species
 - 6 Bryozoa: detached zooids from colonies (possibly *Electra pilosa*)
 - 7 Annelida: Oligochaeta, Tubificidae and Naidiidae
 - 8 Annelida: Polychaeta, spionid and nereid neochaetes
 - 9 Mollusca: Bivalvia, newly settled spat, ?tellinid species
 - 10 Mollusca: Gastropoda, newly metamorphosed larvae
 - 11 Crustacean nauplii
 - 12 Cladocera, mostly *Alona affinis* but also 2 species of *Pleuroxis*
 - 13 Amphipoda, juveniles possibly *Corophium* species
 - 14 Hexapoda: Diptera: Nematocera: Chironomidae, larvae
-

Section 4: Table 6. Other Meiofaunal Groups Observed in Thames Sediment Samples, October - December 1989

Stn.	G R O U P													
	1	2	3	4	5	6	7	8	9	10	11	12	13	14
1		+	+		+		+				+	+	+	+
2	+	+	+		+		+				+			
3	+		+	+	+		+							+
4	+						+				+			+
5	+						+			+	+			
6	+						+				+			
7s	+						+			+				
7i	+			+	+		+			+	+			
8s	+													
8i	+				+		+			+				
9	+			+	+									
10s	+													
10i	+			+	+		+				+			
11s				+			+		+		+			
11i	+						+							
12s	+													
12i	+			+			+				+		+	
13s	+			+			+	+	+					
13i	+						+		+					
14	+			+				+		+				
15	+			+					+				+	
16	+						+		+					
17	+						+		+					
18	+			+		+	+	+	+	+	+			
19s	+			+				+	+					
19i	+							+	+		+			
20	+								+					
21	+							+		+				
22	+			+				+	+					

Section 4: Table 7. Key to Table of Non-meiofaunal Groups observed in Thames Sediment Samples, January - March 1990

- 1 Filamentous algae
 - 2 Diatoms, mostly centric species, some pennate
 - 3 Desmids
 - 4 Stalked, colonial Protozoa
 - 5 Thecate hydroids, detached hydranths and sections of perisarc
 - 6 Bryozoa: detached zooids from colonies (possibly *Electra pilosa*)
 - 7 Rotifera, loricae of *Keratella* species (e.g. *K. quadrata* and *K. cochlearis*) and contracted remains of *Brachionus* species
 - 8 Annelida: Oligochaeta, Tubificidae and Naididae
 - 9 Annelida: Polychaeta, spionid and nereid neochaetes
 - 10 Mollusca: Bivalvia, newly settled spat, ?tellinid species
 - 11 Mollusca: Gastropoda, newly metamorphosed larvae
 - 12 Crustacean nauplii
 - 13 Cladocera, mostly *Alona affinis* but also 2 species of *Pleuroxis*
 - 14 Amphipoda, juveniles possibly *Corophium* species
 - 15 Calanoid Copepoda; *Eurytemora affinis*
 - 16 Hexapoda: Diptera: Nematocera: Chironomidae, larvae
-

Section 4: Table 8. Other Meiofaunal Groups Observed in Thames Sediment Samples, January - March 1990

Stn.	G R O U P															
	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
1	+	+	+			+	+	+	+							
2	+		+				+	+	+			+	+			
3			+			+	+	+	+					+		+
4	+		+				+	+	+			+	+			+
5	+		+													
6	+		+			+	+					+				+
7s			+			+	+	+				+				
7i			+		+		+	+		+						+
8s			+					+								
8i			+					+								
9								+		+						
10s			+					+			+					
10i			+					+								
11s			+					+				+				
11i			+					+				+				
12s			+					+				+				
12i			+					+		+		+				
13s			+					+				+				
13i		+	+					+			+					
14			+					+	+			+				
15			+			+										
16			+					+	+		+	+				
17			+					+	+			+				
18			+		+	+		+	+					+		
19s			+		+			+	+							
19i			+					+				+				
20			+		+	+		+	+			+				
21				+					+							
22			+						+		+					

Stn.	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
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Section 5

NEMATODE FEEDING TYPES



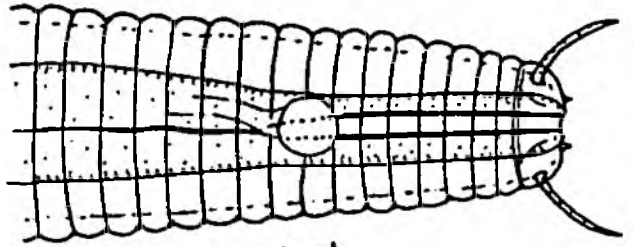
Nematode Feeding Types and the 1B:2A Ratio

Depending on the structure of the buccal capsule, nematodes can be classified into one of four different groups (see Figure 20). These groups appear to relate to their mode of feeding. Type 1 species lack cuticularised teeth whereas type 2 species have between 1 and 3 primary teeth that can be exceptionally large. Type 1 species are subdivided into 1A species which have small or narrow buccal cavities and 1B species that have large unarmed buccal cavities. The type 1A species are believed to be microbivorous or selective deposit feeders and, in the present survey, include species such as those belonging to the families Oxystominidae and the Leptolaimidae. Type 1B species are non-selective detritivores and, although they may ingest whole diatoms, they usually ingest 'plugs' of organically rich sediment. Examples found in the Thames Estuary include most of the xyloid nematodes and *Richtersia inaequalis*.

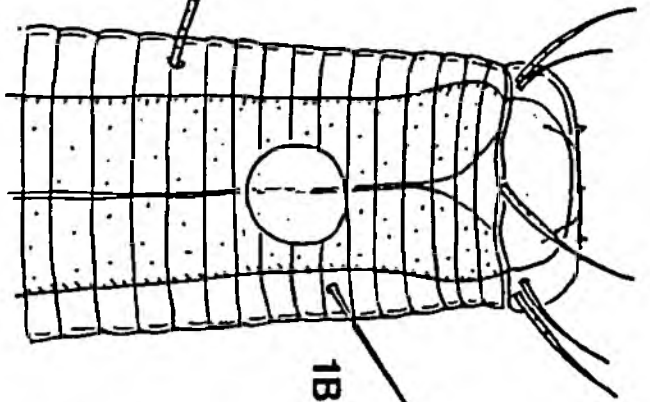
Type 2 species are also sub-divided on the basis of the size of their buccal cavities. 2A species have small cavities armed with teeth. These may be modified to split open frustules of sediment-dwelling diatoms or to rasp epigrowth from the surface of sand grains. Type 2A species attain their highest densities in coarse grained sediments and include most members of the Chromadoridae, Desmodoridae and Cyatholaimidae. Type 2B species have large armed buccal cavities, occasionally with moveable jaws and include predatory as well as omnivorous species. Oncholaimid species such as *Adoncholaimus thalassophygas* and enoplids such as *Enoplus brevis* belong to this group. These are often large nematode species with life-cycles of up to a year. Consequently, their populations are slow to re-establish following a catastrophic disturbance.

The ratio between 1B and 2A feeding types (1B:2A ratio) is a fundamental index used to describe the trophic composition of nematode populations. This has been used to detect changes in the composition of nematode assemblages with shifts to or from non-selective detritivore-dominated populations or selective epigrowth/diatomivorous species populations. The ratios for the stations at the mouth of the estuary and towards the western end of the Estuary are predominantly less than 5.00. Those for the muddier middle reach sediment communities are generally high. This reflects the low densities of selective epigrowth feeders combined with the elevated numbers of non-selective detritus feeding species in the middle reach. Used in combination with multivariate analyses of meiofaunal species complements, the 1B:2A nematode feeding type ratio is a powerful tool for the assessment of pollution status and in the detection of changes in prevailing environmental conditions in aquatic ecosystems.

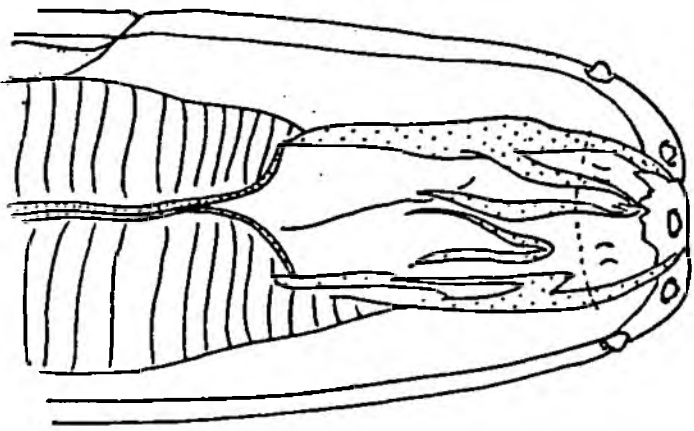
Figure 20. The buccal structures of the four principal nematode feeding types used in the investigation of trophic structures of nematode assemblages. See text for descriptions.



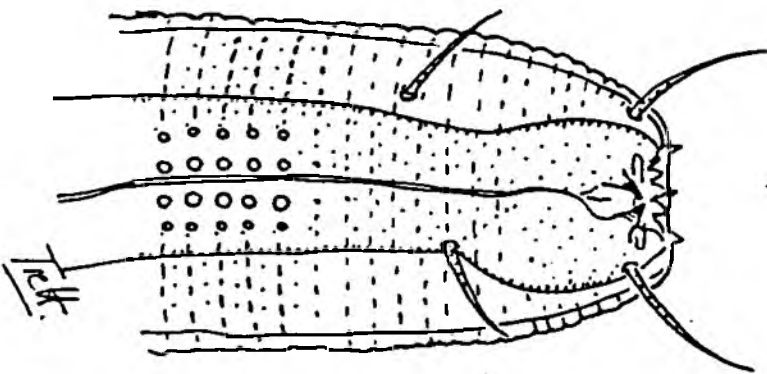
1A



1B



2B



2A



Appendix II

PARTICLE SIZE DISTRIBUTIONS

(October - December 1989)

Site Code: TW1

Total dry weight: 471.00g

Decription: Mud and stones.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	1.77
63 μm - 212 μm	5.99
212 μm - 600 μm	10.37
600 μm - 1 mm	2.52
1 mm - 2 mm	3.21
> 2 mm	76.14

Site Code: TW2

Total dry weight: 254.25g

Decription: Fine sand and gravel

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	1.77
63 μm - 212 μm	5.99
212 μm - 600 μm	10.37
600 μm - 1 mm	2.52
1 mm - 2 mm	3.21
> 2 mm	76.14

Site Code: TW3

Total dry weight: 430.20g

Decription: Sand with some organic matter.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	7.35
63 μm - 212 μm	16.45
212 μm - 600 μm	74.74
600 μm - 1 mm	0.66
1 mm - 2 mm	0.32
> 2 mm	0.48

Site Code: TW4

Total dry weight: 204.54

Decription: Fine sand and stones.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	25.59
63 μm - 212 μm	11.21
212 μm - 600 μm	47.35
600 μm - 1 mm	2.21
1 mm - 2 mm	0.39
> 2 mm	13.25

Site Code: TW5

Total dry weight: 312.50g

Decription: Sand.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	2.27
63 μm - 212 μm	9.89
212 μm - 600 μm	85.20
600 μm - 1 mm	1.80
1 mm - 2 mm	0.61
> 2 mm	0.24

Site Code: TW6

Total dry weight: 172.57g

Decription: Silty mud with some stones.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	24.78
63 μm - 212 μm	20.26
212 μm - 600 μm	22.59
600 μm - 1 mm	2.38
1 mm - 2 mm	5.83
> 2 mm	24.15

Site Code: TW7s

Total dry weight: 271.54g

Decription: Sand.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	9.14
63 μm - 212 μm	32.62
212 μm - 600 μm	51.75
600 μm - 1 mm	2.63
1 mm - 2 mm	1.84
> 2 mm	2.02

Site Code: TW7i

Total dry weight: 214.64g

Decription: Silt and mud.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	22.54
63 μm - 212 μm	52.34
212 μm - 600 μm	22.05
600 μm - 1 mm	1.05
1 mm - 2 mm	2.02
> 2 mm	0.00

Site Code: TW8s

Total dry weight: 155.28g

Decription: Silt and mud.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	35.06
63 μm - 212 μm	54.32
212 μm - 600 μm	10.35
600 μm - 1 mm	0.15
1 mm - 2 mm	0.12
> 2 mm	0.00

Site Code: TW8i

Total dry weight: 301.22g

Decription: Mud and stones.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	13.82
63 μm - 212 μm	14.75
212 μm - 600 μm	12.62
600 μm - 1 mm	1.61
1 mm - 2 mm	1.97
> 2 mm	55.23

Site Code: TW9

Total dry weight: 177.58g

Decription: Mud.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	50.65
63 μm - 212 μm	39.61
212 μm - 600 μm	9.24
600 μm - 1 mm	0.09
1 mm - 2 mm	0.20
> 2 mm	0.21

Site Code: TW10s

Total dry weight: 205.36g

Decription: Fine mud.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	40.03
63 μm - 212 μm	50.76
212 μm - 600 μm	8.46
600 μm - 1 mm	0.11
1 mm - 2 mm	0.20
> 2 mm	0.42

Site Code: TW10i

Total dry weight: 175.08g

Decription: Fine mud.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	34.76
63 μm - 212 μm	48.32
212 μm - 600 μm	16.25
600 μm - 1 mm	0.25
1 mm - 2 mm	0.42
> 2 mm	0.00

Site Code: TW11s

Total dry weight: 145.04g

Decription: Fine mud.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	41.19
63 μm - 212 μm	36.54
212 μm - 600 μm	22.12
600 μm - 1 mm	0.15
1 mm - 2 mm	0.00
> 2 mm	0.00

Site Code: TW11i

Total dry weight: 114.35g

Decription: Fine mud.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	67.41
63 μm - 212 μm	16.35
212 μm - 600 μm	16.13
600 μm - 1 mm	0.02
1 mm - 2 mm	0.00
> 2 mm	0.07

Site Code: TW12s

Total dry weight: 180.10g

Decription: Coarse mud.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	36.10
63 μm - 212 μm	44.99
212 μm - 600 μm	12.07
600 μm - 1 mm	0.27
1 mm - 2 mm	2.03
> 2 mm	4.54

Site Code: TW12i

Total dry weight: 167.29g

Decription: Coarse mud.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	44.47
63 μm - 212 μm	45.03
212 μm - 600 μm	9.21
600 μm - 1 mm	0.30
1 mm - 2 mm	0.45
> 2 mm	0.51

Site Code: TW13s

Total dry weight: 296.07g

Decription: Coarse mud.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	38.36
63 μm - 212 μm	52.54
212 μm - 600 μm	8.57
600 μm - 1 mm	0.53
1 mm - 2 mm	0.00
> 2 mm	0.00

Site Code: TW13i

Total dry weight: 434.55g

Decription: Mud.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	22.54
63 μm - 212 μm	51.21
212 μm - 600 μm	26.19
600 μm - 1 mm	0.06
1 mm - 2 mm	0.00
> 2 mm	0.00

Site Code: TW14

Total dry weight: 265.10g

Decription: Mud.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	29.49
63 μm - 212 μm	46.50
212 μm - 600 μm	23.56
600 μm - 1 mm	0.21
1 mm - 2 mm	0.23
> 2 mm	0.00

Site Code: TW15

Total dry weight: 316.54g

Decription: Mud and shell (bivalve).

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	23.80
63 μm - 212 μm	65.26
212 μm - 600 μm	8.72
600 μm - 1 mm	0.45
1 mm - 2 mm	0.60
> 2 mm	1.17

Site Code: TW16

Total dry weight: 365.62g

Decription: Mud.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	35.85
63 μm - 212 μm	54.58
212 μm - 600 μm	9.54
600 μm - 1 mm	0.01
1 mm - 2 mm	0.01
> 2 mm	0.02

Site Code: TW17

Total dry weight: 295.46g

Decription: Mud and shells (bivalve).

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	37.83
63 μm - 212 μm	35.15
212 μm - 600 μm	20.51
600 μm - 1 mm	0.57
1 mm - 2 mm	0.94
> 2 mm	5.01

Site Code: TW18

Total dry weight: 186.81g

Decription: Mud with shells (bivalve) and bored wood.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	20.70
63 μm - 212 μm	18.96
212 μm - 600 μm	13.63
600 μm - 1 mm	2.08
1 mm - 2 mm	2.63
> 2 mm	42.01

Site Code: TW19s

Total dry weight: 289.73g

Decription: Sand with stones and shells (bivalve).

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	5.83
63 μm - 212 μm	29.42
212 μm - 600 μm	52.34
600 μm - 1 mm	0.39
1 mm - 2 mm	0.48
> 2 mm	11.54

Site Code: TW19i

Total dry weight: 319.16g

Decription: Fine sand and shells (bivalve).

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	14.44
63 μm - 212 μm	65.82
212 μm - 600 μm	12.92
600 μm - 1 mm	0.75
1 mm - 2 mm	0.51
> 2 mm	5.53

Site Code: TW20

Total dry weight: 241.02g

Decription: Fine sand.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	20.79
63 μm - 212 μm	47.03
212 μm - 600 μm	32.15
600 μm - 1 mm	0.02
1 mm - 2 mm	0.02
> 2 mm	0.00

Site Code: TW21

Total dry weight: 564.25g

Decription: Fine sand and shells (bivalve).

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	7.81
63 μm - 212 μm	31.48
212 μm - 600 μm	57.06
600 μm - 1 mm	0.56
1 mm - 2 mm	0.51
> 2 mm	2.58

Site Code: TW22

Total dry weight: 245.90g

Decription: Fine sand.

<u>Particle Size</u>	<u>Percentage</u>
< 63 μm	5.71
63 μm - 212 μm	50.50
212 μm - 600 μm	43.28
600 μm - 1 mm	0.29
1 mm - 2 mm	0.07
> 2 mm	0.15
